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Research Article

UV-SPECTROPHOTOMETRIC DETERMINATION OF ROSUVASTATIN AND NIACIN INDIVIDUALLY AND COMBINED TABLET DOSAGE FORM BY SIMULTANEOUS EQUATION AND ABSORBANCE RATIO METHOD

Narayankar Savita M.*, Sakpal Promod. H., Bhingare Chandrashekhar. L.

Deptartment of Advanced Quality assurance Techniques/Pharmaceutical chemistry, Marathwada Mitra Mandal's college of Pharmacy, Savitribai Phule Pune University, Pune, Maharashtra, India

*Corresponding Author Email: Savita.narayankar1985@gmail.com

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ABSTRACT

Two UV-spectrophotometric methods were developed and validated for quantitative determination of Rosuvastatin and niacin in combined tablet dosage form . Method I is based on the simultaneous equation and method II is based on the absorbance ratio method. The absorbance maxima were found to be at 241 and 262nm in water for the Rosuvastatin and niacin respectively. Beer's law is obeyed in the concentration range $5-40\mu g/ml$ with correlation coefficient within range of 0.998 for both the drugs. The accuracy of the methods was assessed by recovery studies and found to be 98-105% for Rosuvastatin and niacin respectively.

Keywords: Simultaneous equation, absorbance ratio method, Rosuvastatin and niacin

INTRODUCTION

Rosuvastatin (statin) HMG-CoA reductase inhibitor and niacin (nicotinic acid) are used, in the primary and secondary prevention of coronary heart disease, carotid artery disease and other atherosclerotic vascular diseases. In US guidelines, the lowering of low-density lipoprotein cholesterol (LDL-C) is the primary goal of lipid-modifying therapy in patients with atherosclerotic disease and those at risk for atherosclerotic disease due to dyslipidemia.

However, in patients with primary hyperlipidemia and atherogenic dyslipidemia and (i.e. those with high triglyceride levels, low high-density lipoprotein cholesterol [HDL-C] levels and small dense LDL particles), LDL-C levels may underestimate the cardiovascular risk. Therefore, the US guidelines recommend lowering both LDL-C and non-HDL-C in patients with hypertriglyceridemia.

In available lipid-modifying drugs, statins are the most effective for lowering plasma LDL-C and are considered the cornerstone of treatment for dyslipidemia and hyperlipidemia.

Niacin at pharmacological doses, displays wide-ranging lipid-modifying activity, reducing levels of all atherogenic lipid and lipoprotein subclasses, including total cholesterol, LDL-C, non-

HDL-C, triglycerides, apolipoprotein B, and lipoprotein(a), and also significantly increasing levels of HDL-C and apolipoprotein A. Furthermore, the combination of two lipid-lowering agents in one formulation may potentially improve patient compliance. Niacin is also used in the treatment of hyperlipidemia because it reduces very low density lipoprotein (VLDL), a precursor of low density

lipoprotein (LDL) or "bad" cholesterol, secretion from the liver and inhibits cholesterol synthesis

Literature survey revealed that numerous methods have been reported for estimation of Rosuvastatin and Niacin in pharmaceutical formulations individually or with other drug combination but no UV-spectrophotometric method has been reported for this combination. Present study involves development of UV-spectrophotometric method which is simple, economical, sensitive and rapid for quantification of Rosuvastatin and Niacin in individual as well as combined tablet dosage forms as well as subsequent validation of developed method according to ICH guidelines.

MATERIALS AND METHODS Instrumentation

UV-Visible Spectrophotometer instrument

Make: - Shimadzu Model: - UV 1800 Software: - UV Probe

Shimadzu Ultraviolet-visible spectrophotometer UV 1800 is a computer controlled double beam scanning spectrophotometer. It covers the range from 200-1000 nm with setting accuracy at 0.2nm.

Selection of common solvent

On the basis of solubility of both the drugs water is selected as common solvent for developing more economical and simple method.

Preparation of Standard Stock Solution

A stock solution of Rosuvastatin and Niacin was prepared by accurately weighed 50mg of drug, transferred to 50ml of volumetric flask, containing 50ml of Double distilled water dissolving it to obtain final standard solution of 1mg/ml of Rosuvastatin and Niacin. Pipette out 10ml and makeup the volume to 100ml to get solution of $100\mu g/ml$.

