



BIOCHEMICAL ANALYSIS OF CABBAGE (*BRASSICA OLERACEA*) AFTER INFECTION OF PEST

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Article Received on: 16/03/13 Revised on: 07/04/13 Approved for publication: 11/05/13

DOI: 10.7897/2230-8407.04628

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ABSTRACT

Plants of Brassicaceae family include worldwide commercially grown crops. Experiments on this crop were conducted during the year 2011 in Vidhania and Jaisinghpura of Jaipur district Rajasthan, India. The aim of this study is to find out primary metabolites like chlorophyll, sugar starch protein total phenol of cauliflower. Levels of plant metabolites are strongly affected by genetic and environmental factors. Growth factors such as light, temperature, humidity, type of soil, application of fertilizers, damage caused by microorganisms and insects, stress induced by UV radiation, heavy metals and pesticides all alter metabolite composition of plants. Different types of pests' cause changes in plant metabolite production. The results revealed the evidence of different infestation of cabbage by common herbivores. In this review we report primary metabolites of the cabbage along with the quantification after the pests' effect.

Keywords: Brassicaceae, cabbage, fertilizers, humidity, primary metabolites.

INTRODUCTION

Vegetables contain essential components of human nutrition. Nutrients have traditionally been viewed as food components that either cannot be synthesized in the body (for example, vitamin C) or whose synthesis requires a specific factor that may in certain circumstances be absent or inadequate (for example, some amino acids, fatty acids, and vitamins). However, there is now recognition that many other compounds of plant food, such as dietary fibre, flavonoids, sterols, phenolic acids, and glucosinolates are associated with lower disease risk. This has been widely reported, sometimes erroneously, by the popular press. Nevertheless, a large number of phytochemical capable of antioxidant, anti mutagenic, cytotoxic, antifungal, and antiviral activities have been identified in broccoli, cauliflower, Brussels sprouts, turnips, kale, mustard, asparagus, spinach, lettuces and endives^{1,2}. The links between fruit and vegetable consumption and protection against cancers of stomach, oesophagus, lung, pharynx, endometrium, pancreas and colon have also been extensively reported^{3,4}. These phytochemicals have been linked to many positive effects on human health, including coronary heart diseases, diabetes, high blood pressure, cataracts, degenerative diseases and obesity^{5,6}. Brassicaceae family plants are the most popular vegetables consumed all over the world and considered to be a good source of bioactive phytochemicals. Cabbage is a good source of beta-carotene, vitamin C and fibre. It is a cruciferous vegetable and has been shown to reduce the risk of some cancers, especially those in the colorectal group. This is possibly due to the glucosinolates found in Cole crops, which serve as metabolic detoxicants or due to the sulphoraphane content, also responsible for metabolic anti-carcinogenic activities. Purple cabbage also contains anthocyanins, which in other vegetables have been proven to have anti-carcinogenic properties⁷. Along with other Cole crops, cabbage is a source of indole-3-carbinol, a chemical that boosts DNA repair in cells and appears to block the growth of cancer cells⁸. Research suggests that boiling these vegetables reduces their anti-carcinogenic properties⁹. Additionally, these species and varieties are increasingly becoming a research model in plant science, as a consequence of the importance of their primary and

secondary metabolites. Plant interaction with environmental stress factors including animals and insects' herbivore, pathogens, metal ions, light, among others, is known to lead to the activation of various defence mechanisms resulting in a qualitative and/or quantitative change in plant metabolite production. Pre-harvest and/or post-harvest conditions are also known to affect this, since plants produce signalling molecules (e.g. salicylic acid, jasmonic acid etc.), that cause a direct or indirect activation of metabolic pathways. That ultimately affects the production of phytochemicals, such as carbohydrates (sucrose and glucose), amino acids, phenolics (phenylpropanoids and flavonoids) and glucosinolates. These phytochemicals have diverse applications due to their antimicrobial, antioxidant and anticarcinogenic properties, but on the other hand these compounds or their breakdown products can act as anti-nutritional factors in diet. Primary metabolites, for example; sugars, proteins, lipids, and starch are of prime importance and essentially required for growth of plants¹⁰. Nutrient composition of vegetables is very complex and difficult to assess. Levels of plant metabolites are strongly affected by genetic and environmental factors as well as transportation and storage conditions. Growth factors such as light, temperature, humidity, type of soil, application of fertilizers, damage caused by microorganisms and insects, stress induced by UV radiation, heavy metals, and pesticides all alter metabolite composition of plants¹¹. Plant primary metabolism, which is shared with insects and other living organisms, provides carbohydrates, amino acids, and lipids as essential nutrients for the insect. Most of the insects considered common vegetable pests undergo a developmental process known as metamorphosis, which simply means that the insect changes form during its life. Metamorphosis may be complete or incomplete. Complete metamorphosis consists of four stages-egg, larva, pupa, and adult. Vegetable parts chewed by the insects this processes is known as herbivory. Pest and insects are also affecting the primary metabolites of vegetable plants. Pest of vegetables is also reduces the quality of food. Food quality is largely determined by the availability of these nutrients (protein sugar carbohydrates) and its importance for longevity, size, fecundity, and death rates in herbivorous insects has been recognized early on by Painter¹².

