

HPLC METHOD DEVELOPMENT, VALIDATION AND ITS APPLICATION TO STABILITY STUDIES OF CHLORPROMAZINE HYDROCHLORIDE TABLETS

Venkatesh S^{1*}, Mandal Badal Kumar¹, Sridevi Ramachandran^{1,2} and Navalgund Sameer G.²

¹Environmental and Analytical Chemistry Division, School of Advanced Sciences, VIT University, Vellore-632014, India

²International Specialty Chemicals (India) Pvt Ltd, Hyderabad, A.P., India

*Venkatesh S, Environmental and Analytical Chemistry Division, School of Advanced Sciences, VIT University, Vellore, India Email: badalmandal@vit.ac.in

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ABSTRACT

Various formulations of chlorpromazine (CPZ) are coming out in the market in combination with different excipients. Although a few high-performance liquid chromatography (HPLC) methods were in action, they were not able to separate in a short time with low cost and easily available mobile phases without affecting the drug components. The main objective of the present study was to develop a new simple HPLC method for identification and quantification of CPZ and its degradation products in tablets and formulations. The advantage of the developed HPLC method was that it was simple isocratic method without using any complicated mobile phase. It was highly sensitive and would be applied to the oxidative degradation studies of CPZ.HCl for monitoring the quality of CPZ.HCl during storage. It could be applicable to different derivatives and metabolites of CPZ and its formulations as well as to other formulations with different new excipients.

KEYWORDS: Chlorpromazine tablet, HPLC, stability studies, degradation, formulation

INTRODUCTION

Chlorpromazine (CPZ), a propylamino phenothiazine compound, acts as an antagonist of dopamine-2 receptor¹. It is sold as a typical antipsychotic drug under the trade names Largactil® and Thorazine®. Chlorpromazine HCl (CPZ.HCl) has an aliphatic side chain, typical for low to middle potency neuroleptics². Currently, spectrophotometric/titrimetric methods are used for assay of CPZ in various formulations, while the purity is checked by thin-layer chromatography³. Nowadays liquid chromatography has been developed a lot and researchers are applying it for purification, identification and quantification using octadecyl³, octyl⁴⁻⁵ and C≡N phases⁶. Figure 1 shows the chemical structure of chlorpromazine hydrochloride.

The degradation of CPZ.HCl with single active pharmaceutical ingredient (API) and its metabolites in alkaline mobile media restricts uses of alkaline mobile phases for separation studies^{1,7}. The present study intended to develop a suitable method which would be able to monitor the oxidative degradation products of CPZ.HCl along with different super disintegrants.

There were many HPLC methods available for the determination of CPZ in the ppb range in human blood plasma⁸⁻¹². Chlorpromazine HCl was studied with different analytical instrumental methods in combination drugs/formulations and oral doses with preservatives or without preservatives. Different types of excipients/placeboes were also used during the preparation of tablets⁸⁻¹². But in the present study a new excipient was used for the preparation of tablets. So majority of the published HPLC methods were not able to fit our study of oxidative degradation products of CPZ with a new excipient (super disintegrant) in tablet dosage form.

Chlorpromazine, a basic compound, typically shows poor chromatographic behavior resulting in broad, tailing peaks when analyzed by reversed phase HPLC (RP-HPLC). Various mobile phase

modifiers, such as short-chained amines, decylamine, triethylamine and ion-pair reagents have been used to obtain acceptable chromatograms¹³⁻¹⁵. The formulation got degraded with storage into some degradants and other components. Methylparaben and propylparaben have been used as preservatives in CPZ.HCl in a liquid oral pharmaceutical formulation. Although ion-pair agents were added to mobile phase to improve the peak shape of CPZ during assay of the active component and preservatives in CPZ.HCl oral solution using HPLC, retention time of CPZ was increased substantially¹⁵. Alkaline solutions would cause the drug to oxidize. Keeping in mind all these difficulties the following objectives were taken to fulfill the goal of the present study.

The objectives of the present study were i) to develop a new simple, specific, precise and accurate HPLC method, ii) to validation of the developed method as per ICH guidelines, iii) to carry degradation studies of CPZ.HCl, iv) to study the effects of oxidative degradation processes of excipients (used here as super disintegrants) on CPZ.HCl and v) to assess its suitability for formulation.