Determination of \(\lambda \) max

The standard solution of Rosuvastatin and Niacin were separately scanned at different concentration in the range of 200-400 nm and the λ max was determined for each drug. The λ max Rosuvastatin and Niacin were found to be 241nm and 262nm respectively and 254 nm as λ max of common absorbance (isobestic wavelength).

METHOD VALIDATION² Linearity and calibration curve

A series of standard solution were prepared having concentration in the range of 5-40µg/ml for both Rosuvastatin and Niacin The absorbance of resulting solutions were measured at $\lambda max241nm$, 262nm and 254 nm and calibration curves were plotted. Both the drugs obeyed linearity in the concentration range. (Table 1-3, Figure 1 & 2)

Method I: Simultaneous equation method⁶

This method of analysis was based on the absorption of Rosuvastatin and niacin at the wavelength maximum of each other. Two wavelengths selected for the development of simultaneous equations were 241nm and 262nm which were λ max of Rosuvastatin and niacin respectively. The absorbances of Rosuvastatin and niacin measured at selected wavelengths. Absorptivity values were calculated.

The concentrations of both the drugs in mixture can be calculated by using following equations:

$$C_{x} = \frac{A_{2}ay_{1} - A_{1}ay_{2}}{ax_{2}ay_{1} - ax_{1}ay_{2}}$$
 eqn -(1)

$$C_{y} = \frac{A_{1}ax_{2} - A_{2}ax_{1}}{ax_{2}ay_{1} - ax_{1}ay_{2}} eqn - (2)$$

Where,

 A_1 and A_2 are absorbances of mixture at 241nm and 262nm respectively.

 ax_1 and ax_2 are the absorptivities of Rosuvastatin at 241nm and 262nm respectively.

Ay₁and ay₂ are the absorptivities of Niacin at 241nm and 262nm respectively.

C_x and C_y are concentrations of Rosuvastatin and niacin respectively

Precision

The intra-day precision study of Rosuvastatin and niacin was carried out by estimating the correspondence responses six times on the same day with $10\mu g/ml$ concentration and inter-day precision study of Rosuvastatin and niacin was carried out by estimating the correspondence responses six times next day with $10\mu g/ml$ concentration. (Table 4)

Accuracy (recovery test)

The accuracy of the method was done by recovery study. The recovery experiments were performed by adding known amounts of the pure drug to the preanalyzed sample. The recovery was done at three levels: 50%, 100%, and 150% of the label claim. Three samples were prepared for each recovery level. (Table 5)

Method II: Absorbance ratio method/Q-analysis method

In quantitative assay of two components by absorption ratio method (Q-analysis), absorbances were measured at the isobestic wavelength (254 nm) and maximum absorption of one of the two components. From overlain spectra of Rosuvastatin and Niacin shown in figure no.4, absorbances were measured at the selected wavelengths of 254 nm (isobestic wavelength) and 262 nm (wavelength of maximum absorption of Niacin). From the following sets of equations, the concentration of each component in sample solution can be calculated. (Table 6 & 7, Figure 3 & 4)

$$C_{x} = \frac{Q_{m} - Q_{y}}{Q_{x} - Q_{y}} \times \frac{A_{1}}{ax_{1}} \qquad eqn - (3)$$

$$C_{y} = \frac{Q_{m} - Q_{x}}{Q_{y} - Q_{x}} \times \frac{A_{2}}{ax_{1}} \qquad eqn - (4)$$

Where,

 C_x and C_y are concentrations of Rosuvastatin and niacin respectively $Q_m\!=\!A_2/A_i\!=\!absorbance$ of sample at 254nm/ absorbance of sample at262nm $Q_x\!=\!ax_2/ax_1\!=\!The$ absorptivity of Ros at 254nm/The absorptivity of Rosat 262nm.

 $Q_y=ay_2/ay_1=The$ absorptivity of Niaat 254nm/The absorptivity of Niaat 262nm.

