



Research Article

DESIGN DEVELOPMENT AND EVALUATION OF TOPICAL MICROEMULSION

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ABSTRACT

A microemulsion based gel was designed for the topical and targeted delivery of sertaconazole nitrate for the treatment of superficial fungal infection. The microemulsion region was obtained using a ternary diagram, different ratio of oil and Smix were used. The microemulsion of sertaconazole containing 2% (w/w) of sertaconazole, 6.67% (w/w) of oil phase (Eugenol+Oleic acid 1:1), 60.18% (w/w) of surfactant mixture 2:1 ratio (Tween-80 and Transcutol-P) and 33.15% (w/w) with distilled water. The prepared microemulsion gel and commercial cream of sertaconazole were evaluated for in-vitro and ex-vivo studies. The highest drug retention was achieved with Tween 80 and Transcutol P (T₈₀TC45) when the optimized formulation was converted to a gel. The designed formulation MG2 was safe to be used over the skin as the PDI=0 when compared with commercial cream and MG1. The optimized formulation also posse's anti-inflammatory activity. The average zone of inhibition of MG2 was (23.19 ± 0.478) which was more than the commercial cream (15.34 ± 0.382) or MG1 (17.78 ± 0.715). *Candida albicans* which may be due to better permeation and retention effect of microemulsion gel 2. The MG2 was found to be stable after six month. The results obtained in this research from *in vitro* and *in vivo* data it can be concluded that the developed microemulsions have great potential for topical drug delivery in the treatment of inflammation and fungal infection.

Key words: Sertaconazole nitrate, microemulsion gels, skin retention, antifungal, anti-inflammatory effect

INTRODUCTION

Delivery of a drug via skin¹⁻² found to be attractive and proven to be very beneficial, as the systemic load of API is avoided and thus side effect are reduced as compared to others routes, drug applied topically avoids a number of parameters. Plasma levels typical for repeated administration of rapidly eliminated drug circumvent the first pass effect and decrease gastrointestinal side effects of a drug administered by the oral route. Local actions include actions on the stratum corneum, or within the dermis. Topical delivery³⁻⁴ has become an important means of drug delivery. Delivery of drugs to skin for systemic and local effect is called topical delivery. Topical delivery involves in the availability of drug molecules continuously from the surface, through its layers, and maintain a constant concentration within. Thus it's a valuable alternative to the conventional topical, oral and parenteral route of drug administration. Several topical therapeutic systems are being developed successfully and recently commercialized. The reason for selecting a skin, as the route of delivery of API, is mainly because of the fact that this method avoids the irritation to the GIT that can often occur, causing bleeding, etc. Additionally, in some instances administration through this route allows the drug to bypass the metabolism, allowing more of the drug's active ingredient to be utilized. Furthermore, a high drug concentration can be delivered to a particular diseased or affected area (e.g. bacterial or fungal infection). Ingredients selected must be tolerable to the patient and non-corrosive to the applied area. Absorption rate must be considered along with the total amount of drug delivered and the rate of elimination of active ingredient if found in the bloodstream.

Microemulsion could be an alternative carrier in topical drug delivery and as it has high Solubilization capability and nanometer size, it is believed that microemulsion will be a better candidate in delivering drug topically. Microemulsions composed of surfactant, water, and oil having co-surfactants provide better therapeutic action when compared to the traditional cream and lotions.

Chemically, Sertaconazole contains a benzothioephene ring which makes it unique from other imidazole antifungal. A benzothioephene ring is a sulfur analog of the indole ring found in the amino acid tryptophan. Tryptophan is found in the fungal membrane in addition to lipids such as ergosterol. The benzothioephene ring in Sertaconazole mimics tryptophan and increases the drugs ability to form pores in the fungal cell membrane. If the cell membrane is made sufficiently leaky by these pores the fungal cell will die.

MATERIALS AND METHODS

Sertaconazole nitrate was purchased from Hangzhou Holypharm Biotech Co. Ltd. (Zhejiang, China, Eugenol, Tween-80, propylene glycol was purchased from Sigma Aldrich Mumbai Transcutol P was gifted from gattefosse, India. All other chemicals used in the study were of analytical reagent grade.

Screening of excipients

Screening of excipients is most important criteria to find Sertaconazole solubility⁵⁻⁶ in different excipients such as oil, surfactants, and cosurfactants. Maximum solubility is to be fined in each component and with the help of ternary diagram microemulsion region is obtained. Smix has a vital in the

formulation as its presence makes the interfacial tension very low, and hence microemulsions formed spontaneously, with an average droplet diameter of 10-200 nm or smaller. The spectrophotometer was used at 260 nm for analysis of drug.

Drug Solubility

Drug solubility⁷ in number of oil, surfactants and co-surfactant (Oleic Acid, Eugenol, Olive oil, Captex 300, Captex 355, Ethyl oleate and IPM) surfactants (Labrasol, Tween 20, Tween 80 and Cremophor RH-40) and co-surfactants (Transcutol P, Capryol, PEG 400, Ethanol and Propylene glycol) were detected by adding an excess amount of active pharmaceutical ingredients (API) in 2ml of the selected components in 5 ml capped vials with cap or aluminum foil cap separately, mixture were vortexed and the mixture vials were kept at $37^{\circ}\text{C} \pm 5^{\circ}\text{C}$ in incubator shaker for 72 hours, later microemulsions were centrifuged at five thousand RPM for fifteen minutes. The supernatant was separated and filtered using $0.45\mu\text{m}$ membrane filter, different Excipients solubility is illustrated in the table 1 and figure 1 to 3. The API was detected in each component using spectrophotometer at (260nm).

Analytical Method

High-performance liquid chromatography and UV spectrophotometric method were developed and validated for the quantitative determination of the bulk sertaconazole nitrate⁸ and its micro emulsion formulation. For HPLC, LC GC Qualisil BDS C18 column (4.6×250 mm, $5\mu\text{m}$ particle size) with the mobile phase consisted of acetonitrile-water (65:35% v/v) and flow rate of 1.8 ml/min were used for the analysis. The sertaconazole nitrate peak is monitored at a wavelength of 260 nm; the retention time was 20.16 min. The method is considered reliable for the determination of sertaconazole nitrate. Nearly 99.6% of sertaconazole nitrate from microemulsion formulation were recovered by applying this method with RSD 0.18% (n=9).

Construction of pseudo -ternary phase diagram

According to solubility studies Oleic acid + Eugenol 1:1 ratio was chosen as the oil, Tween-80 (HLB value 15) selected as a surfactant and Transcutol P, Polyethylene glycol was selected as Cosurfactant, for aqueous phase water was used. Different Smix ratios 1:1, 1:2, 1:3, 2:1, 3:1, and 4:1. The ratio⁹⁻¹¹ was selected in different concentration. Firstly concentration of surfactant was increased as compared to Co-surfactant and in second condition concentration of Co-surfactant was increased as compared to surfactant, different ratio of oil and Smix were varied as 9.5:0.5, 9:1, 8.5:1.5, 8:2, 7.5:2.5, 7:3, 6.5:3.5, 6:4, 5.5:4.5, 5:5, 4.5:5.5, 4:6, 3.5:6.5, 3:7, 2.5:7.5, 2:8, 1.5:8.5, 1:9, 0.5:9.5 Were chosen so as to cover maximum ratio which will be important to define the ternary diagram. Aqueous titration method was deployed to develop phase diagram. Slow titration with water and constant stirring after each water addition, the tube was observed for clarity and stability. Point where solution became turbid marked as the end point. The quantity of the distilled water added was noted, the same process was repeated for all other surfactant/co-surfactant ratios. Those formulations¹² which remain stable after water titration and further addition of aqueous do not destabilize microemulsion. The result of preliminary trial batches of microemulsion presented in Table 5.6. (Oil phase 5-95% in each batch) A three component ternary diagram with each axis representing an oil phase, Smix, and water with fix mass ratio. The microemulsion area was drawn using Smix software.

Selection of microemulsion on the basis of stability studies

The optimized formulation was evaluated for following stability testing methods

Centrifugation

Remi Model R-8C Centrifuge instrument at 5000 rpm for 15 min to find the stability of formulation by analyzing¹³ separation of phase occurs or not. Formulations do not undergo phase separation were taken to next stability testing methods.

Thermal stability of microemulsion

Stability of optimized Formulations was detected by placing in 10 ml transparent borosil volumetric flask at three different temperatures i.e. 4, 25 and $45^{\circ}\text{C} \pm 1^{\circ}\text{C}$ in a temperature controlled oven or in an incubator for the duration of 48-72 hours. Samples were removed periodically for assessment to detect any physical changes like loss of coalescence, clarity, and turbidity etc.

Freeze-Thaw Method

The freeze-thaw methods¹⁴ were employed where temperature ranging from -4 to 40°C for the duration of twenty-four hours. Samples were periodically checked visually to find any physical changes like clarity loss, the presence of coalescence and turbidity etc.

Stability Study Microemulsions

Here, we use

- 'x' for unstable and '√' for stable after 24 hrs.
- 'x' separation of phase and '√' non-separation of phase after centrifugation.
- Freeze Thaw method √ – Pass, x - Fail

Clarity/Dispersibility test

The stability of microemulsion¹⁵⁻¹⁷ was assessed for clarity for infinite dilution. Dilutions were checked using XXII USP dissolution apparatus. Test formulations were transferred in 900 ml 0.1 HCl and distilled water respectively at $37 \pm 0.5^{\circ}\text{C}$. The aim of this research was to detect the best grade of formulation in reference to given table 4

% Transmittance¹⁸⁻¹⁹ was checked with respect to distilled water using spectrophotometer at 650 nm by dilution of 1.0 ml of the formulation with distilled water up to 100 ml. The microemulsion was examined for clarity by finding the transparency in term of Transmittance. Having water in as an external phase, %T value less than 98% suggest less clarity of microemulsion. Table 3

Preparation of microemulsions

The microemulsion which passed the test as described in Table 5 was used for further investigation.