MATERIALS AND METHODS

Chemicals

USP reference standards acdisol (FMC polymer), polyplasdone XL (ISP), chlorpromazine HCl (Emco laboratory) were used for validation. HPLC grade (Qualigens) mobile phases, Milli-Q water and all other reagents of analytical grade were used. The buffer was prepared by dissolving 2.72g of potassium dihydrogenorthophosphate and 5ml triethylamine (TEA) in 1 liter and pH was adjusted to 3.1 with ortho phosphoric acid. The mobile phase consisted of buffer and acetonitrile in the ratio of 65:35. Amber glassware was used for all samples and standard preparations.

Instrumentations

A HPLC system (Model 2695, WATERS, Milliford - USA) consisting of a pneumatic pump, a degasser, column heater, an auto injector and a photo diode array detector 2996 was controlled by computer and empower software. The column Zorbax extended C₁₈ column (250 x 4.6 mm, 5 μ m, 10 A° (Agilents, Santa Clara, USA) was used. The stability study of chlorpromazine hydrochloride was undertaken to test its stability with super disintegrants under three different conditions 25°C/60% RH, 30°C/65% RH and 40°C/75% RH stability chamber. Hardness, thickness, diameter were checked by using Inkarph Pharma tester. Disintegration and friability were tested by using Erweka manufacturer apparatus.

Analytical methods

HPLC Method validation was carried out as per ICH protocol¹⁶. The terms selectivity and specificity are often used interchangeably. The selectivity of an analytical method was tested by comparing results of samples containing impurities with the results of samples without impurities. Precision of the developed method was the degree of agreement among individual test results when the procedure was applied repeatedly to multiple samplings. It was measured by injecting a series of standards. Accuracy is the measure of how close the experimental value is to the true value. The accuracy of the present method was the degree of agreement of test results generated by the method to the true value. A linear regression equation applied to the results should have an intercept not significantly different from zero¹⁷. The range was expressed in the same units as test results (for example percent, parts per million) obtained by the analytical method. The limit of detection is the point at which measured value is larger than the uncertainty associated with it is the lowest concentration of the analyte in a sample that can be detected, but not necessarily quantified. In chromatography the detection limit was the injected amount which resulted in a peak with a height at least twice as high as the baseline noise. The limit of quantification was the injected amount which resulted in a reproducible measurement of peak areas (equivalent to amounts). Peak heights were typically required to be at least 10 times higher than baseline noise. A ruggedness test was examined to assess the affect operational and environmental conditions on the analysis results. It was the degree of variance in test results obtained by the analysis of the same samples under a variety of different test conditions. As per ICH definition¹⁶⁻¹⁷ robustness being a measure of the method's capability was unaffected by small, but deliberate variations in method parameters. Robustness could be partly assured by good system suitability specifications. Thus, it was important to set tight, but realistic, system suitability specifications. Testing variation in some or all

conditions, e.g., age of columns, column type, column temperature, and pH of buffer in mobile phase, reagents were performed to validate the method¹⁶. Theoretical plate number (N) was calculated for the method as discussed elsewhere¹⁸.

Oxidative degradation study of CPZ.HCl

The ingredients (CPZ.HCl, lactose monohydrate, microcrystalline cellulose, polyplasdone XL or Acdisol, aerosol, Mg-stearate) of CPZ.HCl tablets were mixed in dry condition and compressed in such a manner to get a tolling size of 7mm. The study of CPZ.HCl was undertaken to test its stability with super disintegrants under three different conditions [Ambient, 30°C-65% and 40°C-75% Relative Humidity (RH)]. As super disintegrants contained peroxide, the present study was also undertaken to ensure whether the peroxide had any effect on the stability of CPZ.HCl or not. Analysis of these tablets was performed for duration of 1 to 180 days using HPLC method developed in the present study.

Physical parameters such as color (visual/hunter), weight (wt. gain/wt. uniformity), friability test, hardness test, disintegration test, diameter/thickness and chemical parameters such as assay (Impurity/content uniformity), moisture content and dissolution of tablets were carried out repeatedly on days 0, 30, 60, 90, and 180 respectively.