Table 1: Linearity data of Rosuvastatin

Concentration in µg/ml	Concentration in gm/lit.	Absorbance at 241 nm
5	0.005	0.18755
10	0.010	0.39136
15	0.015	0.54402
20	0.020	0.73745
25	0.025	0.91797
30	0.030	1.127970
35	0.035	1.24234
40	0.040	1 36193

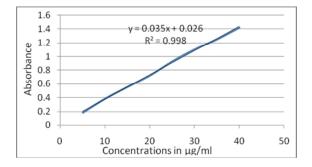


Figure 1: Linearity Graph Rosuvastatin at 254 nm

Concentration Concentration Absorbance at in gm/lit. in μg/ml 262 nm 0.005 0.14156 10 0.010 0.28682 15 0.015 0.407440.020 0.54525 25 0.025 0.67906

0.030

0.035

0.040

0.84407

0.88014

0.91786

30

35

40

Table 2: Linearity data of Niacin

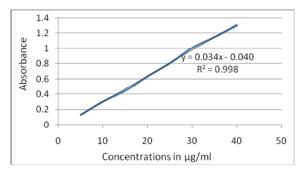


Figure 2: Linearity Graph Niacin at 262 nm

Table 3: Linear regression data for rosuvastatin and niacin

Parameters	Rosuvastatin	Niacin
Linearity range	5-40 μg/ml	5-40 μg/ml
R ² (Regressioncoefficient)	0.998	0.998
Slope	0.035	0.034
Intercept	0.026	0.040

Table 4: Results for precision study of tablet dosage form

Component	Concentration (µg/ml)	Mean*	Standard Deviation	Percentage Relative Standard Deviation	Standard Error
Rosuvastatin	10	98%	0.25	0.26	0.14
Niacin	10	102%	0.14	0.13	0.08

Table 5: Statistical validation of Accuracy (recovery test)

Component	Percentage	Mean*	Standard Deviation	Percentage Relative Standard Deviation	Standard Error
Rosuvastatin	50%	98.01	0.259	0.26	0.14
at 241nm	100%	100.2	0.58	0.579	0.3
	150%	99.13	0.68	0.68	0.3
Component	Percentage	Mean*	Standard	Percentage Relative Standard	Standard
•			Deviation	Deviation	Error
Niacin	50%	101.2	0.14	0.139	0.081
at 262nm	100%	103.3	0.47	0.457	0.27
	150%	102.1	0.14	0.138	0.081

Table 6: At iso-absorptive point (254nm)

Concentration in µg/ml	Concentration in gm/lit.	Absorbance at 254 nm	
5	0.005	0.15508	
10	0.010	0.46051	
15	0.015	0.68112	
20	0.020	0.92300	
25	0.025	1.14496	
30	0.030	1 40027	

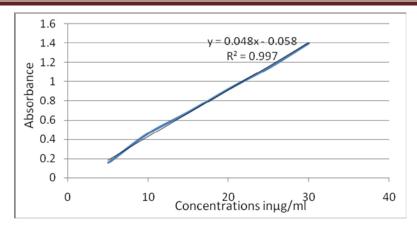
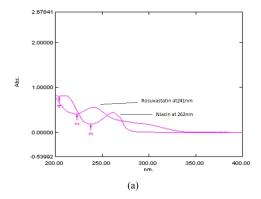


Figure 3: Linearity Graphat iso-absorptive point (254nm)

Table 7: Statistical validation of Accuracy (recovery test)

Component	Percentage	Mean*	Standard Deviation	Percentage Relative Standard Deviation	Standard Error
Rosuvastatin	50%	100.1	0.105	0.105	0.0608
at 254nm	100%	100.1	0.141	0.141	0.081
	150%	99.6	0.169	0.17	0.9
Component	Percentage	Mean*	Standard	Percentage Relative Standard	Standard
_			Deviation	Deviation	Error
Niacin	50%	100	0.124	0.124	0.072
at 262nm	100%	100.9	0.105	0.104	0.0608
	150%	100.8	0.38	0.38	0.2



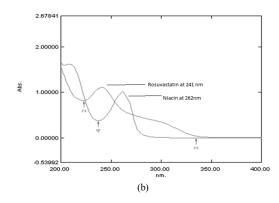


Figure 4: Overlain spectra of Rosuvastatin (241nm) and Niacin (262nm)

CONCLUSION

The proposed method is found to be simple, sensitive and reproducible and hence it can beused in routine analysis for simultaneous determination of Rosuvastatin and Niacin in bulk as well as in pharmaceutical preparation. Statistical analysis of the results has been carried out revealing linear, high accuracy and good precision.

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