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HEPATOPROTECTIVE ACTIVITY OF BAUHINIA RACEMOSA LINN

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ABSTRACT

Present work was undertaken to evaluate hepatoprotective activity of *Bauhinia racemosa* stem bark. The biochemical parameters such as SGOT, SGPT, Total bilirubin, ALP were estimated, the histopathological paraffin sections of rat liver were performed, analyzed to assess the hepatoprotective effect. The test material was found effective as hepatoprotective, through *in vivo* and histopathological studies. The hepatoprotective effect of ethyl acetate extract was comparable to that of silymarin, a standard hepatoprotective agent. **KEY WORDS:** *Bauhinia racemosa*, histopathological studies, silymarin

INTRODUCTION

The plant *Bauhinia racemosa Linn* (Fabaceae) is the most widely cultivated variety of the genus *Bauhinia* and is distributed in the sub-Himalayan ranges of India, Sri-Lanka, Mexico, Arabia and South Western Africa. The use of stem bark of *Bauhinia racemosa* has been reported in all kinds of pain, good vesicant, expectorant, stimulant and abortifacient. The decoction of the root bark is used as a stimulant, analgesic and diuretic. The stem bark is used in the treatment of jaundice¹. The pods are edible, seeds are useful as purgative, antipyretic, cures eye diseases, head complaints and are used in venereal affections².

A investigated flavones glycoside has been investigated from the stem bark of *Bauhinia racemosa*³. The present study was undertaken to screen hepatoprotective activity of the stem bark of *Bauhinia racemosa*.

MATERIALS AND METHODS

Collection of Plant material

Bauhinia racemosa Linn stem bark were collected from Mangalore D.K.District, Karnataka, India. The plant was identified and authenticated by Dr. Gopalkrishna Bhat, Department of Botany, Poornaprajna College, Udupi. A voucher specimen (NGSM 049) was deposited in Pharmacognosy Department of N.G.S.M. Institute of Pharmaceutical Sciences, Mangalore.

Preparation of extract

The stem bark was collected locally it was air dried under shade, powdered mechanically and stored in airtight containers. About3 kg of the powdered material was boiled with 10 liter of distilled water for 30 min and filtered to obtain the aqueous extract. The extract was concentrated under reduced pressure and finally dried using the water bath. Another 1 kg of the powdered material was subjected for soxhlation. It was refluxed with ethanol for 48 h in batches of 250 g each. The extracts were pooled together and concentrated in vacuum using rotary flash evaporator (Buchi, Flawil, Switzerland).

Animals

Male Wistar albino rats weighing 150-200 g were obtained from animal house located at K.S.Hegde Medical academy Deralakatte, Mangalore and Karnataka. They were maintained

at standard housing conditions and fed with commercial diet (Hindustan Lever Ltd., Bangalore) and provided with water *ad libitum* during the experiment.

Evaluation of hepatoprotective activity

The acute toxicity studies were carried out as per staircase method⁴. Fifty male rats were divided into five groups of 10 each and were administered with aliquot doses of the extracts orally (200, 400, 800, 100 and 2000 mg/kg). Mortality was not noticed up to 2000 mg/kg. The LD50 of the extracts was found to be 2000 mg/kg body weight. One-tenth of this dose was selected as the therapeutic dose for the evaluation⁵. Five groups of animals containing six each were used for the study. The animals from Group I served as the control and received the vehicle 1% w/v gum tragacanth at a dose of 1 ml/kg/day of p.o. for 14 days. Groups II–IV received 0.1

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ml/kg/ day i.p. of CCl4 (E-Merck, Mumbai, India) for 10 days⁶. The standard drug Silymarin (Ranbaxy Lab. Dewas) was administered to Group III animals in the dose of 100 mg/kg/day p.o. for 14 days. While, Groups IV was treated with ethyl acetate extract of *Bauhinia racemosa* Linn in the dose of 25 mg/kg/day, p.o. (as per acute toxicity studies) for 14 days, respectively. The CCl4, silymarin and the extracts were administered concomitantly to the respective groups of animals.

Assessment of hepatoprotective activity

All the animals were killed on day 14 under light ether anaesthesia. The blood samples were collected separately by carotid bleeding into sterilized dry centrifuge tubes and allowed to coagulate for 30 min at 37°C. The clear serum was separated at 2500 rpm for 10 min and biochemical investigations were carried out to assess liver function viz., total bilirubin⁷ serum transaminases⁸ and serum alkaline phosphatase⁹ (Raichem, San Diego, CA 92111-1203). The results are expressed as mean± SEM of six animals from each group. The data were evaluated by one-way ANOVA followed by Tukey's multiple comparison tests. P values < 0.01 were considered statistically significant. Histopathology After draining the blood, liver samples were excised, washed with normal saline and processed separately for histological observations. Initially, the materials were fixed in10% buffered neutral formalin for 48 h and then with bovine solution for 6 h. Paraffin sections were taken at 5 mm thickness, processed in alcohol-xylene series and were stained with alum haematoxylin and $eosin^{10}$.