Statistical analysis

All the statistical parameters present in the results were calculated as per Microsoft Excel-Windows 2003.

RESULTS

Method development

All parameters mentioned in Table 1 were optimized to get good resolution of the separated compounds in the present study.

Method validation

Diameter, thickness and hardness of the tablets were checked by using Inkarph Pharma Tester and all agreed the pre-specifications. Hardness was also verified through friability test using ERWEKA Friabilator (Ahemadabad, India) and was within control limit. Time of disintegration was checked to be 61-103 sec for innovator sample and 61-89 sec for ISP sample which were mostly identical (data not shown). The above results of stability study indicated that oxidative degradation of CPZ.HCl in the tablet formulations did not occur under the following test conditions i.e., ambient condition, 30°C and 65 % RH, and with 40°C and 75 % RH up to three months.

Method validation is generally a one-time process performed after the method development to demonstrate that the method was scientifically sound and that it served the intended analytical purpose¹⁸. The test method was validated for specificity, linearity, precision, accuracy, range, ruggedness and robustness. The specificity is the ability to assess unequivocally the analyte of interest in the presence of component that may be expected to be present, such as impurities, degradation products, and matrix components stored under relevant stress conditions (e.g., light, heat, acid hydrolysis, alkali hydrolysis, and H₂O₂ oxidation). The recovery ranges of CPZ were 104.2-106.9% under acid hydrolysis, 23.3-96.4% under 1-10% H₂O₂ oxidation, 2.02-98.1% during photolysis for a period of 1 min to 3 hr, 99.4-23.3% under heating for 1-2hr and 75.2-111.6% under alkali hydrolysis for a period of 15 min to 1 hr respectively.

As per ICH acceptance criteria RSD (%) of precision study should not be more than 2.0 %. For precision study all solutions were prepared in triplicate at levels 80%, 100% and 120% of test concentrations using CPZ.HCl working standard as per the test method. In the present study RSD% was found to be 0.21%, whereas method precision and sample precision were 1.7% and 0.2% (n=6) using both chlorpromazine ISP and chlorpromazine innovator. As per ICH acceptance criteria recovery should be within 95 to 105%. In the present study the observed acceptable level of accuracy was 98.7 to 101.5% of test concentration for the assay of CPZ.HCl (Table 2).

For linearity study a series of solutions were prepared using CPZ.HCl working standard at concentration levels from 1ppm to 200 ppm. The value of R² was 0.99 and RSD% was 0.10-1.11%. The range should be reported in % with respect to test concentration from the linearity, precision and accuracy

data. The range of the analytical method for the assay of CPZ.HCl was 92% to 107% of test concentration.

For ruggedness study six samples individually were prepared using single batch of CPZ.HCl and each solution was injected in triplicate using different column, system, analyst on different days. RSD% was found to be 1.5%. Robustness was calculated using the developed HPLC method for the effect of temperature, buffer concentration i.e., pH and time on recovery and it was found to be 90.3-101.2%. In addition, limit of detection (LOD) was calculated based upon a signal-to-noise ratio of 3 (as calculated in the external standard calibration solution and adjusting for the dilution factor of sample preparation). Samples were injected 6 times and then LOD was calculated. The limit of quantification (LOQ) was the injected amount which resulted in a reproducible measurement of peak areas (equivalent to amounts) or peak heights typically required to be at least 10 times higher than baseline noise. System suitability study showed that for the developed HPLC method RSD (%), USP tailing and USP plate count were 0.39%, 1.22 and 5310 respectively which were within USP acceptance limit for CPZ.HCl.

The results supported the predetermined acceptance criteria. The validated method was found to be specific, linear, precise, accurate, robust and rugged for the assay of CPZ.HCl tablets in the present study (Table 2).