According to ternary diagram sertaconazole, loaded microemulsion²⁰⁻²² was selected comprising of different component ratio. The microemulsion was prepared with reference to the area in the ternary diagrams, the Sertaconazole nitrate-loaded microemulsion was selected having different oil and Smix ratio. Sertaconazole nitrate (2% w/w) were dissolved in oil (oil phase was varied from 5% to 95%, and the drug was dissolved with the help of ultrasonication. (Oleic acid + Eugenol 1:1). The optimized quantity of surfactant (Tween 80)

and co-surfactant (Propylene glycol or Transcutol P) were added and vortexed for five minutes, the aqueous phase was added slowly with continued stirring, turbidity appearance is considered end point. Selected microemulsion formulation is given in table 6

Following parameters were employed for evaluation of Microemulsion.

Microscopic Evaluation

OLYMPUS microscope and optical microscope were employed to detect the homogeneity of on formulation.

Microemulsion droplet size analysis

The size and distribution of formulation were obtained by Malvern Zetasizer²³⁻²⁴ version 6.20 laser scattering principle is employed. Malvern instrument having laser light scattering zeta sizer with argon laser was employed for evaluating the size of globule in microemulsion and size distribution, at 90° angles and 25 °C scattering of light was monitored. The microemulsion size was obtained from the intensity, volume and bimodal distribution assuming particles to be spherical

Zeta Potential

It's an important parameter that provides an indication of the stability in colloidal systems and indicates charge present on the colloidal systems. Highly positive or highly negative²⁵⁻²⁶ charge on oil globules indicate higher stability because of the anticipated surface repulsion between similarly charged globules hence inhibiting aggregation of the colloidal oil globules

Refractive index & pH

Refractive index of optimized formulations was detected using an Abbe-type refractometer²⁷. To standardized, the instrument castor oil was used. It's a parameter in finding droplet size distribution of microemulsion as the droplet size measurement is done by light scattering observed at 90° angles. Benchtop pH Meter was employed to find the pH of the optimized formulation. pH meter was standardized with pH 4 and pH 7 buffers before use.

Conductivity Measurement

SIMTRONICS conductivity²⁸ meter having magnetic stirrer was used to find the conductivity²⁸ of formulation, having two platinum plates which are separated by a defined distance and having liquid between the platinum plates act as a conductor. It helps to determine the type of microemulsion and detect phase inversion phenomenon.

Viscosity

In the present study, the viscosity²⁹ of microemulsion and its gel formulation were detected using Brookfield Viscometer (LV DV-III+ Pro EXTRA) rheometer used to measure viscosity and shear stress at given shear rates. It consists of the sample holder, and water jacket, and spindle. The rheometer uses a calibrated spring to drive a spindle that is immersed in the test fluid. DV-III Ultra programmable rheometer is able to measure viscosity over an extremely large range of 0.1 to > 800 million cP.

TEM Analysis

Morphology of microemulsion was studied using TEM, TOPCON 002B used at 200 KV and of a 0.18 nm providing point to point resolution. Increasing magnification, Bright field imaging modes were used find the type and size³⁰⁻³¹ of the microemulsion. In order to perform the TEM an observation, the microemulsion was diluted with distilled water (1/100). A small drop of diluted microemulsion was deposited on the Copper holey film grid and observed by having a fixing agent and drying it in the filtered air.

Permeation, retention studies

Rat skin was obtained from already approved experiment (Reference No Med/IAEC/2012/136) Subharti University, Meerut to carry the permeation studies using the skin. Franz diffusion apparatus having an effective diffusion¹⁶⁻³² area of 3.14 cm² receptor volume was 20 ml were employed for the permeation study. The optimized skin was placed at 25° C for 30 minutes before conducting the experiment. The skin was washed with distilled water and skin was clamped on the Franz diffusion apparatus. The subcutaneous side should face up into the donor compartment and the dermal side should face the receptor compartment. 2% optimized formulations (ME1- ME8) was administrated on the subcutaneous side of individual skin samples. The upper part of the cell was covered with aluminum foil. The receptor chambers were filled with methanolic phosphate buffer 7.4 (30:70%, V/V). The receiver compartment was stirred at 100 rpm and 37±1 °C was maintained. The whole methanolic PB was replaced with new at an interval of thirty minutes until the skin was stabilized. Practically it was found that after 2.5 hours skin stabilization was achieved. When complete stabilization was achieved, Specified amount of formulation was placed into the donor compartment and sealed as to maintained occlusive conditions. Samples were withdrawn at regular intervals 0.5, 1, 2, 3, 4, 5, 6, 8, 10, 12, 20 and 24 hrs and filtered through membrane filter size 0.45µ and analyzed for drug content by HPLC. *Ex-vivo* Skin permeation profile of Microemulsions is given in Figure 5.

Optimized formulation selection

Formulation ME 2 and ME 6 have lowest release profile hence both the formulation ME 2 and ME 6 was converted to gel formulation (MG1 and MG2).

Preparation of gel of microemulsion

Carbomer 934 was used to make gel matrix³³. The polymer was swelled with a small amount of water for 24 h resulting in a solution of high viscosity. Sertaconazole microemulsion was added little by little to the viscous solution under constant stirring. The concentration of Carbomer 934 in MB gel was 1% (w/w). Cumulative drug release from Microemulsion MG1 and MG2 is described in Figure 6

Permeation data analysis

The drug permeated or retained³⁴⁻³⁷ through the skin (mg cm⁻²) using franz diffusion was calculated. Drug flux (permeation rate) at steady state (J_{ss}) was calculated by dividing the slope of the graph linear portion with the diffusion cell area (mg cm⁻²h⁻¹). K_p , Permeability coefficient was calculated by dividing J_{ss} by the initial concentration of the drug in the donor cell (cm⁻¹). Er Enhancement ratio was calculated by dividing J_{ss} of the

respective formulation by J_{ss} of the control formulation. The permeation profile is given in table 8.

Characterization Of Microemulsion Gel

Clarity Test

The clarity test employed to detect the stability of gel; it was detected by visual inspection under background which is black and white.

Satisfactory +
 Good ++
 Excellent (glassy) +++

Spreadability

Spreadability was determined using wooden block apparatus, which was provided by a pulley at one end. By this method, Spreadability³⁸⁻³⁹ was measured on the basis of "slip" and "drag". A ground glass slide was fixed on this block. An excess of gel (about 2 g) under study was placed on this ground slide. The gel was then sandwiched between the slides. A weight of 100 g was placed on the top of the two slides for 5 min to expel air and to provide a uniform film of the gel between the slides. Excess of the gel was scrapped off from the edges. The top plate was then subjected to a pull of 20 g weight with the help of a string attached to the hook and the time (in seconds) required by the top slide to cover a distance of 7.5 cm was noted.

$$S = M.L / T$$

Homogeneity

A little portion of the gel is checked by pressing in between the thumb and the index finger and the consistency of the gel is noticed (whether homogeneous or not, if their coarse particle appeared or detached on fingers).

Skin irritancy test

For this investigation, Wistar rat of any sex was used. The rats were obtained from the animal house of SV Subharti University, Meerut, U.P, India, Ref. No Med/IAEC/2014/378. Rats in the range of weighing 180-200 g were chosen for the study⁴⁰⁻⁴¹. The day before study hairs from the site of the study of the animal was removed with the help of hair clippers and scissors, complete hair removal should be done from 2 cm² area the portion was cleaned with surgical spirit. A 10 μ l of sample formulation gel was then applied the following day to the site if investigation.

Test Materials

Microemulsion formulation (MG1 & MG2) were selected to be tested against the control 2% Sertaconazole cream (SERACON, AS Life Science) The dose of each test material was taken 10 μ l.

Clinical Observations

Assessing the site where the formulation was applied was scored once daily at 1, 2, 3 and 4 days after microemulsion application in the form of MG1 & MG2. Reaction on skin at the application site scored as follows grading of skin reaction (Table 9,10)

Primary Dermal Irritation Index CALCULATION (PDI)

The PDI was calculated with the help of following formula and the result was predicted according to Figure 8-11

$$PDI = \text{Combined index for 1, 2, 3 and 4 days} / 4$$

Anti-inflammatory activity

Anti-inflammatory activity of MG1 and MG2 were compared with the marketed formulation. The study was carried out with the help of carrageenan⁴² that was used to induce paw edema as developed by (Winter *et al.*, 1962). in albino rats. Rat weighing 180-210 g overnight fasted with free water. Groups were divided into 2 groups of 2 animals each. Dorsal part of hair of animal was first trimmed and shaved 12 h before starting the experiments. The control animals were kept intact without any disturbance. The first batch (control) received carrageenan only without the drug. The second batch received an application of optimized formulation in a dose of 5 mg/kg on the shaved region of all animals (except control group) half an hour before subplantar mode of carrageenan. The animals were injected with 0.1 ml of carrageenan suspension (1%, w/v, in distilled water) in the right paw. Paw edema was obtained before carrageenan injection as well as after 1 to 6 h following the carrageenan injection using mercury displacement method. The % inhibition of edema volume was calculated as follows:

$$\% \text{ Inhibition} = 100 \times [1 - (A - x / B - y)]$$

Where A is paw volume after administration of carrageenan at time t,

X is paw volume before administration of carrageenan.

B is the mean paw volume of control rats after administration of carrageenan at time t

y is mean paw volume of control rats before administration of carrageenan.

Antifungal activity In vitro

Cup plate methods

The sterilized media was poured into Petri-plates^{43,44} of 100 mm size. For each formulation, three plates were prepared and kept for solidifying. One hole was bored in each plate with a stainless steel borer of 9mm diameter. The test solution was delivered with a micropipette into the holes. The volume of all the formulation to be tested was kept uniform (0.5 ml in each hole). The Petri dishes were left aseptically for an hour for diffusion^{45,46} of the drug solutions. The antifungal property of optimized formulation from (MG 1 and MG2) and the control 2% Sertaconazole marketed formulation was determined using *Candida albicans*. (ATCC 10231) as representative fungi, adopting the Petri plate method.

Stability Studies

Optimized formulations (ME 2 and ME 6) and (MG1 and MG2) were subjected to stability studies. Formulations were transferred in ampoules and placed in Stability chambers as described in Table 16. Samples were withdrawn at 0, 1, 3 and 6 months⁴⁷⁻⁴⁹ to evaluate their physical stabilities. The stability of optimized formulations was investigated for different parameters.