Table 1: Various Primary Metabolites in Pest Free Vegetable Parts

Primary metabolites	leaf	Stem	Root
chlorophyll a	6.03±0.31 gm/gdw	2.17±0.41 gm/gdw	0.00 gm/gdw
chlorophyll b	6.11±0.53 gm/gdw	2.82±0.63 gm/gdw	0.00 gm/gdw
carotenoid	3.51±0.18 gm/gdw	1.05±0.44 gm/gdw	0.34±0.6 gm/gdw
sugar	4.00±0.65 gm/gdw	2.93±0.43 gm/gdw	2.00±0.49 gm/gdw
starch	6.00±0.56 gm/gdw	4.08±0.13 gm/gdw	3.25±0.35 gm/gdw
proteins	69.57±0.56 µg/mg	70.52±0.25 µg/mg	61.01±0.57 µg/mg
Total phenol	86.14±0.56 µg/mg	80.78±0.35 µg/mg	79.18±0.34 µg/mg

Table2: Various Primary Metabolites in Pest Infected Vegetable Parts

Primary metabolites	leaf	Stem	Root
chlorophyll a	4.14±0.3 gm/gdw	1.38±0.4 gm/gdw	0.00 gm/gdw
chlorophyll b	5.78±0.34 gm/gdw	2.82±0.23 gm/gdw	0.00 gm/gdw
carotenoid	1.71±0.81 gm/gdw	1.57±0.43 gm/gdw	1.28±0.16 gm/gdw
sugar	4.98±0.46 gm/gdw	3.15±0.14 gm/gdw	4.27±0.37 gm/gdw
starch	5.07±0.43 gm/gdw	2.34±0.32 gm/gdw	1.32±0.95 gm/gdw
proteins	72.65±0.53 µg/mg	87.34±0.32 µg/mg	76.42±0.11 µg/mg
Total Phenol	98.87±0.7 µg/mg	87.28±0.4 µg/mg	88.97±0.19 µg/mg