Dissolution study

There were many literatures available on the method development for CPZ using different columns and mobile phases^{1,12}. In the present study the column used was C₁₈ column (Waters, Milliford, USA) and the mobile phase taken for the first trial was methanol and water system. The peak was asymmetric but tailing was high (Fig. 2) in methanol, because CPZ.HCl being highly soluble in methanol was eluted at RT of 8 min and degraded quickly in slightly alkaline pH. Methanol-buffer system was also tried, but it was not satisfactory (Fig. 3). Later on methanol was replaced with acetonitrile and the phosphate buffer pH was adjusted to acidic at pH 3.1. The proportion of acetonitrile to buffer and pH were optimized and the symmetric peak with minimal retention time of 2.35 min was obtained (Fig. 4). Finally, the method was optimized as per ICH guidelines¹⁷ and validated (Table 2). Finally, the present method was applied to check impurity profile and degradation products of CPZ.HCl under different conditions for various lots at different time intervals.

Results on stability study

Reference Std. compound of CPZ.HCl (100 mg/L) was used to create an external standard calibration for the assay of CPZ.HCl stability study. The retention time of CPZ.HCl was 2.35 minutes (Fig. 4) using buffer-acetonitrile system. Stability study was carried out under normal condition, 25°C/60%RH, 30°C/65%RH and 40°C/75%RH for a period of three months. It was found that tablets made with polyplasdnone XL (ISP excipient) had recovery ranges of 98.5-104.1% while tablets made with acdisol (Innovator excipient) had recovery ranges of 97.2-103.9% (Table 2). Similar type of results was obtained from content uniformity study using 10 tablets. Dissolution study was carried out in 0.1N HCl using paddle apparatus at 50 rpm. Results were within USP specifications and same as that of stability study.

DISCUSSION

The aim of the present study was to develop a method for identification and quantification of CPZ.HCl and its validation as well as application to stability study. The present HPLC method was developed with the trials of different mobile phases such as methanol and water system, methanol and phosphate buffer system or acetonitrile and phosphate buffer system respectively. The acetonitrile buffer system produced the optimized separation capacity (Fig. 4) using Zorbax C₁₈ column. The developed method was validated as per ICH guidelines¹⁷⁻¹⁸ (Table 2).

The validation parameters such as specificity, precision (% RSD), linearity (R² as 0.9998), accuracy (%RSD), ruggedness and robustness, system suitability results met the requirements (Table 2) and fulfilled the ICH guidelines for CPZ. HCl^{16-17,19}.

The optimized developed HPLC method was fast, accurate, precise and reproducible. The validation parameters tallied nicely with ICH guidelines. The linearity study showed a R² of 0.9998 and

system suitability results (in % RSD) were less than 2. The specificity, ruggedness and robustness were comparable to ICH guidelines.

Although Kollmorgen et al.¹³ used mobile phase methanol-acetate buffer system for the separation of CPZ.HCl and methyl paraben, this mobile system did not work in the present study. Also, the HPLC method described by Boehme and Strobel¹ was tried, but it was not suitable for the present study. The developed and validated method was applied for the stability study of CPZ.HCl with super disintegrants. According to ICH guidelines for stability study¹⁹ the optimized method was performed and results were strictly within the pre-specified limits.

The stability results of physical and chemical parameters such as assay, content uniformity, dissolution, friability, disintegration time, diameter, thickness and hardness were within the intended limits. The results of comparative study between innovative and ISP products showed that the drug formulated in ISP was stable and they did not degrade with super disintegrants.

The assay test of tablets showed comparable values with ICH within the range of 90 to 105% for all the tablet formulations. In addition, there was no peak appeared in the samples collected from the oxidative degradation study of the tablets (Fig. 5). So, the tablets were free from oxidative degradation during the period of observation time.

CONCLUSIONS

The HPLC method developed in the present study is simple, reliable, sensitive, reproducible, less time consuming and is applicable to test CPZ.HCl in bulk drug, raw materials and tablet doses. The advantage of the present test procedures is that it is a simple isocratic method without using any complicated mobile phase. The present work gets validated, highly sensitive and selective method for determination of CPZ.HCl in pharmaceutical dosage forms. Also, it would be applied to the oxidative degradation studies of CPZ.HCl for monitoring the quality of CPZ.HCl during storage. The oxidative degradation study of CPZ.HCl confirms no oxidative degradation of CPZ.HCl in tablet formulations under ambient conditions as well as under accelerated stability conditions. The method could be applicable for chlorpromazine derivatives and their metabolites. This technique would be applicable to other formulations with different new excipients.