The stability study was performed as per ICH guideline conditions can be decided based on climatic condition of that particular zone. As per guideline, stability is carried out as per given parameters.

Table 1 solubility drug in different component

Component	Solubility (mg/ml)	Component	Solubility (mg/ml)
Eugenol	39.23 ± 0.22	Span 80	21.66 ± 0.57
Oleic acid	31.03 ± 1.527	Tween 80	37.33 ± 0.012
Oleic acid + Eugenol (1:1)	41.13 ± 0.44	Span 20	3.02 ± 1.645
Light Liquid Paraffin	9.33 ± 0.577	Tween 20	28.03 ± 0.605
Cardamom oil	17.13 ± 1.527	Propanol	23.7 ± 2.645
Peppermint oil	24.33 ± 2.516	Acconon CC-6	33.03 ± 0.79
Castor oil	10.33 ± 1.527	Isopropyl alcohol	17.33 ± 0.201
Cinnamon oil	28.66 ± 1.527	Cremophor RH-40	25.66 ± 1.081
Labrafac	28.33 ± 1.154	Transcutol P	37.02 ± 1.358
Capryol 90	21.66 ± 2.081	Propylene glycol	35.66 ± 1.969
Captex 355	23.33 ± 2.309	Polyethylene glycol	26.17 ± 1.732
Isopropyl myristate	20.66 ± 2.081		

Table 2 Smix ratio used for Microemulsion formulation

S.No	Surfactant volume	Co surfactant volume	Smix ratio
1	50	50	1:1
2	33.3	66.7	0.5:1 or 1:2
3	25	75	1:3
4	66.7	33.3	2:1 or 1:0.5
5	75	25	3:1
6	80	20	4:1

Table 3 Microemulsion stability result

Smix Ratio (S: Cs)	Code	% V/V			Observation according to Thermodynamic Stability			Inference
		oil	Smix	Aqueous	Stable at room Temp.	Centrifuge	Freeze Thaw	
1:1	Oil: Tween 80: propylene glycol							
	T ₈₀ PG 2	9.12	53.65	37.23	√	√	√	Passed
	T ₈₀ PG 3	11.97	46.19	41.84	√	√	√	Passed
	T ₈₀ PG 4	13.03	43.81	43.16	√	√	√	Passed
	T ₈₀ PG 5	15.27	38.5	46.23	√	√	√	Passed
1:2	T ₈₀ PG 17	9.95	64.93	25.12	√	√	√	Passed
	T ₈₀ PG 19	12.43	53.97	33.6	√	√	√	Passed
	T ₈₀ PG 20	12.89	50.84	36.27	√	√	√	Passed
	T ₈₀ PG 23	18.23	30.66	51.11	√	√	√	Passed
1:3	T ₈₀ PG 34	10.43	16.23	73.34	√	√	√	Passed
2:1	T ₈₀ PG 43	11.26	67.38	21.36	√	√	√	Passed
	T ₈₀ PG 44	14.32	63.79	21.89	√	√	√	Passed
	T ₈₀ PG 45	16.12	61.15	22.73	√	√	√	Passed
	T ₈₀ PG 46	17.33	55.48	27.19	√	√	√	Passed
3:1	T ₈₀ PG 58	11.11	55.55	33.34	√	√	√	Passed
	T ₈₀ PG 59	10.25	51.28	38.47	√	√	√	Passed
	T ₈₀ PG 61	13.33	53.33	33.33	√	√	√	Passed
4:1	T ₈₀ PG 72	7.28	38.14	54.58	√	√	√	Passed
	T ₈₀ PG 73	7.87	36.5	56.23	√	√	√	Passed
	T ₈₀ PG 74	8.43	32.89	58.68	√	√	√	Passed
	T ₈₀ PG 75	11.2	31.02	57.01	√	√	√	Passed
1:1	Oil: Tween 80: Transcutol P							
	T ₈₀ TC 1	10.76	48.06	41.18	√	√	√	Passed
	T ₈₀ TC2	10.93	39.87	49.2	√	√	√	Passed
	T ₈₀ TC3	9.09	21.21	69.70	√	√	√	Passed
1:2	T ₈₀ TC18	5.88	52.94	41.18	√	√	√	Passed
	T ₈₀ TC19	7.22	46.4	46.38	√	√	√	Passed
	T ₈₀ TC20	9.52	38.10	52.38	√	√	√	Passed
1:3	T ₈₀ TC31	6.25	56.25	37.50	√	√	√	Passed
	T ₈₀ TC32	7.22	51.7	41.08	√	√	√	Passed
	T ₈₀ TC33	10.53	42.11	47.37	√	√	√	Passed
2:1	T ₈₀ TC45	6.67	60.18	33.15	√	√	√	Passed
	T ₈₀ TC46	7.91	55.07	37.02	√	√	√	Passed
	T ₈₀ TC47	11.76	47.06	41.18	√	√	√	Passed
	T ₈₀ TC48	13.64	31.82	54.55	√	√	√	Passed
	T ₈₀ TC49	14.72	28.25	57.03	√	√	√	Passed
	T ₈₀ TC50	16.00	24.00	60.00	√	√	√	Passed
3:1	T ₈₀ TC58	7.94	71.43	20.63	√	√	√	Passed
	T ₈₀ TC59	9.09	36.36	36.39	√	√	√	Passed
4:1	T ₈₀ TC71	7.03	66.4	26.57	√	√	√	Passed
	T ₈₀ TC72	10.4	53.21	36.39	√	√	√	Passed
	T ₈₀ TC73	11.27	47.06	41.67	√	√	√	Passed

Table 4 different grade of the microemulsion

S.No	Observation	Grade
1	Forming rapidly within one minute. Microemulsion is clear to slightly bluish	A
2	Forming rapidly little bit less clear and bluish color	B
3	Fine milky type emulsion	C
4	Emulsion grayish with slightly oily in appearance	D

Table 5 Clarity/Dispensability test, Transmittance of Microemulsion formulations

Batch no.	% of Oil	% of Smix	Dispensability tests in distilled water and 0.1 N HCl		Appearance after 100 times Dilution	*% T at 650 nm	**% T at 650 nm (after 100 times Dilution)	Inference
			Water	0.1NHCL				
T ₈₀ PG 17	9.95	64.93	A	A	Clear	99.74±0.3	99.21 ± 0.10	Pass
T ₈₀ PG 43	11.26	67.38	A	A	Clear	99.73 ± 0.2	99.13 ± 0.62	Pass
T ₈₀ PG 44	14.32	63.79	A	A	Clear	99.67 ± 0.27	99.03 ± 0.1	Pass
T ₈₀ PG 59	10.25	51.28	A	A	Clear	99.71±0.21	99.01±0.23	Pass
T ₂₀ TC 1	10.76	48.06	A	A	Clear	99.64±0.23	99.02±0.16	Pass
T ₂₀ TC45	6.67	60.18	A	A	Clear	99.71 ± 17	99.16± 0.2	Pass
T ₂₀ TC46	7.91	55.07	A	A	Clear	99.66±03	99.02±0.17	Pass
T ₂₀ TC71	7.03	66.4	A	A	Clear	99.79±0.27	99.23±0.21	Pass

A- Grade A microemulsion

Table 6 Selected Microemulsion Formulations (with 2% Sertaconazole)

Code	Selected Microemulsion composition					
	%wt/wt				Oil/S _{mix} ratio	S _{mix} ratio
	Drug	Oil	S _{mix}	Water		
ME1	2	9.03	63.09	27.88	1:9	1:2
ME2	2	10.67	65.73	23.6	2:8	2:1
ME3	2	14.1	62.7	23.2	3:7	2:1
ME4	2	11.06	49	39.94	2:8	3:1
ME5	2	9.87	48.62	41.51	1:9	1:1
ME6	2	7.18	59.03	33.79	1:9	2:1
ME7	2	8.29	54.19	37.52	2:8	2:1
ME8	2	8.7	63.9	27.4	1:9	4:1

Table 7 refractive index of placebo and drug loaded microemulsion

S No	Code	Refractive Index ± SD	
		Placebo formulation	Sertaconazole-loaded formulation
1.	ME 1	1.401 ± 0.001	1.411 ± 0.005
2.	ME 2	1.371 ± 0.005	1.363 ± 0.002
3.	ME 3	1.353 ± 0.003	1.391 ± 0.004
4.	ME 4	1.361 ± 0.006	1.381 ± 0.003
5.	ME 5	1.303 ± 0.003	1.321 ± 0.002
6.	ME 6	1.319 ± 0.003	1.407 ± 0.001
7.	ME 7	1.321 ± 0.004	1.357 ± 0.005
8.	ME 8	1.309 ± 0.005	1.402 ± 0.003

Table 8 Permeation data analysis

Formulation	J _{ss} (mg cm ⁻² h ⁻¹)	K _p x 10 ⁻² (cm h ⁻¹)	Er
ME 1	0.0672±0.010	0.0034±0.001	3.0685
ME 2	0.0429±0.020	0.0021±0.001	1.9589
ME 3	0.0764±0.008	0.0038±0.0009	3.4886
ME 4	0.0675±0.022	0.0034±0.0009	3.0822
ME 5	0.0739±0.029	0.0037±0.0008	3.3744
ME 6	0.0388±0.004	0.0019±0.0014	1.7717
ME 7	0.0591±0.018	0.0030±0.0014	2.6986
ME 8	0.0780±0.004	0.0039±0.0013	3.5616
MG 1	0.0348±0.016	0.0017±0.0007	1.5890
MG2	0.0326±0.003	0.0016±0.0011	1.4886
Marketed cream	0.0219±0.0008	0.005±0.006	--

Marketed cream was used as a control.

J_{ss} – steady state flux, K_p – permeability coefficient, Er – enhancement ratio.