RESULTS AND DISCUSSION

The sections were examined microscopically for histopathological changes. Histological profile of the control animals showed normal hepatocytes. Group II animals exhibited intense centrilobular necrosis (N), vacuolization (F) and macro vesicular fatty change. The sections of liver taken from the animals treated with standard drug silymarin showed the hepatic architecture, which was similar to that of control. The animals treated with ethyl acetate extract exhibited significant liver protection against the toxicant as evident by the presence of normal hepatic cords, absence of necrosis and lesser fatty infiltration. The administration of CCl4 to the animals resulted in a marked increase in total bilirubin, serum amino transaminase (SGOT and SGPT) and serum alkaline phosphatase activities. The toxic effect of CCl4 was controlled in the animals treated with the ethyl acetate extract by way of restoration of the levels of the liver function biochemistry similar to that of the standard drug silvmarin (Table 1)

The Carbon tetrachloride (CCl4) has been used to induce hepatotoxicity in experimental animals^{11, 12}. This toxic

chemical caused peroxidative degradation in the adipose tissue resulting in fatty infiltration of the hepatocytes. The increase in the levels of serum bilirubin reflected the depth of jaundice and the increase in transaminases and alkaline phosphatase was the clear indication of cellular leakage and loss of functional integrity of the cell membrane¹³. Administration of ethyl acetate extract of Rauhinia flowers showed racemosa significant hepatoprotective reactivity, which was comparable with the standard drug silymarin. The effect was more pronounced with methanol extract. Many phytochemical reports revealed that the ethanolic extract of the plant was found to contain higher concentrations of flavonoids glycosides¹⁴. The qualitative phytochemical and investigations on the ethyl acetate extract of Bauhinia racemosa Linn also showed positive for flavonoids by ferric chloride, alkaline reagent and Shinoda tests. Further, it has been reported that the flavonoid constituents of the plant possess antioxidant properties¹⁵ and was found to be useful in the treatment of liver damage¹⁶. The administration of hepatoprotective drugs may induce the hepatocytes to resist the toxic effect of CCl4. The results indicate that the ethyl acetate extract of Bauhinia racemosa Linn has significant hepatoprotective activity. This may be probably due to the higher content of flavonoids.

REFERENCES

- 1. Badgujar SB, Patil MB. Ethnomedicines for jaundice used in tribal areas of North Maharastra. Natural Product Radiance. 2008; 7(1): 79-81.
- 2. Kirtikar KR and Basu BD "Indian Medicinal Plants", International Book Distributor, Dehradun, Ist Edn. 1980; Vol.III: p.676.
- Francesca Boralli and Angelo A. lizzo. The plant kingdom as a source of anti-ulcer remedies. Phytotherapy Research. 2000; 14:8: 581-591.
- 4. Ghosh MN. Fundamentals of experimental pharmacology. 2nd ed. Calcutta Scientific Book Agency; 1984; 55.
- Jalalpure SS, Patil MB, Prakash NS, Hemalata K, Manvi FV. Hepatoprotective activity of fruits of *Piper longue* Linn. Indian J Pham Sci 2003;65;360-6
- 6. Jaiprakash B, Aland R, Karadi RV, Savadi RV, Hukkeri VI. Hepatoprotective activity of fruit pulp of *Balanites aegyptiaca*. Indian Drugs 2003;40;296-7.
- 7. Mallory HT, Evelyn EA. The determination of bilirubin with photoelectric colorimeter. J Biol Chem 1937;119:481-5.
- 8. Reitman S, Frankel S. A Colorimetric method for the determination of serum glutamic oxoloacetic and glutamic pyruvic transaminase Am J Clin Pathol, 1957, 28, 56-63.
- 9. Bessey OA, Lowery DH, Brock MJ. A method for the rapid determination of alkaline phosphatase with five cubic meters of serum. J Biol Chem 1964;164: 321-9.
- 10. Galighter AE, Koyloff EN. Essentials of practical micro techniques. Philadelphia: Lea and Febiger; 1971; 227.
- 11. Recnagel RO. Carbon tetrachloride hepatotoxicity status and future prospects, Trends Pharmacol Sci 1983, 4, 129-31.
- 12. Okuno H, Hazama H, Muraze T, Shiozaki Someshima YT. Drug metabolizing activity in rats with chronic liver injury,

carbon tetrachloride relationship with the induced by hydroxyproline content in liver. Japan J Pharmacol 1986, 41, 363-71.

13. Saraswat B, Visen PK, Patnaik GK, Dhawan BN. Anticholestic effect of picroliv, active hepatoprotective principle of Picrorhiza kurroa, against carbon tetra chloride induced cholestatis. Indian J Exp Biol 1993;31:316-8.

- Harborn JB. Phytochemical methods. 2nd ed. 1979; 112. 14.
- 15. Hesham R. El- Seedi, Shgeru Nishiyama. Chemistry of Bioflavonoids. Indian J Pharm Edu 2002, 36, 191-4.
- 16. Maurya R, Singh R, Deepak M, Handa SS, Yadav PP, Mishra PK. Constituents of Pterocarpus marsupium; an ayurvedic crude drug. Phytochemistry 2004;65: 915-20.

Design of treatment	Liver (wt/100g	Dose	SGPT	SGOT	ALP	Total Bil
	body wt)	(mg/kg)	U/L	U/L	U/L	(mg %)
Control	358 <u>+</u> 0.03		131.45 <u>+</u> 1.44	45.9 ± 0.85	164.3 <u>+</u> 3.23	0.74 ± 0.05
CCl ₄	6.5 <u>+</u> 0.35	1.25 ml/kg	222.5 <u>+</u> 4.3	345.5 <u>+</u> 2.3	384.4 <u>+</u> 18.53	2.13 <u>+</u> 0.04
Silymarin $+ CCl_4$	3.4 <u>+</u> 0.39*	100	134.0 <u>+</u> 1.15**	82.2 <u>+</u> 9.33*	213.6 <u>+</u> 5.47**	$0.6 \pm 0.06*$
Ethyl acetate extract + CCl ₄	4.6 <u>+</u> 0.45*	400	115.2 <u>+</u> 1.13*	64.4 <u>+</u> 5.35*	294.5 <u>+</u> 5.56*	0.85 ± 0.09

Table 1: Effect of Bauhinia racemosa on CCl4 treated rats

115.2 + 1.13* \overline{N} = 6 animals in each group. *P < 0.001; **P < 0.01 when compared with control. Values are expressed as mean \pm SE.

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