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REFERENCES

1. Boehme CL, Strobel HW. High performance liquid chromatographic methods for the analysis of haloperidol and chlorpromazine metabolism in vitro by purified cytochrome P450 isoforms. *J Chromatogr B* 1998; 718:259-266.
2. Indian pharmacopoeia 1996, 1:178.
3. USP XXIII, The United States Pharmacopeia, Rand McNally, Taunton, MA, 1995; 353-356.
4. Tamai G, Yoshida H, Imai H. High-performance liquid chromatographic drug analysis by direct injection of whole blood samples. I. Determination of moderately hydrophobic drugs incorporated into blood corpuscles. *J Chromatogr* 1987; 423:163-168.
5. Kiel JS, Morgan SL, Abramson RK. Computer-assisted optimization of a high-performance liquid chromatographic separation for chlorpromazine and thirteen metabolites. *J Chromatogr* 1989; 485:589-596.
6. Ohkubo Z, Shimoyama R, Sugawa K. Determination of chlorpromazine in human breast milk and serum by high-performance liquid chromatography. *J Chromatogr* 1993; 614:328-332.
7. Kountourellis JE, Makoupoulou CK. A simultaneous analysis by high performance liquid chromatography of bupropion combined with tricyclic antidepressants and/or antipsychotics in dosage forms. *J Liquid Chromatogr* 1991; 14:2969-2977.

8. Fekete J, Del Castilho P, Kraak JC. Reversed-phase liquid chromatography for the separation of chlorpromazine, imipramine and some of their metabolites. *J Chromatogr A* 1981; 204:319-327.
9. Cooper JK, McKay G, Midha KK. Subnanogram quantitation of chlorpromazine in plasma by HPLC with electrochemical detection. *J Pharm Sci* 2006; 72: 1259-1262.
10. Pistos C, Stewart JT. Direct injection HPLC method for the determination of selected phenothiazines in plasma using a Hisep column. *Analyst* 1985; 110:1117-1124.
11. Mahadik KR, Aggarwal H, Kaul N. Development and validation of HPLC method for simultaneous estimation of trihexyphenidyl hydrochloride and chlorpromazine hydrochloride from tablet dosage form. *Indian Drugs*, 39, 441-445, 2002.
12. Kircherer H, Khun-Veltan WN. Quantitative determination of forty- eight antidepressants and antipsychotics in human serum by hplc tandem mass spectrometry A multi level single – sample approach. *J Chromatogr B* 2006; 843:100-113.
13. Kollmorgen D, Kraut B. Determination of methylparaben, propylparaben and chlorpromazine in chlorpromazine hydrochloride oral solution by high-performance liquid chromatography. *J Chromatogr B* 1998; 707:181-187.
14. Snyder LR, Glajch JL, Kirkland JJ. *Practical HPLC Method Development*, Wiley, New York, 232, 1988.
15. Method # 0742.06, Alparma Inc., USPDI, (1994).
16. ICH Topic Q2B, "Validation of Analytical procedures": Methodology, 6th Nov 1996.
17. ICH Topic Q2A, "Validation of Analytical Procedures": Text, 6th Nov 1996.
18. USP 26, General Information, Validation of compendial methods 2003; 2440-2442.
19. ICH Stability Testing: Photostability Testing of New Drug Substances and Products, ICH, Geneva, Switzerland, November 1996.

