INTRODUCTION
Liver is a vital organ in the body and it plays an important role for the metabolism of endogenous and exogenous agents. It is the first organ to be exposed to the damaging effects of the newly formed toxic substance during metabolism. As a traditional medicine, plant products are used for the treatment of liver disorders all over the world. Tuberculosis (TB) continues to be a health problem worldwide, WHO estimated 9.27 million new cases of TB in 2007. According to DOTS (Directly Observed Treatment Shortcourse), chemotherapeutic regimens of TB involves isoniazid (H), rifampicin (R), pyrazinamide (Z) and/ or ethambutol (E) is highly effective. However, hepatotoxicity is a serious adverse drug reaction of these currently used anti-TB drugs. Therefore, a search for an alternative therapy/ supplementation of plant drugs useful for the treatment of hepatotoxicity and also the efforts to explore the mechanism of hepatoprotective effect of natural products thus carry a great clinical significance. Barleria Montana, (Synonym Barleria Purpurea) commonly known as Mountain Barleria has been traditionally used for centuries for treating wounds, diabetes, cough, inflammation and is known to possess hepatoprotective activity. The leaves of Barleria Montana (BM) are reported as having antimicrobial activity, pharmacognostic and physicochemical analysis, antibacterial activity, antidiabetic activity and proved to possess in-vitro antioxidant activity. Still there is no scientific report on Antihepatotoxic potential of 95% hydroalcoholic extract of Barleria Montana L. leaves against anti-TB drugs induced hepatotoxicity. Therefore, the present study was undertaken for antihepatotoxic effect of 95% hydroalcoholic extract of Barleria Montana against anti-TB drugs induced hepatotoxicity in rats.

MATERIAL AND METHODS
Collection and Authentication of Plant Material
Leaves of Barleria montana were collected from Tirumala hills, Andhra Pradesh. The plant was identified, authenticated and certified by Dr. K. Madhavachetty, Assistant Professor, Department of Botany, S.V.University, Tirupathi and A.P., India.

Preparation of plant extract
The air dried powder was extracted in Soxhlet apparatus using 95% hydroalcoholic solution as solvent. Appearance of colorless solvent in the siphon tube was taken as the endpoint of extraction. The extract was concentrated to ¼ of its original volume by distillation.

Acute toxicity studies
Acute toxicity studies were performed for 95% hydroalcoholic extract of Barleria montana according to OECD guidelines 423. 10 mice were selected for the study and oral administration of 95% hydroalcoholic extract of Barleria montana at a dose of 100, 1000, 2000, 4000 mg/kg given at 48 hrs interval simultaneously. In this acute toxic study, animals were observed for any changes in consumption of food, water; body weight; behavioral changes and mortality rates.

Animals
Healthy adult albino rats (150–250 gm) were used and they were purchased from In vivo biosciences, Bangalore. The animals were housed in clean metabolic cages, maintained in controlled temperature (22±3°C) and light cycle (12 hour light and 12 hour dark). They were fed with standard pellet diet and water ad libitum. The protocol was approved by the Institutional animal ethical committee (IAEC) of Krishna Teja Pharmacy College (1521/PO/a/11/CPCSEA).
Study protocol
Hepatotoxicity was induced by using H-Isoniazid (27 mg/kg, p.o), R-Rifampicin (40 mg/kg, p.o), Z-Pyrazinamide (66 mg/kg, p.o) and E-Ethambutol (53 mg/kg, p.o) were dissolved in normal saline for 35 days and Silymarin (100 mg/kg, p.o) was used as the standard. The oral doses of anti-TB drugs were extrapolated from daily human dose using the conversion table based on body surface area.