Table 9 Grading reaction of skin (Erythema and Eschar Formation)

No erythema	0
Very slight erythema (barely perceptible)	1
Well defined erythema	2
Moderate to severe erythema	3
Severe erythema (beef redness) to eschar formation preventing grading of erythema	4

Table 10 Skin reactions grading (Oedema Formation)

No oedema	0
Very slight oedema (barely perceptible)	1
Slight oedema (edges of area well defined by definite raising)	2
Moderate oedema (raised approximately 1 mm)	3
Severe oedema (raised more than 1 mm and extending beyond area of exposure)	4

Table 11 Evaluation PDI (primary dermal index)

Evaluations	Score
Non Irritant	0.0
Negligible Irritant	0.1-0.4
Slight Irritant	0.41-1.9
Moderate Irritant	2.0 - 4.9
Severe Irritant	5.0 - 8.0

Table 12 Summary of observed irritation of skin scores of 2 % Sertaconazole (SERACON) Control

Animal no.	Sex	Time Period after 2 % Sertaconazole (SERACON) marketed preparation (Control) in days							
		1		2		3		4	
		Ery.	Oed.	Ery.	Oed.	Ery.	Oed.	Ery.	Oed.
1.	M	1	0.0	0.0	0.0	0.0	0.0	0.0	0.0
2.	M	1	0.0	0.0	0.0	0.0	0.0	0.0	0.0
3.	M	1	0.0	0.0	0.0	0.0	0.0	0.0	0.0
Total		3	0	0.0	0.0	0.0	0.0	0.0	0.0
Mean		1	0.00	0	0.00	0.0	0.00	0.0	0.00
Combined index		0.5		0.0		0.0		0.0	
PDI		0.12							

* (Ery. = Erythema; Oed. = Oedema)

Table 13 Summary of observed primary skin irritation scores of MG1

Animal no.	Sex	Time Period after MG 1 (in days)							
		1		2		3		4	
		Ery.	Oed.	Ery.	Oed.	Ery.	Oed.	Ery.	Oed.
1	M	1	0.0	1	0.0	0	0.0	0	0.0
2	M	1	0.0	0	0.0	0	0.0	0	0.0
3	M	1	0.0	1	0.0	1	0.0	0	0.0
Total		3	0.0	0.66	0.0	0.33	0.0	0	0.0
Mean		1	0.00	0.33	0.00	0.16	0.00	0	0.00
Combined index		0.5		0.16		0.08		0	
PDI		0.185							

* (Ery. = Erythema; Oed. = Oedema)

Table 14 Summary of observed primary skin irritation scores of MG2

Animal no.	Sex	Time Period after MG 2 (in days)							
		1		2		3		4	
		Ery.	Oed.	Ery.	Oed.	Ery.	Oed.	Ery.	Oed.
1.	M	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
2.	M	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
3.	M	0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
Total		0.0	0.0	0.0	0.0	0.0	0.0	0.0	0.0
Mean		0.0	0.00	0.00	0.0	0.0	0.0	0.0	0.0
Combined index		0.0		0.0		0.0		0.00	
PDI		0.00 (Zero)							

* (Ery. = Erythema; Oed. = Oedema)

Table 15 Anti-inflammatory activity of MG1, MG2 and Marketed formulation

Sl.No.	Body weight (gm)	Treatment	Dose	Paw Volume (ml) as measured by Mercury displacement																					
				0 min.		15 min.		30 min.		60 min.		90 min.		120 min.		150 min.		180 min.		210 min.		240 min.			
FOR CONTROL				R	L	R	L	R	L	R	L	R	L	R	L	R	L	R	L	R	L				
1	190 to 200	Control	0.1	0.15	0.13	0.15	0.25	0.15	0.24	0.15	0.31	0.15	0.31	0.15	0.34	0.15	0.36	0.15	0.34	0.15	0.36	0.15	0.37		
2				0.25	0.17	0.25	0.27	0.25	0.32	0.25	0.32	0.25	0.33	0.25	0.34	0.25	0.36	0.25	0.37	0.25	0.36	0.25	0.36		
3				0.13	0.21	0.13	0.29	0.13	0.31	0.13	0.32	0.13	0.35	0.13	0.14	0.13	0.35	0.13	0.36	0.13	0.35	0.13	0.35		
4				0.27	0.29	0.27	0.35	0.27	0.37	0.27	0.37	0.27	0.37	0.27	0.37	0.27	0.26	0.27	0.37	0.27	0.37	0.27	0.37	0.27	0.36
Mean				0.20	0.20	0.20	0.29	0.20	0.31	0.20	0.33	0.20	0.34	0.20	0.36	0.20	0.36	0.20	0.36	0.20	0.36	0.20	0.36		
% Increase in oedema				0%		45%		55%		65%		70%		80%		80%		80%		80%		80%			
FOR TREATMENT (market formulation)																									
1	190 to 200	Marketed formulation	2%	0.25	0.25	0.25	0.28	0.25	0.29	0.25	0.3	0.25	0.23	0.25	0.32	0.25	0.32	0.25	0.33	0.25	0.35	0.25	0.35		
2				0.2	0.2	0.2	0.33	0.2	0.34	0.2	0.33	0.2	0.33	0.2	0.35	0.2	0.35	0.2	0.34	0.2	0.34	0.2	0.34	0.2	0.34
3				0.23	0.23	0.23	0.36	0.23	0.36	0.23	0.38	0.23	0.37	0.23	0.37	0.23	0.38	0.23	0.4	0.23	0.41	0.23	0.41	0.23	0.41
4				0.24	0.24	0.24	0.27	0.24	0.29	0.24	0.31	0.24	0.39	0.24	0.32	0.24	0.39	0.24	0.41	0.24	0.42	0.24	0.42	0.24	0.42
Mean				0.23	0.23	0.23	0.31	0.23	0.32	0.23	0.33	0.23	0.33	0.23	0.34	0.23	0.36	0.23	0.37	0.23	0.38	0.23	0.39		
% Increase in oedema (for Treatment)				0%		34.78%		39.13%		43.48%		43.47		43.48%		56.52%		60.86%		65.21%		69.56%			
% Anti-inflammatory effect				0%		22.70%		28.85%		33.10%		37.88		31.67%		29.34%		23.91%		18.48%		13.04%			
FOR TREATMENT (MG1)																									
1	190 to 200	MG1	2%	0.24	0.24	0.24	0.23	0.24	0.24	0.24	0.25	0.24	0.27	0.24	0.24	0.24	0.25	0.24	0.26	0.24	0.27	0.24	0.28		
2				0.17	0.17	0.17	0.27	0.17	0.29	0.17	0.29	0.17	0.31	0.17	0.3	0.17	0.33	0.17	0.35	0.17	0.35	0.17	0.35		
3				0.25	0.25	0.25	0.31	0.25	0.32	0.25	0.33	0.25	0.36	0.25	0.34	0.25	0.36	0.25	0.34	0.25	0.37	0.25	0.37	0.25	0.37
4				0.22	0.22	0.22	0.35	0.22	0.35	0.22	0.37	0.22	0.38	0.22	0.38	0.22	0.38	0.22	0.38	0.22	0.37	0.22	0.37	0.22	0.38
Mean				0.22	0.22	0.22	0.29	0.22	0.30	0.22	0.31	0.22	0.3	0.22	0.31	0.22	0.33	0.22	0.33	0.22	0.34	0.22	0.35		
% Increase in oedema (for Treatment)				0%		31.81%		36.36%		40.90%		36.36%		45.45%		50%		50%		54.54%		59.09%			
% Anti-inflammatory effect				0%		29.29%		33.88%		37.06%		48.05%		41.55%		37.5%		37.5%		31.81%		26.13%			
FOR TREATMENT (MG2)																									
1	190 to 200	MG2	2%	0.24	0.25	0.24	0.25	0.24	0.27	0.24	0.27	0.24	0.25	0.24	0.25	0.24	0.25	0.24	0.27	0.24	0.27	0.24	0.28		
2				0.19	0.2	0.19	0.29	0.19	0.32	0.19	0.31	0.19	0.31	0.19	0.31	0.19	0.33	0.19	0.36	0.19	0.35	0.19	0.37		
3				0.27	0.28	0.27	0.33	0.27	0.34	0.27	0.35	0.27	0.35	0.27	0.35	0.27	0.36	0.27	0.35	0.27	0.37	0.27	0.37	0.27	0.37
4				0.26	0.23	0.26	0.37	0.26	0.35	0.26	0.39	0.26	0.37	0.26	0.37	0.26	0.38	0.26	0.38	0.26	0.38	0.26	0.37	0.26	0.38
Mean				0.24	0.24	0.24	0.31	0.24	0.32	0.24	0.33	0.24	0.32	0.24	0.32	0.24	0.33	0.24	0.34	0.24	0.34	0.24	0.35		
% Increase in oedema (for Treatment)				0%		29.16%		33.33%		37.5%		33.33%		33.33%		37.5%		41.66%		41.66%		45.83%			
% Anti-inflammatory effect				0%		35.18%		39.39%		42.30%		52.38%		52.38%		53.12%		47.91%		47.91%		42.70%			

Table 16 Stability studies as per ICH guidelines

(a)	30°C/65% RH	6 months	Intermediate stability
(b)	40°C/75% RH	6 months	Accelerated study

Table 17 Stability of Microemulsion ME 2

Formulation	Period In Month	30°C/65% RH					40 ±°C/75% RH				
		Droplet size	Zeta potential	Viscosity (mPa.s)	pH	Drug content	Droplet size	Zeta potential	Viscosity (mPa.s)	pH	Drug content
ME2	0 Month	33.21	-33.27	119.14±0.6	5.86 ±0.10	99.01 ± 0.2	33.21	-33.27	119.14±0.6	5.86 ±0.10	99.01 ± 0.2
	1 Month	34.27	-35.08	118.37±0.19	5.61 ±0.17	95.67 ± 0.16	34.97	-35.61	117.03±0.36	5.47 ± 0.02	94.41 ± 0.11
	3 Month	35.02	-31.71	117.33±0.34	5.41 ±0.04	93.55 ± 0.19	36.18	-31.07	114.75±0.12	5.61 ± 0.02	91.01 ± 0.02
	6 Month	36.07	-30.91	117.71±0.41	5.59 ±0.32	88.19 ± 0.22	35.04	-30.21	108.87±0.80	5.77 ± 0.01	86.12 ± 0.3