Table 1: Optimized HPLC conditions for the elution of CPX.HCL

Parameters	Details
Stationary phase (column)	Zorbax Extend – C ₁₈ , 4.6×250mm, 5µm, 10 A°
Mobile Phase	0.02M KH ₂ PO ₄ (pH~3.1) : Acetonitrile (65:35 v/v), Isocratic
Flow rate	1 mL/min
Run time	20 minutes
Column temperature	35°C
Volume of injector loop	10 µl
Wavelength (λ _{max})	256 nm
Tailing factor	Not more than 2.0
Retention time	2.3 mins

Table 2: Comparison between observed results obtained by validated method and ICH guidelines

S.No	Validation parameters	ICH guidelines	Observed results
1	Method precision	% RSD not more than 2.0%	1.7%
2	System precision	% RSD not more than 2.0%	0.21%
3	Accuracy	Recovery should be within 95% to 105%	98.7% - 101.5%
4	Linearity	R ² (correlation coefficient) > 0.999	0.9998
5	Range	It should be within 80% to 120%	Complies
6	Ruggedness	% RSD not more than 2.0%	1.5%
7	System suitability	% RSD not more than 2.0%	0.39%
8	LOD & LOQ	Signal to noise ratio (s/n): 3:1 & 10:1	Complies

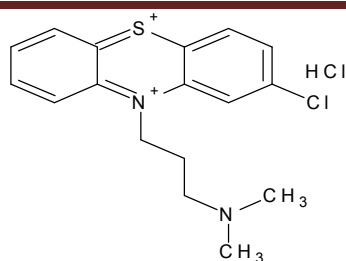


Figure 1: Structure of Chlorpromazine Hydrochloride

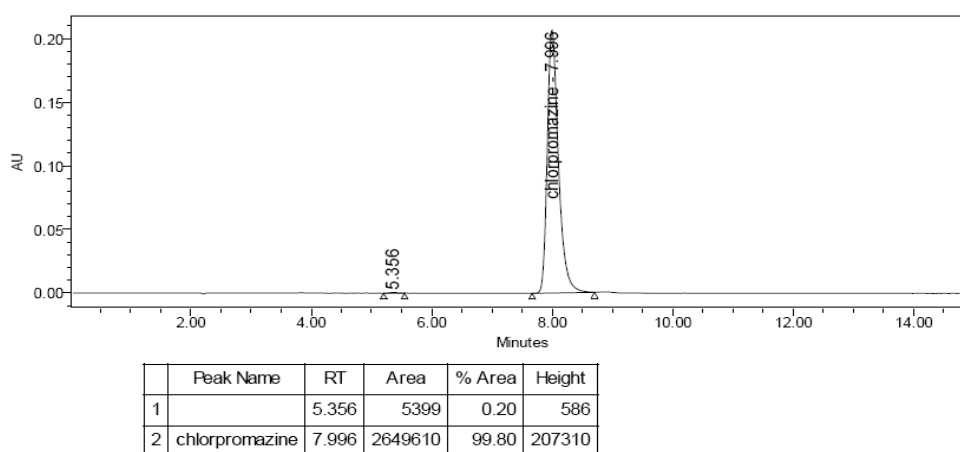


Figure 2: HPLC chromatogram of Chlorpromazine HCl. The concentration of the sample was 100mg/L. The mobile phase of methanol:water (85:15 v/v) was used at a flow rate of 1ml/minute for elution. The injection volume was 10 μ L and the column temp was 35°C.

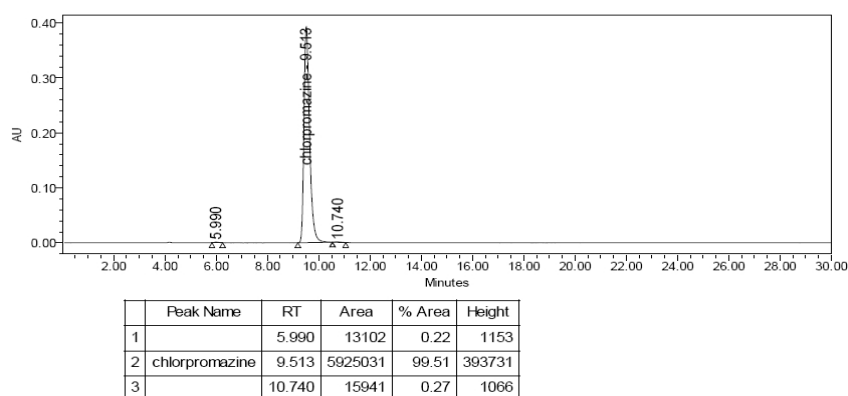


Figure 3: HPLC chromatogram of chlorpromazine HCl. The concentration of the sample was 100mg/L. The mobile phase of 0.02 M KH_2PO_4 and methanol containing 0.5% TEA (buffer:acetonitrile :: 20:80, v/v, pH 7.6) was used at a flow rate of 1ml/minute. The runtime was 15 min and the injection volume was 10 μ L and the column temp was 35°C.