Experimental procedure
Experimental animals were randomly divided into 5 groups, each group containing 6 animals and the treatment schedule for 35 days as follows. Group I: Control (Normal saline 1ml/kg, p.o), Group II: Hepatic control (anti – TB drugs – HRZE, p.o.), Group III: Silymarin (100 mg/kg, p.o) + one hour prior administration of anti-TB drugs, Group IV: 95% hydroalcoholic extract of Barleria Montana (250 gm/kg, p.o)+one hour prior administration of anti-TB drugs and Group V: 95% hydroalcoholic extract of Barleria Montana (500 gm/kg, p.o) + one hour prior administration of anti-TB drugs. On 36th day, blood is collected for estimation of biochemical parameters. On the same day, liver is removed and stored in 10% formalin solution for the estimation of antioxidant parameters; and processing for histopathological studies.

RESULTS

Estimation of Biochemical and Antioxidant Parameters
SGOT and SGPT were estimated by Reitman and Frankel method, ALP was estimated by kind king’s method. Total Bilirubin, total cholesterol were estimated by Jendrassik and Grof’s method and CHOD/POD method respectively. Antioxidant parameters were estimated by according to reported methods SOD, CAT, GSH, GPx, GRx and lipid peroxidation.

Histopathological studies
Livers from rats were fixed in 10% neutral formalin solution, dehydrated in graded alcohol and embedded in paraffin. Fine sections obtained were mounted on glass slides and counter-stained with Hematoxylin Eosin (H&E) for light microscopic analyses.

Statistical analysis
The results are presented as Mean ± S.E.M (n=6 in each group). Analyses were performed using One-way ANOVA followed by Tukey posthoc for the difference between the control and treatment groups.

Table 1: Effect of Barleria Montana on serum SGOT, SGPT, ALP, TB, Total cholesterol and HDL on anti-TB drugs induced hepatotoxicity in rats

<table>
<thead>
<tr>
<th>Groups</th>
<th>Treatment</th>
<th>SGOT (IU/L)</th>
<th>SGPT (IU/L)</th>
<th>ALP (IU/L)</th>
<th>TB (IU/L)</th>
<th>Total cholesterol (mg/dl)</th>
<th>HDL (mg/dl)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Group-I</td>
<td>Control</td>
<td>122.67±6.38</td>
<td>63.33±4.02</td>
<td>182.66±27</td>
<td>0.06±0.01</td>
<td>83.56±6.3</td>
<td>33.8±2.1</td>
</tr>
<tr>
<td>Group-II</td>
<td>Anti-TB Drugs (Hepatic control)</td>
<td>386.50±14.60</td>
<td>639.83±5.46</td>
<td>315.23±7.46</td>
<td>0.39±0.01</td>
<td>131.3±16.2</td>
<td>17.4±1.9</td>
</tr>
<tr>
<td>Group-III</td>
<td>Silymarin (100mg/kg)</td>
<td>166.67±2.90</td>
<td>136.33±1.53</td>
<td>138.7±6.55</td>
<td>0.08±0.02</td>
<td>88.7±5.9</td>
<td>33.6±2.4</td>
</tr>
<tr>
<td>Group-IV</td>
<td>Barleria Montana (250mg/kg)</td>
<td>277.50±7.68</td>
<td>232.67±7.45</td>
<td>205±3.50</td>
<td>0.08±0.01</td>
<td>116.9±9.5</td>
<td>28.5±2.8</td>
</tr>
<tr>
<td>Group-V</td>
<td>Barleria Montana (500mg/kg)</td>
<td>181.67±6.58</td>
<td>147.16±1.15</td>
<td>150.83±7.52</td>
<td>0.06±0.04</td>
<td>89.1±7.1</td>
<td>34.1±1.7</td>
</tr>
</tbody>
</table>

Data are expressed as Mean ± S.E.M (n=6), One-way ANOVA Tukey posthoc; *p≤0.05 vs. Control (Group I); **p≤0.01 vs. Hepatic control (Group II); ***p≤0.001 vs. Hepatic control (Group II)

Table 2: Effect of Barleria Montana on antioxidant parameters on anti-TB drugs induced hepatotoxicity in rats