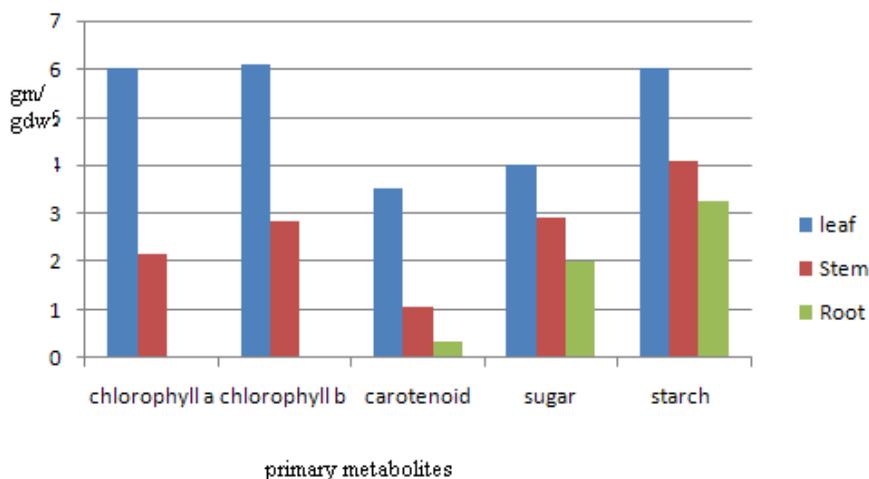


Figure 1: Primary metabolites of fresh cabbage

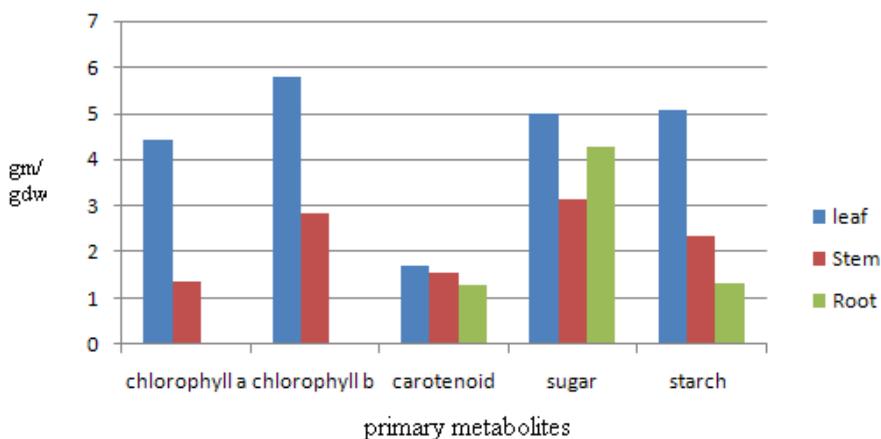


Figure 2: Primary metabolites of pest infected cabbage

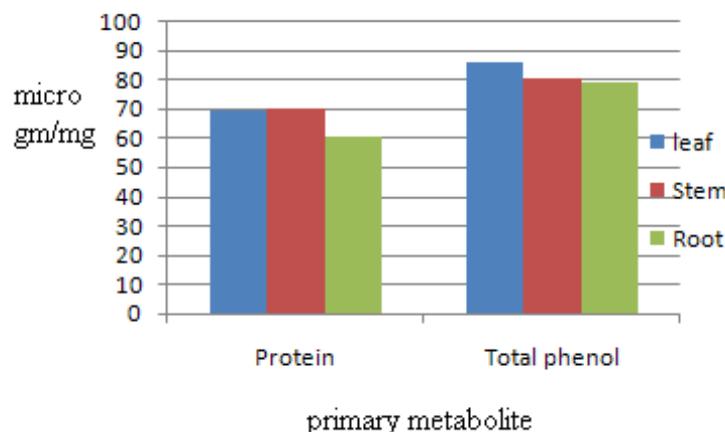


Figure 3: Protein and Phenol content of fresh cabbage

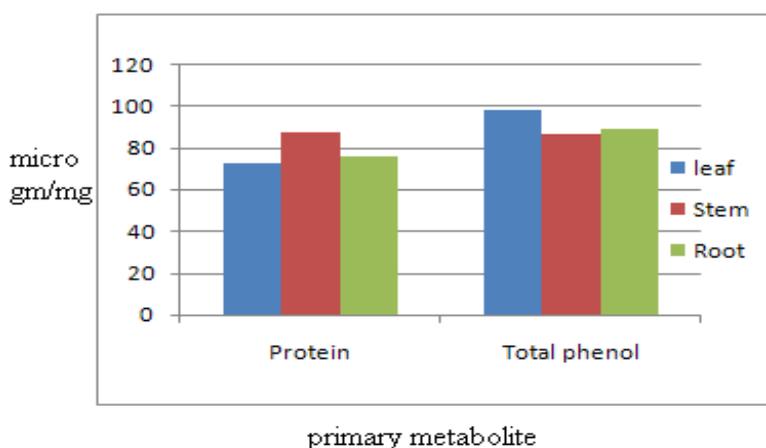


Figure 4: Protein and Phenol content of pest infected cabbage

Numerous studies have shown that herbivore causes large-scale changes in gene expression¹³⁻²⁰. In this article we report primary metabolites of the Cauliflower along with the quantification after the effect of pests like cut worm white fly diamond back moth flea beetles cabbage lopper etc.