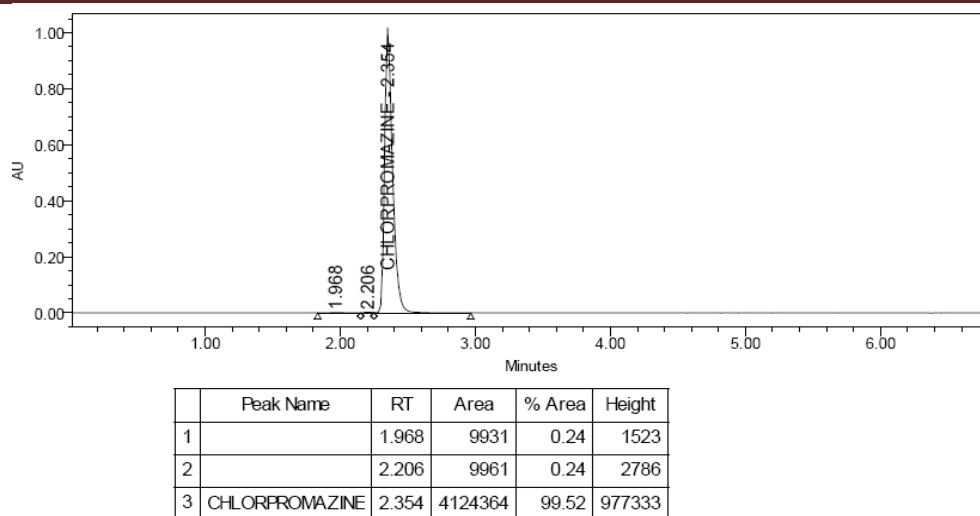


Figure 4: HPLC chromatogram of chlorpromazine HCl. The concentration of the sample was 100mg/L. The mobile phase of 0.02 M KH_2PO_4 and acetonitrile containing 0.5% TEA (buffer:acetonitrile :: 35:65, v/v, pH 3.1) was used at a flow rate of 1ml/minute. The runtime was 15 min and the injection volume was 10 μL and the column temp was 35°C.

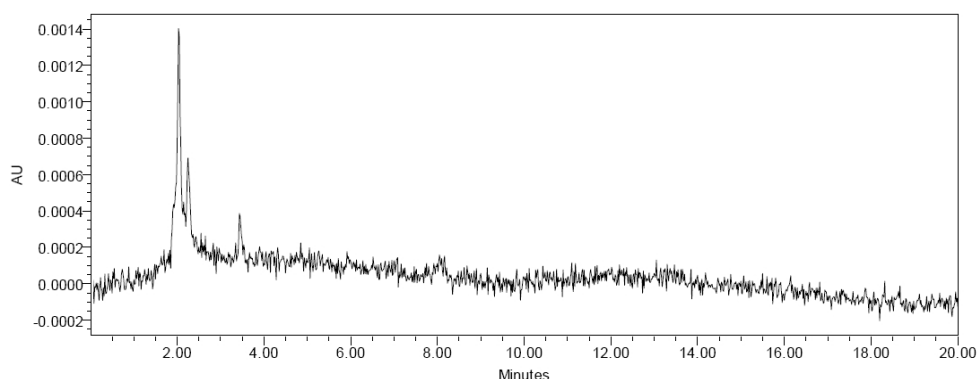


Figure 5: HPLC chromatogram of CPZ.HCl with placebo polyplasdone after oxidation degradation. The concentration of the sample was 100mg/L. The mobile phase of 0.02 M KH_2PO_4 and acetonitrile containing 0.5% TEA (buffer:acetonitrile :: 35:65, v/v, pH 3.1) was used at a flow rate of 1ml/minute. The runtime was 20 min and the injection volume was 10 μL and the column temp was 35°C.

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