<table>
<thead>
<tr>
<th>Group</th>
<th>Treatment</th>
<th>SOD (umoles/min/mg)</th>
<th>CAT (umoles/min/mg)</th>
<th>GPx (umoles/min/mg)</th>
<th>GRx (umoles/min/mg)</th>
<th>TARRS (mg/min/mg)</th>
<th>GSH (mg/min/mg)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Group I</td>
<td>Control</td>
<td>3.55±0.26</td>
<td>34.99±0.98</td>
<td>28.99±0.64</td>
<td>30.81±0.94</td>
<td>34.95±1.9</td>
<td>2.47±0.12</td>
</tr>
<tr>
<td>Group II</td>
<td>Anti-TB Drugs (Hepatic Control)</td>
<td>3.62±0.66</td>
<td>15.49±0.57</td>
<td>12.64±0.66</td>
<td>18.34±0.31</td>
<td>88.29±5.89</td>
<td>0.74±0.06</td>
</tr>
<tr>
<td>Group III</td>
<td>Silymarin (100mg/kg)</td>
<td>3.71±0.19</td>
<td>33.89±0.08</td>
<td>29.91±0.82</td>
<td>30.86±0.44</td>
<td>33.89±3.23</td>
<td>2.27±0.09</td>
</tr>
<tr>
<td>Group IV</td>
<td>Barleria Montana (250mg/kg)</td>
<td>2.67±0.43</td>
<td>28.88±0.05</td>
<td>23.17±0.23</td>
<td>25.87±0.58</td>
<td>41.98±3.07</td>
<td>1.53±0.05</td>
</tr>
<tr>
<td>Group V</td>
<td>Barleria Montana (500mg/kg)</td>
<td>3.27±0.04</td>
<td>34.09±0.29</td>
<td>30.69±0.31</td>
<td>31.11±0.26</td>
<td>34.32±2.13</td>
<td>2.37±0.04</td>
</tr>
</tbody>
</table>

Data are expressed as Mean ± S.E.M (n=6), One-way ANOVA Tukey posthoc; *p≤0.05 vs. Control (Group I); **p≤0.01 vs. Hepatic control (Group II); ***p≤0.001 vs. Hepatic control (Group II)

RESULTS

On acute toxicity studies
The 95% hydroalcoholic extract of Barleria Montana was found to be safe as no animal died even at the dose of 4000mg/kg when administered orally and the animals did not show any gross behavioral changes.

On Biochemical Parameters
Animals treated with anti-TB drugs (hepatic control) showed a significantly elevated levels of SGOT, SGPT, alkaline phosphatases, total bilirubin and total cholesterol levels; and significantly decreased in HDL levels when compared to control group. 95% hydroalcoholic extract of Barleria Montana 250 mg/kg and 500 mg/kg given with one hour prior administration of anti-TB drugs showed a significant decreased serum diagnostic liver enzymes and increased HDL levels when compared to hepatic control. The results are presented in the Table I.

In vivo Antioxidant parameters
In the present study, antioxidant parameters were assessed in the liver homogenate. Oral administration of anti-TB drugs (hepatic control) decreased SOD, CAT, GPx, GRx, GSH and significantly increased TBARS when compared to control group. Pretreatment of 95% hydroalcoholic extract of Barleria Montana 250mg/kg and 500mg/kg with one hour
prior administration of anti-TB drugs showed significantly increased the enzymatic and non-enzymatic levels and significantly decreased TBARS levels when compared to hepatic control. The results are presented in Table 2.

**Histopathological study of the liver**
Hepatic control group animals showed significant liver cell necrosis and inflammation in the centrilobular region with portal triaditis as compared to normal control group. 95% hydroalcoholic extract of *Barleria Montana* 500mg/kg showed protective effect on the hepatocellular necrosis and their lobular structure is normal similarly like control and silymarin treated animals (Figure 1 to 5).