MATERIAL AND METHODS

Collection of plant material

Vegetable sample were collected from Vidhania and Jaisinghpura khor village of Jaipur districts Rajasthan, India. Fresh cabbage and pest infected cabbage were collected from the different fields

Preparation of extracts

The stem, leaf and roots of cabbage was cut into small pieces, dried and powdered. The resultant was then subjected for successive extraction with petroleum ether, benzene, chloroform, ethanol and water with soxhlet apparatus. The extracts were then concentrated in vacuum under reduced pressure using rotary flash evaporator and dried in desiccators. These extracts were then subjected to preliminary phytochemical screening for the detection of various plant constituents. Each of these extracts was processed further to evaluate the presence of carbohydrates, proteins, starch and chlorophyll following the established protocols. The powder was treated with acids like 1N HCl,

H₂SO₄, HNO₃, Acetic acid and alkaline solutions like 1N NaOH and ammonia. Root, stem and leaf parts of cauliflower were evaluated quantitatively to estimate the total levels of soluble sugars, starch, proteins, lipids and phenols following the established methods for the sugars, starch, lipid, protein¹⁹ and phenol²⁰. All experiments were repeated five times for precision and values were expressed in mean \pm standard deviation in terms of air dried material.

RESULTS AND DISCUSSION

Primary metabolites proteins, lipid, soluble sugar, starch and total phenol contents are quantified in different plant parts (root, stem and leaves) and shown in (Table 1). Fresh vegetable part contained total sugar (4.00 \pm 0.65gm/gdw) in leaves, starch (6.00 \pm 0.56gm/gdw) in leaves, proteins (70.52 \pm 0.25 μ g/mg) in stem, carotenoid (3.51 \pm 0.18gm/gdw) in leaves, chlorophyll a (6.03 \pm 0.31gm/gdw) and chlorophyll b (6.11 \pm 0.53gm/gdw) in leaves. Leaves show maximum concentration of metabolites as compared to its roots and stem. Pest infected vegetable parts contained total sugar (4.98 \pm 0.46gm/gdw) in leaves starch (5.07 \pm 0.43gm/gdw) in leaves, proteins (87.34 \pm 0.32 μ g/mg) in stem, carotenoid (1.71 \pm 0.81gm/gdw) in leaves, chlorophyll a (4.14 \pm 0.30gm/gdw) and chlorophyll b (5.78 \pm 0.34 gm/gdw) in leaves. Leaves show maximum primary metabolites as compared to its roots and stem. Plant synthesizes primary

metabolites (lipid, protein, starch, sugars, phenol etc.) for the normal growth and development of itself. These results are suggestive of primary bioactive compound of commercially importance and may result in great interest in plants pharmaceuticals. Therefore, economic use depends partially on the quantitative and qualitative aspects of their organic reserves, specially carbohydrates, proteins, phenols and lipids. These primary metabolites further can be used for biosynthesis of secondary metabolites or bioactive compounds.