Table 18 Stability of Microemulsion ME 6

Formulation	Period In Month	30°C/65% RH					40 ±°C/75% RH				
		Droplet size	Zeta potential	Viscosity (mPa.s)	pH	Drug content	Droplet size	Zeta potential	Viscosity (mPa.s)	pH	Drug content
ME6	0 Month	41.29	-23.9	93.76±0.73	5.13 ±0.05	99.11±0.41	41.29	-23.9	93.76±0.73	5.13 ±0.05	99.11±0.41
	1 Month	41.97	-20.07	91.96±0.41	5.36 ±0.05	98.02 ± 0.14	42.19	-24.97	90.12±0.10	5.39 ± 0.02	98.10 ± 0.10
	3 Month	43.07	-19.01	91.13±1.19	5.55 ±0.04	96.65 ± 0.41	43.66	-23.99	87.93±1.04	5.41 ± 0.02	96.57 ± 0.19
	6 Month	44.6	-21.77	90.53±0.92	5.43 ±0.07	95.2 ± 0.23	45.67	-25.07	84.76±0.40	5.67 ± 0.01	94.10 ± 0.2

Table 19 Stability of Microemulsions Gel 1 (MG 1)

Formulation	Period In Month	30°C/65% RH					40 ±°C/75% RH				
		Droplet size	Zeta potential	Viscosity Pa.s	pH	Drug content	Droplet size	Zeta potential	Viscosity Pa.s	pH	Drug content
MG1	0 Month	39.19	-33.17	63.07±0.32	6.53 ±0.07	99.01 ± 0.2	39.19	-33.17	63.07±0.32	6.53 ±0.07	99.01 ± 0.2
	1 Month	41.32	-33.78	62.27±0.41	6.17 ±0.012	95.67 ± 0.16	42.41	-32.25	61.43±0.10	6.17 ± 0.02	93.01 ± 0.11
	3 Month	42.61	-31.53	60.73±0.55	6.03 ±0.02	90.55 ± 0.19	44.27	-30.71	60.03±0.78	6.11 ± 0.02	89.01 ± 0.02
	6 Month	44.03	-34.73	60.06±0.17	5.7 ±0.032	87.19 ± 0.22	46.09	-34.11	58.7.7±0.2	6.03 ± 0.01	84.12 ± 0.3

Table 20 Stability of Microemulsions Gel 2 (MG 2)

Formulation	Period In Month	30°C/65% RH					40 ±°C/75% RH				
		Droplet size	Zeta potential	Viscosity Pa.s	pH	Drug content	Droplet size	Zeta potential	Viscosity Pa.s	pH	Drug content
MG2	0 Month	43.06	-24.90	51.03±0.047	6.79 ±0.05	99.07 ± 0.26	43.06	-24.90	51.03±0.047	6.79 ±0.05	99.07 ± 0.26
	1 Month	44.19	-23.30	50.5±0.98	6.15±0.005	99.02 ± 0.14	45.16	-23.03	50.11±1.82	6.22 ± 0.02	98.10 ± 0.10
	3 Month	47.15	-21.68	49.07±0.25	5.91 ±0.04	98.65 ± 0.41	47.18	-25.70	49.77±0.02	5.91 ± 0.02	97.57 ± 0.19
	6 Month	48.5	-20.11	49.1±0.43	5.88 ±0.057	97.2 ± 0.23	51.62	-22.81	49.13±0.14	5.77 ± 0.01	96.10 ± 0.2

Table 21 Droplet size, Polydispersity, Refractive index, pH, Viscosity and Zeta potential of selected microemulsion formulations

S. No.	Code	Droplet size (nm)	Polydispersity	Mean Viscosity (mPa. s) ± SD	Zeta Potential (mV)	pH	Conductivity (µS/cm) ± S.D
1	ME1	10.11	0.163	187.02 ± 0.2	-0.258	5.21 ± 0.05	121 ± 1.5
2	ME2	33.21	0.152	119.5 ± 0.6	-33.27	5.86 ± 0.10	142 ± 2.8
3	ME3	10.19	0.174	327.0 ± 0.7	-0.212	5.73 ± 0.03	187 ± 2.3
4	ME4	9.236	0.128	121.02 ± 0.8	-2.02	5.57 ± 0.15	133 ± 4.5
5	ME5	15.23	0.175	216.3 ± 0.2	-6.34	5.69 ± 0.15	194 ± 3.2
6	ME6	41.29	0.134	93.2 ± 0.2	-23.9	5.13 ± 0.05	157 ± 4.1
7	ME7	23.17	0.335	243.4 ± 1.2	-1.92	5.09 ± 0.6	143 ± 1.5
8	ME8	17.43	0.143	257.6 ± 1.03	-0.214	5.03 ± 0.08	172 ± 5.7

Table 22 characterization of microemulsion gel

Formulation	Droplet size	Zeta potential	pH	Spreadability (g/cm/sec)	Viscosity (Pa.s)	Refractive index
MG1	37.19	-33.17	6.53 ± 0.07	15.48 ± 0.64	63.07 ± 0.328	1.351 ± 0.006
MG2	43.06	-24.9	6.79 ± 0.05	14.18 ± 0.15	51.03 ± 0.047	1.381 ± 0.001

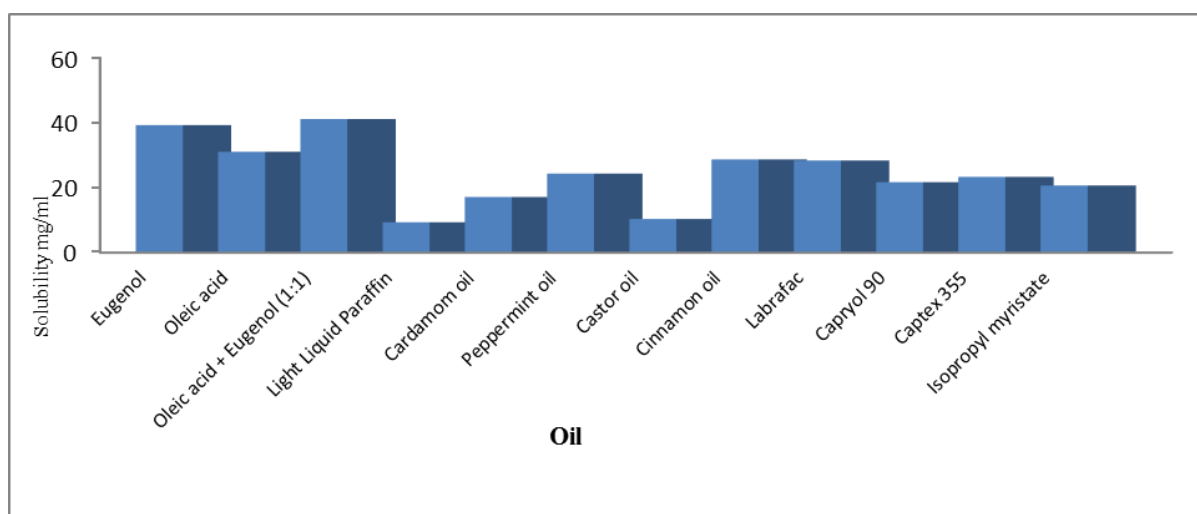


Figure 1 Sertaconazole solubility in different oil

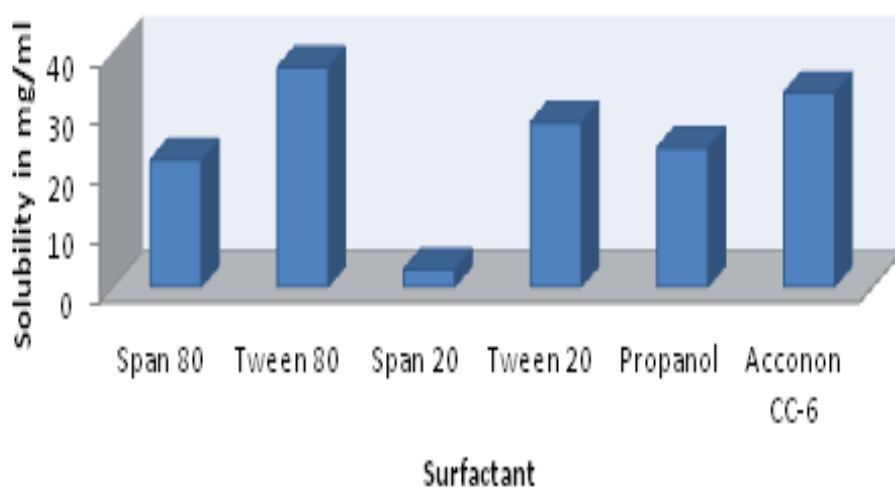


Figure 2 Sertaconazole solubility in different surfactants

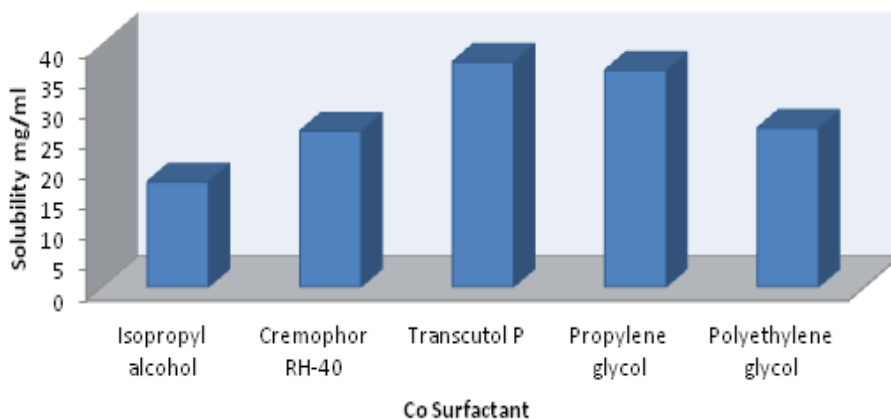
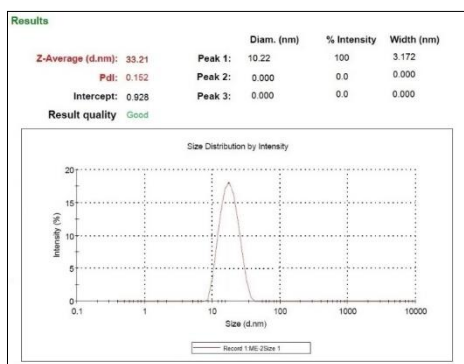
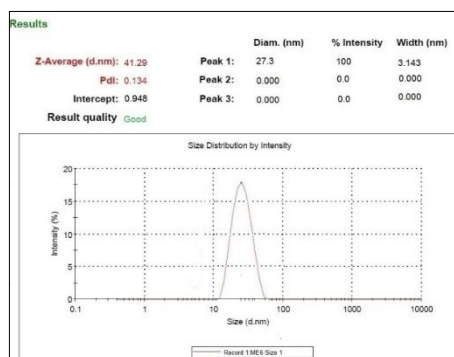