**DISCUSSION**
Anti-TB drugs like Isoniazid, Rifampicin, and pyrazinamide are independently itself are potentially hepatotoxic\(^\text{18}\), when given in combination; their toxic effect is enhanced in a synergistic manner. In the present study, the combination of anti-TB drugs is used as a tool to induce the hepatotoxicity in experimental animals\(^\text{19}\). The exact mechanisms of hepatotoxicity by these drugs are not clear, but several researchers have suggested that hepatotoxicity is mediated through release of reactive/toxic metabolites binding covalently with liver cell macromolecules causing liver injury\(^\text{20}\). Secondly, cytochrome P450 2E1 is involved in induction of hepatic damage\(^\text{21}\). Third reason is oxidative stress can be explained by release of free radicals, which is source for destruction and damage to cell membranes\(^\text{22}\). Finally, an alteration of various cellular defense mechanisms involves enzymatic and non-enzymatic components such as reduced GSH have been reported in INH and RIF induced hepatotoxicity\(^\text{23}\). The reduced GSH mediated by free radical injury leads to progress in lipid peroxidation\(^\text{24}\). 95% hydroalcoholic extract of *Barleria Montana* is used as supplementation with anti-TB drugs to reduce hepatotoxicity caused by anti-TB drugs. In hepatic control animals elevated levels of serum diagnostic enzymes such as SGOT, SGPT and ALP levels indicate hepatic injury. At the time of hepatic injury, these enzymes leak out from liver into the blood due to the tissue damage. Pretreatment with 95% hydroalcoholic extract of *Barleria Montana*, the levels of these marker enzymes in serum were near normal, this may be a consequence of the stabilization of plasma membrane as well.
as repair of hepatic tissue damage caused by anti-TB drugs. Hepatotoxicity is characterized by cirrhosis liver condition which in turn increased the bilirubin release, which was observed in anti-TB drug treatment. Pretreatment with 95% hydroalcoholic extract of Barleria Montana restored the level of bilirubin to normal, suggesting that Barleria Montana stabilizes biliary dysfunction of rat liver. The cholesterol levels are increased which might be due to uptake of LDL from the blood by the tissues. Thus, 95% hydroalcoholic extract of Barleria Montana may be effective on reduced cholesterol synthesis, and there by causes increased HDL levels.

The body has a defense mechanism such as enzymatic and non-enzymatic antioxidants to prevent and neutralize the free radicals induced damage. Enzymatic antioxidants like SOD and CAT play an important role in the elimination of free radicals. The suppression of SOD and CAT activity as a result of hepatic damage was observed in hepatic control animals of our study also with anti-TB drugs. But 95% hydroalcoholic extract of Barleria Montana has significantly recovered the levels of SOD and CAT, which indicates that it possesses antioxidant activity. Depleted GSH level with increased lipid peroxidation in livers of diabetic rats.

Increase in the level of lipid per oxidation in liver reflected the hepatocellular damage causes degradation of cellular macromolecules leading to tissue damage. A marked increase in the concentration of TBARS in anti-TB drugs indicated that enhanced lipid per oxidation, failure of the antioxidant defense mechanism, and increase free radical production leading to oxidative stress induced cell death. 95% hydroalcoholic extract of Barleria Montana showed ability to prevent anti-TB drugs induced elevation of TBARS level, suggesting that Barleria Montana inhibited the hepatic lipid per oxidation. It implies that reduction in free radicals production and subsequent decrease in damage to the hepatocellular membrane.

CONCLUSION
From the results, it is clear that 95% hydroalcoholic extract of Barleria Montana has hepatoprotective activity at the dose of 500 mg/kg as compared to control and silymarin treated animals. On phytochemical investigation of 95% hydroalcoholic extract of Barleria Montana revealed the presence of flavonoids, steroids, triterpenoids, tannins and glycosides, which contributes antioxidant potential and probably this, may be responsible for hepatoprotective activity. Histopathological studies of the liver evidenced that Barleria Montana attenuated the hepatocellular necrosis and inflammation, which might be attributed to its hepatoprotective effects. Further investigations are required for the identification and isolation of constituents, which are responsible for the hepatoprotective activity.

REFERENCES


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