REFERENCES

- Prior RL, Cao G. Antioxidant phytochemical in fruits and vegetables diet and health implications. HortScience 2000; 35(4): 588–92.
- Goldberg G. Plants diet and health. The report of a British nutrition foundation task force. Oxford, U.K., Blackwell Publishing Ltd; 2003. <http://dx.doi.org/10.1002/9780470774465>
- Temple NJ, Gladwin KK. Fruit vegetables and the prevention of cancer: research challenges. Nutrition 2003; 19(5): 467–70. [http://dx.doi.org/10.1016/S0899-9007\(02\)01037-7](http://dx.doi.org/10.1016/S0899-9007(02)01037-7)
- Hung HC, Huang MC, Lee JM, Wu DC, Hsu HK, Wu MT. Association between diet and esophageal cancer in Taiwan J Gastroenterol Hepatol 2004; 19(6): 632–7. <http://dx.doi.org/10.1111/j.1440-1746.2004.03346.x> PMID:15151616
- Liu S, Manson JE, Lee IM, Cole SR, Hennekens CH, Willett WC, Buring JE. Fruit and vegetable intake and risk of cardiovascular disease: the women's health study. Am J. Clin Nutr 2000; 72: 922–930. PMID:11010932
- Djouss'e L, Arnett DK, Coon H, Province MA, Moore LL, Ellison RC. Fruit and vegetable consumption and LDL cholesterol. The National Heart Lung and Blood Institute Family Heart Study. Am. J. Clin. Nutr 2004; 79: 213–220.
- Katz and Weaver. Encyclopedia of Food and Culture Scribner Library of Daily Life 2009; 3: 283–284.
- Fan S, Meng Q, Auburn K, Carter T, Rosen EM. BRCA1 and BRCA2 as molecular targets for phytochemicals indole-3-carbinol and genistein in breast and prostate cancer cells. British Journal of Cancer 2006; 94(3): 407–426. <http://dx.doi.org/10.1038/sj.bjc.6602935> PMID:16434996 PMID:2361140
- Wu Y, Feng X, Jin Y, Wu Z, Hankey W, Paisie C, Li L, Liu F. A Novel Mechanism of Indole-3-Carbinol Effects on Breast Carcinogenesis Involves Induction of Cdc25A Degradation. Cancer Prevention Research 2010; 3(7): 818–828. <http://dx.doi.org/10.1158/1940-6207.CAPR-09-0213> PMID:20587702
- Santhi R, Lakshmi G, Priyadarshani AM and Anandaraj AL. Phytochemical screening of Nerium oleander leaves and *Momordica charantia* leaves. International Research Journal of Pharmacy 2011; 2(1): 131-135.
- Orcutt DM, Nilsen ET. The Physiology of Plants under Stress, A biotic Factors Wiley 07; 1996.
- Painter RH. The food of insects and its relation to resistance of plants to insect attack. Am Nat 1936; 70: 547-566. <http://dx.doi.org/10.1086/280696>
- Cheong YH, Chang HS, Gupta R, Wang X, Zhu T, Luan S. Transcriptional profiling reveals novel interactions between wounding, pathogen, abiotic stress and hormonal responses in Arabidopsis. Plant Physiology 2002; 129: 661–677. <http://dx.doi.org/10.1104/pp.002857> PMID:12068110 PMID:161692
- Reymond P, Bodenhausen N, Van Poecke RM, Krishnamurthy V, Dicke M, Farmer EE. A conserved transcript pattern in response to a specialist and a generalist herbivore. Plant Cell 2004; 16: 3132–3147. <http://dx.doi.org/10.1105/tpc.104.026120> PMID:15494554 PMID:527203
- Smith CM, Rodriguez BM, Karlsson J, Campbell MM. The response of the poplar transcriptome to wounding and subsequent infection by a viral pathogen. New Phytologist 2004; 164: 123–136. <http://dx.doi.org/10.1111/j.1469-8137.2004.01151.x>
- Voelckel C, Baldwin IT. Herbivore-induced plant vaccination. Part II. Array-studies reveal the transience of herbivore-specific transcriptional imprints and a distinct imprint from stress combinations. Plant J 2004; 38: 650–663. <http://dx.doi.org/10.1111/j.1365-313X.2004.02077.x> PMID:15125771
- Zhu-Salzman K, Salzman RA, Ahn JE, Koiwa H. Transcriptional regulation of sorghum defense determinants against a phloem-feeding aphid. Plant Physiol 2004; 134: 420–431. <http://dx.doi.org/10.1104/pp.103.028324> PMID:14701914 PMID:316321
- Ralph SG, Yueh H, Friedmann M, Aeschliman D, Zeznik JA, Nelson CC, Butterfield YSN, Kirkpatrick R, Liu J, Jones SJM et al. Conifer defense against insects: microarray gene expression profiling of sitka spruce (*Picea sitchensis*) induced by mechanical wounding or feeding by spruce budworms (*Choristoneura occidentalis*) or white pine weevils (*Pissodes strobi*) reveals large-scale changes of the host transcriptome. Plant Cell Environ 2006; 29: 1545–1570. <http://dx.doi.org/10.1111/j.1365-3040.2006.01532.x> PMID:16898017
- Thompson GA, Goggin FL. Transcriptomics and functional genomics of plant defense induction by phloem-feeding insects. J Exp Bot 2006; 57: 755–766. <http://dx.doi.org/10.1093/jxb/erj135> PMID:16495409
- Broekgaarden C, Poelman E, Steenhuis G, Voorrips R, Dicke M, Vosman B. Genotypic variation in genome-wide transcription profiles induced by insect feeding: *brassica oleracea*, *Pieris rapae* interactions. BMC Genomics 2007; 8: 239-240. <http://dx.doi.org/10.1186/1471-2164-8-239> PMID:17640338 PMID:1940009
- Lehninger A. Principles of Biochemistry, 3rded. Worth Publishers, New York; 1993.p.184-185
- Lowery OH, Rosenbrough NJ, Farr AL, Randall RJ. Protein measured with folin phenol reagent. J Bio Chem 1951; 193: 265-275.

Cite this article as:

Deeplata Sharma and D.V. Rao. Biochemical analysis of Cabbage (Brassica oleracea) after infection of pest. Int. Res. J. Pharm. 2013; 4(6):127-130

Source of support: Nil, Conflict of interest: None Declared