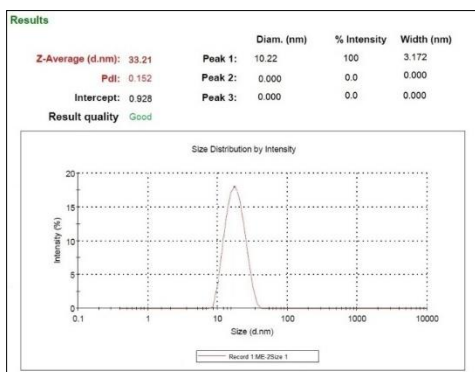
Figure 3 Sertaconazole solubility in Co Surfactants



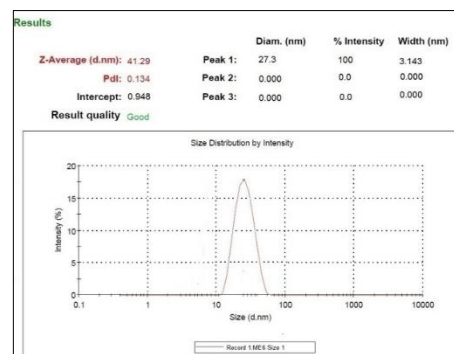
(a) Size distribution ME2



(b) Size distribution ME6



(c) Zeta potential report ME2



(d) Zeta potential Report ME6

Figure 4 Size distribution and Zeta potential study of microemulsion

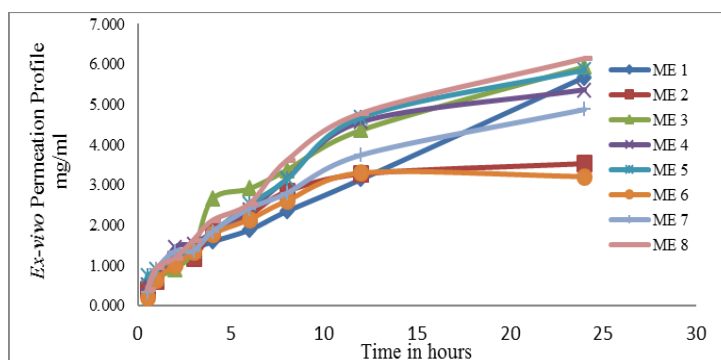


Figure 5 Ex-vivo Skin permeation, retention profile of Microemulsion

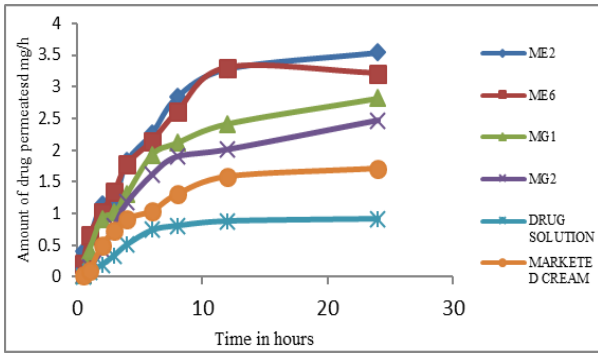


Figure 6 Comparative skin permeation profile of Sertaconazole Nitrate from ME2, ME6, MG1, MG2, drug solution and marketed cream

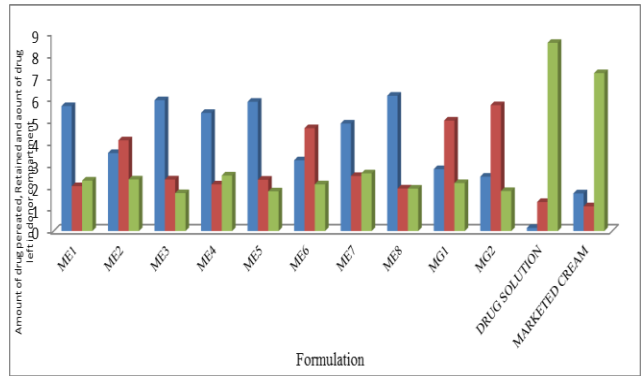


Figure 7 Ex Vivo permeation / retention study



Figure 8 MG2 application day 1



Figure 9 MG2 application day 2



Figure 10 MG2 application day 3



Figure 11 MG2 application day 4

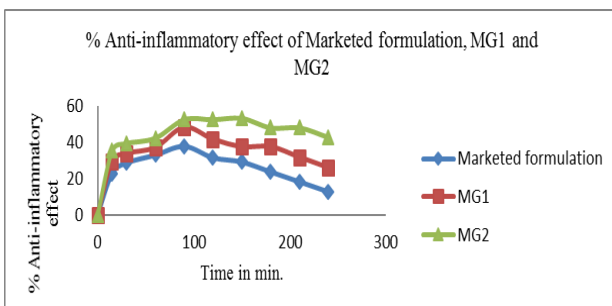


Figure 12 Anti-inflammatory activity of MG1, MG2 and Marketed formulation

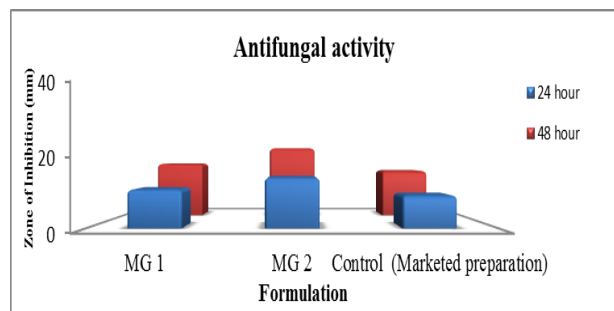


Figure 13 Antifungal activity data

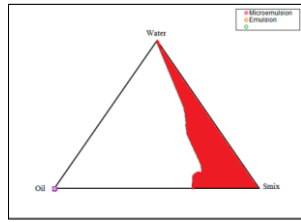


Figure 14 Ternary diagrams of ratio (1:1)

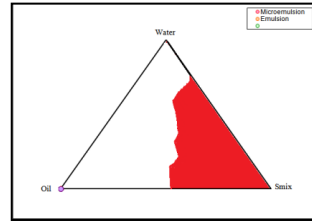


Figure 17 Ternary diagrams of ratio (2:1)

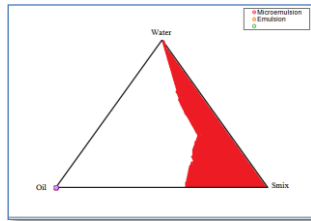


Figure 15 Ternary diagrams of ratio (1:2)

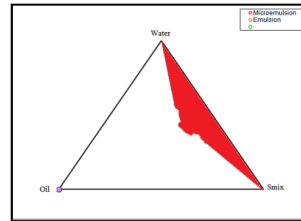


Figure 18 Ternary diagrams of ratio (3:1)

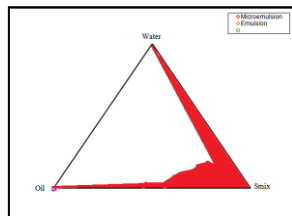


Figure 16 Ternary diagrams of ratio (1:3)

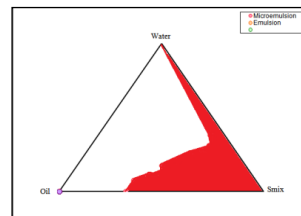


Figure 19 Ternary diagrams of ratio (4:1)

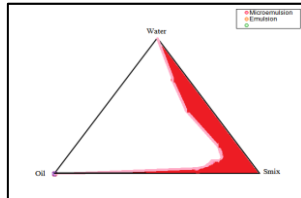


Figure 20 Ternary diagrams of ratio (1:1)

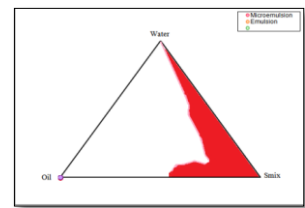


Figure 23 Ternary diagrams of ratio (2:1)

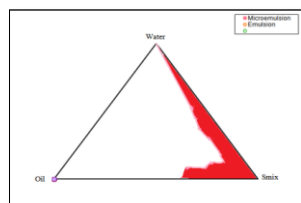


Figure 21 Ternary diagrams of ratio (1:2)

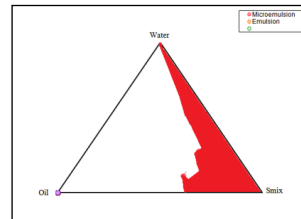


Figure 24 Ternary diagrams of ratio (3:1)

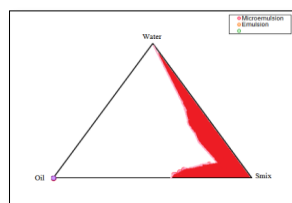


Figure 22 Ternary diagrams of ratio (1:3)

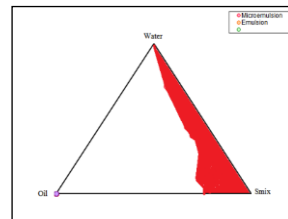


Figure 25 Ternary diagrams of ratio (4:1)

RESULT AND DISCUSSION

The physicochemical properties of sertaconazole suggest that it has good potential for topical as well as targeted drug delivery. The important criterion for selection of materials for the microemulsion formulation development is that the components are pharmaceutically acceptable, nonirritant and sensitizing to the skin and fall under the GRAS (Generally Regarded as Safe) category. Non-ionic surfactants are less toxic than ionic surfactants. The higher solubility of the drug in the oil phase is important for microemulsion to maintain the drug in solubilized form. The right blend of low and high hydrophilic lipophilic balance (HLB) surfactants leads to the formation of a stable microemulsion formulation. In this research, we selected Tween 80 as a surfactant having the HLB value 15. Transient negative interfacial tension and the fluid interfacial film are rarely achieved by the use of a single surfactant, usually necessitating the addition of a co-surfactant. The presence of co-surfactant decreases the bending stress of the interface and allows the interfacial film sufficient flexibility to take up different curvatures required to form a microemulsion over a wide range of compositions. Thus, two co-surfactants were selected for the study Propylene glycol HLB 2.5 and Transcutol-P with the HLB value of 4.2. Therefore, the aim of the present study was to develop and evaluate thermodynamically stable o/w microemulsion of sertaconazole for topical drug delivery. This microemulsion were prepared by using a combination of Eugenol + oleic acid as oil phase, Tween 80 as a surfactant, Propylene glycol and Transcutol P as co surfactant.

The most important criterion for the screening of components is the solubility of a poorly soluble drug in oil, surfactants, and cosurfactant. Since the aim of this study is to develop a topical formulation, it is important to determine drug solubility in oils, surfactants, and cosurfactant. The solubility of sertaconazole was found to be highest in Eugenol + oleic acid (1:1) 41.13 ± 0.44 , maximum solubility in cosurfactant was found in Tween 80 37.33 ± 0.012 , two co surfactant were used Propylene glycol solubility 35.66 ± 1.969 and Transcutol P solubility 37.02 ± 1.358 .

Pseudo-ternary phase diagram

Care was taken to ensure that observations were not made on metastable systems; although the free energy required to form an emulsion is very low, the formation is thermodynamically stable. The relationship between the phase behavior of a mixture and its composition can be captured with the aid of a phase diagram. Pseudo-ternary phase diagrams were constructed separately for each Smix ratio (Figure. 1), so that o/w microemulsion regions could be identified and microemulsion formulations could be optimized.

As two co surfactant were used for the formulation of o/w microemulsion such as Tween 80: propylene glycol and Tween 80 Transcutol P.

oil, smIx (Tween 80: Propylene glycol) & Water

In Figure. 14, Smix with a ratio 1:1 showed small microemulsion area. O/w microemulsion region was found towards the Smix rich apex, there was the formation of large emulsion region. In Figure. 15 Smix ratio 1:2 there was the formation of large microemulsion region and less emulsion region. When cosurfactant was added along with surfactant, the interfacial film became more fluid and no liquid crystalline area was found in the phase diagram. A large o/w microemulsion area was observed. In Figure 16 Smix ratio 1:3, microemulsion region was observed along both oil and water apex. Less

microemulsion region was observed and more emulsion region was formed and microemulsion was less stable. Smix ratio 2:1 Figure 17 has large microemulsion area this may be due to further reduction of the interfacial tension, increasing the fluidity of the interface, thereby increasing the entropy of the system. There may be greater penetration of the oil phase in the hydrophobic region of the surfactant monomers. As we further increased surfactant concentration in Smix to 3:1 Figure. 18, the microemulsion region decreased as compared to 2:1 and it was confined in between Smix and oil region resulting in the formation of large emulsion area and less stable formulation. When the Smix ratio of 4:1 was studied Figure. 19, the area of microemulsion increased but result in the formation of the less stable microemulsion.

Oil, Smix (Tween 80: Transcutol P) & Water

In Figure. 20, Smix ratio 1:1 showed narrow o/w microemulsion area and a large emulsion region was found. further in Figure 21 and 22 Smix ratio 1:2 and 1:3 same microemulsion region was obtained, the microemulsion obtained was more stable as compared to 1:1 ratio. Figure 23 Smix ratios 2:1 has microemulsion area more when compared with 1:1, 1:2 and 1:3, the microemulsion obtained was stable in nature, this may be due to further reduction of the interfacial tension, increasing the fluidity of the interface, thereby increasing the entropy of the system. in Smix to 3:1 Figure. 24, the microemulsion region was more at Smix apex this was due to the addition of a large amount of Surfactant. This also results in the formation of gel type macroemulsion which was not stable for a long duration. Further reduction in microemulsion region was observed in Figure 25 Smix ratio 4:1

It is well known that large amounts of surfactants cause skin irritation, it is therefore important to determine the surfactant concentration properly and use the optimum concentration of surfactant in the formulation. From Pseudoternary phase diagrams, the formulations in which the amount of oil phase completely solubilized the drug and which could accommodate the optimum quantity of Smix and distilled water were selected for the study. The ratios of the optimized formulation were chosen from all the batches.

Characterization of the microemulsion

Droplet size of microemulsion

The Droplet size of microemulsion range from 9.236 to 41.29 large droplets of microemulsion will result in decreased flux in skin and help in retention of the formulation will lead to better therapeutic effect. The size of optimized microemulsion ME2 and ME6 was found to be 33.21 and 41.92 nm respectively.

Viscosity

Viscosity is an important parameter for topical drug delivery. A formulation having less viscosity will not retain over the skin for a prolonged time. The viscosity of microemulsion was found to 93.2 ± 0.2 to 327.0 ± 0.7 . The viscosity of optimized microemulsion was found to be 93.2 ± 0.2 and 119 as given in Table 21.

Zeta Potential

Highly positive or highly negative charge on oil globules indicate higher stability because of the anticipated surface repulsion between similarly charged globules hence inhibiting aggregation of the colloidal oil globules. the optimized

formulation ME2 and ME6 are considered to be stable as the zeta potential was - 33.27 and - 23.9. Figure 4a – 4d

pH

pH of microemulsion was in the range of 5.03 ± 0.08 to 5.86 ± 0.10 .

Refractive Index

Conductivity measurement using conductivity meter provides a way to determining whether a microemulsion is oil continuous or water continuous. More conductivity more will be the percentage of water, which allows more freedom for mobility of ions. Refractive index of Placebo formulation and sertaconazole loaded microemulsion was found to be near water so it as confirmed that it is oil in water microemulsion

Polydispersity Poly disparity index is a measure of particle homogeneity and it varies from 0.128 to 0.335 tables 21

Characterization of microemulsion gel

The droplet size of microemulsion gel was in the range of 37.19 to 43.06nm this is due to the addition of carbopol for converting microemulsion to microemulsion gel. Zeta potential of microemulsion gel was -24.9 to -33.17 (Table 22) highly negative or positive zeta potential values indicate stable formulation. ph of the formulation was 6.53 ± 0.07 to 6.79 ± 0.05 . Spreadability of microemulsion gel was found to be 14.18 ± 0.15 to 15.48 ± 0.64 increases in viscosity help in retention of the dosage form to the skin for a long duration. The designed formulation must have sufficient viscosity as it can easily spread over the affected or infected part. Microemulsion gel was homogeneous in nature that was confirmed by homogeneity test.

Skin irritancy test

Individual skin scores and of Primary Dermal irritation Index (PDI) of microemulsion (MG 1, MG2 and 2 % sertaconazole nitrate marketed preparation (control) are given in table 12-14. The tables show that the 2 % sertaconazole nitrate marketed preparation (control) and MG1 is 'Negligible Irritant having PDI=0.12 and PDI=0.185 respectively. From the Table 14, it was observed that microemulsion gel MG2 having PDI=0 considered is not irritating.

Permeation/Retention study

A superficially applied microemulsion is subjected to penetrate the stratum corneum and exist intact in the whole Horney layer. The main aim of the research work was to allow adequate concentration of the drug over and within the skin as to increase the chance of eradication of fungal infection. The main demerit of marketed cream was un able to maintain adequate concentration within the skin, as the maximum amount of drug was left intact over the donor compartment. Microemulsion gel MG 2 has adequate concentration over and within the skin and can provide effective cure rate. Figure 7. The permeation parameter also reveals that the MG2 has least permeation rate and least permeability coefficient. Table 8

Anti-inflammatory activity of MG1, MG2, and Marketed formulation

Anti-inflammatory activity of MG1, MG2 and marketed formulation was carried out using carrageenan induced induce paw edema. Microemulsion gel MG2 showed maximum anti

inflammatory activity when compared with MG1 and marketed formulation as given in Table 15 Figure 12.

Antifungal Activity

The antifungal property of optimized formulation from (MG 1 and MG2) and the control 2% Sertaconazole marketed formulation was determined using *Candida albicans*. (ATCC 10231) as representative fungi, adopting the Petri plate method. Average zone of inhibition for control (Marketed formulation), MG1 and MG2 was 15.34 ± 0.382 , 17.78 ± 0.715 and 23.19 ± 0.478 respectively. It is concluded that MG2 is having maximum antifungal activity.

Stability Studies

The data indicate that all the parameters of microemulsion were found to be stable systems. Stability of microemulsion was observed at different time intervals i.e., 0 (initial), 1, 3 and 6 months. All the characteristics of formulation ME 6 and MG 2 were found stable even after 6 months period. In case of formulation ME 2 the drug content was drastically decreased from 99.01 ± 0.2 to 88.19 ± 0.22 at $30^\circ\text{C}/65\% \text{ RH}$ and from 99.01 ± 0.2 to 86.12 ± 0.3 respectively, in case of MG 1 also the drug content were decreased from 99.01 ± 0.2 to 87.19 ± 0.22 at $30^\circ\text{C}/65\% \text{ RH}$ and 99.01 ± 0.2 to 84.12 ± 0.3 at $40^\circ\text{C}/75\% \text{ RH}$ as described in Table 17 to 20. All other parameter was found to be stable.

CONCLUSION

In the current study, the application of microemulsion systems in gel form for topical delivery of sertaconazole was investigated and pseudo ternary phase diagram was utilized to detect stable formulation. The microemulsion formulation of sertaconazole containing 2% (w/w) of sertaconazole, 6.67% (w/w) of oil phase (Eugenol+Oleic acid 1:1), 60.18% (w/w) of surfactant mixture (Tween-80 and Transcutol-P) and 33.15% (w/w) of distilled water has been optimized. The result suggests that the microemulsion gel MG2 was having more antifungal activity as compared to commercial cream and MG1. Permeation study of microemulsion gel MG 2 has adequate concentration over and within the skin and can provide effective cure rate. The anti-inflammatory activity of MG2 was more when compared with commercial cream and MG1. The skin irritation test of MG2 PDI=0 confirms that the formulation is safe to be used topically. The formulation was stable after storing at $30^\circ\text{C}/65\% \text{ RH}$ and $40^\circ\text{C}/75\% \text{ RH}$ for six months. From *in vitro* and *in vivo* data it can be concluded that the developed microemulsions have great potential for topical drug delivery.

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REFERENCES

1. Hoar H, Schulraan J H. The oleopathic hydro micelle. *Nature* 1943; 152: 102–103.
2. Friend D, Catz P, Heller, Barry BW. Lipid-protein partitioning theory of skin penetration enhancement. *Journal of Control Release* 1991; 15: 237-248.
3. Remington: The Science and Practice of Pharmacy. 19th edition, Mack Publishing Company, Easton, PA 1995; 2, 628–638.

4. Yalkowsky S H, Roseman, T J. Teclinique of Solubilizing of drugs by co solvent. *Drugs and Pharmaceutical Science*, Marcel Dekker Inc 1981, 12, 91-134.
5. Aulton M E. *Pharmaceutics: The science of dosage form design*, second edition, Churchill Livingstone,2002:120.
6. Azeem A, Rizwan M, Ahmad F J, Iqbal Z, Khar R Ket al., Nanoemulsion Components Screening and Selection. A Technical Note, American Association of Pharmaceutical Scientists 2009; 10:1: 69-76
7. Shakeel F, Baboota S, Ahuja A, Ali J, Aqil M et al., Nanoemulsions as vehicles for transdermal delivery of aceclofenac. *American Association of Pharmaceutical Scientists* 2007; 8: E1-E9.
8. Faizi M, Singh U K. RP-HPLC and UV Spectrophotometric methods for estimation of Sertaconazole nitrate in the microemulsion. *Journal of Chemical and Pharmaceutical Research* 2016; 8:740-745.
9. Baboota S, Shakeel F, Ahuja A, Ali J, Design development and evaluation of novel nanoemulsion formulations for the transdermal potential of celecoxib. *Acta Pharmaceutica* 2007; 57: 315-332.
10. Paolino D, Ventura C A, Nishi A, Puglisi Y, Fresta Y. Lecithin microemulsion for topical administration of ketoprofen, percutaneous adsorption through human skin and in vivo human skin tolerability, *International journal of pharmaceutics* 2002; 244: 21-31.
11. Attwood D, Kreuter J. *Colloidal Drug Delivery System*. New York, Marcel Dekker 1994; 31-71.
12. Shah D O. *Micelles, Microemulsion and Monolayers*. Science and technology Marcel Dekker 1998;1-610.
13. Ashish Y, Pawar, Vilas M, Aurangabadkar, Sunil K, Mahajan et al., formulation development and evaluation of topical microemulsion gels for nimesulide. *Journal of Pharmacy Research* 2011; 4: 1004-1006.
14. Bajpai M, Sharma P K, Mittal A. A study of the oleic acid oily base for the tropical delivery of dexamethasone microemulsion formulation. *Asian Journal of Pharmaceutics* 2009; 3: 208-214.
15. Shinoda K, Lindman B. Organised surfactant systems. *Microemulsions*, *Langmuir* 1987; 3:135-149.
16. Talegaonkar S, Azeem A, Ahmad FJ, Khar RK et al., Microemulsions: A Novel Approach to Enhanced Drug Delivery, *Recent Patents on Drug Delivery & Formulation* 2008;2: 238-257.
17. Jadhav KR, Jadhav S, Kadam S L, Design V J et al., Evaluation of Microemulsion Based Drug Delivery System; *International Journal of Advances in Pharmaceutical Sciences* 2010;1: 156-166.
18. Lieberman HA, Rieger MM, Banker GS. *Pharmaceutical Dosage Forms, Disperse systems*, New York, Marcel Dekker 2006; 3: 339-344.
19. Lachman L, Lieberman H A, Kanig J L, *The Theory and Practice of Industrial Pharmacy*, 3rd ed. Lea and Fibiger 1986; 510-511.
20. Claudia S, Adriana M, Carlucci C, Study of In Vitro Drug Release and Percutaneous absorption of fluconazole from topical dosage forms, *American Association of Pharmaceutical Scientists*, 2010; 11:2: 986-993.
21. Mrunali R, Rashmin B, Jolly R, Kashyap B, Ajay S, Investigating the effect of vehicle on in vitro skin permeation of ketoconazole applied in oil in water microemulsions, *Acta Pharmaceutica Scientia* 2010; 52: 65-77.
22. Kaur P, Kakkar S, topical delivery of antifungal agents, [expert opinion drug deli](#) 2010; 7: 1303-1327.
23. Spielin P, Homar M, Zupancic-Valant A, Gasperlin M. Sodium ascorbyl phosphate in topical microemulsions, *International Journal of Pharmaceutics* 2003; 256: 65-73.
24. Rhee YS, Choi JG, Park ES, Chi SC. Transdermal delivery of ketoprofen using microemulsions. *International Journal of Pharmaceutics* 2001; 228: 161-170.
25. Kawakami K, Yoshikawa T, Hayashi T, Nishihara Y, Masuda K. Microemulsion formulation for enhanced absorption of poorly soluble drugs. *Journal of Control Release* 2002; 81: 75- 82.
26. Mouluk S P, Paul B K, Structure, dynamics and transport properties of micro emulsions, *Advances in Colloid and Interface Science* 1998; 78: 99-195.
27. Lucero M J, Vigo J, Leon M J. A study of shear and compression deformations on hydrophilic gels of *International Journal of Pharmaceutics* 1994; 106: 125-133.
28. Mehta S K, Kavaljit X X, Bala K. Phase behavior, structural effects, volumetric and transport properties in non-aqueous microemulsions. *Physical reviews* 1999; 59: 4317-4325.
29. Podlogar F, Gasperlin M, Tomsic M, Jamnik A, Rogac M. Structural characterisation of water-Tween 40n microemulsions using different experimental methods. *International Journal of Pharmaceutics* 2004; 276:115-128.
30. Vijayalakshmi G, Amitava M, Chandrasekaran N. Mustard oil microemulsion formulation and evaluation of the bactericidal activity. *International Journal of Pharmacy and Pharmaceutical Sciences* 2012; 4: 497-500.
31. Salerno C, Bregni. Study of In Vitro Drug Release and percutaneous absorption of fluconazole from topical dosage forms. *American Association of Pharmaceutical Scientists* 2010; 11: 2: 986-993.
32. Zhu W, Guo C, Yu A, Gao Y, Cao F et al., Microemulsion based hydrogel formulation of penciclovir for topical delivery, *International Journal of Pharmaceutics* 2009; 378: 152-158.
33. Chen L, Tan F, Wang J, Liu F. Microemulsion: A novel transdermal delivery system to facilitate skin penetration of indomethacin. *Die Pharmazie* 2012; 67:319: 18-23.
34. Gasco MR. Microemulsions in the pharmaceutical field in perspectives and applications, *Industrial applications of microemulsions*. New York, Marcel Dekker Inc 1997; 97-122.
35. Das B, Nayak A K, Nanda U. Topical gels of lidocaine hydrochloride using cashew gum and Carbopol 940: Preparation and in vitro skin permeation, *International Journal of Biological Macromolecules* 2013; 62: 514-517.
36. Pereira E, Scolari P, Gasco M. Transdermal permeation of apomorphine through hairless mouse skin from microemulsions. *International Journal of Pharmaceutics* 2001; 226: 47-51.
37. Shishu, Rajan S, Kamalpreet. Development of novel microemulsion based topical formulations of acyclovir for the treatment of cutaneous herpetic infections. *American Association of Pharmaceutical Scientists*, 2009; 10: 559-65.
38. Laithy HM, El-Shaboury K MF. The development of cutting lip gels and microemulsion gel for topical administration of fluconazole. *American Association of Pharmaceutical Scientists* 2002; 3: 35.
39. Chakole CM, Shinde MA, Khadatkar SN. Formulation and development of novel combined clobetasol propionate and fusidic acid ointment. *International Journal of chem tech, Research* 2009; 1: 103-116
40. OECD. Acute dermal irritation,corrosion. In *OECD Guidelines for Testing of Chemicals*. Guideline 1992; 404: 6.
41. Purewal L, Gupta BN, Pande MS. Development and Evaluation of Herbal Formulations for Hair Growth. *Journal of Chemistry* 2008; 5: 34-38.
42. Winter CA, Risley EA, Nuss GW. Carrageenan induced edema in hind paw of the rat as an assay for anti-

- inflammatory drugs. *Proceedings of the Society for Experimental Biology and Medicine* 1962; 111: 544-547
43. Shekarachi M, Pirali-Hamedhani M, Navidpour L, Adib N and Shafiee. A Synthesis, antibacterial and antifungal activities of 3-aryl-5-(pyridine-3-yl)4,5-dihydropyrrole-1-carbothioamide derivatives. *Journal of the Iranian Chemical Society* 2008; 5:1:150-158.
44. Kini S, Gandhi AM. Novel 2-pyrazoline derivatives as potential antibacterial and antifungal agents. *Indian journal of pharmaceutical science* 2008; 70: 1:102-105.
45. Prashanth KV, Chauhan NS, Padh H, Rajani M. Search for antibacterial antifungal agents from selected Indian medicinal plants. *Journal of Ethnopharmacology* 2006; 107:182-188.
46. Perez C, Paul M, Bezique P. An Antibiotic assay by the agar well diffusion method. *Alta biomed. Group experiences* 1990; 15: 113.
47. Kataria, S, Rees GD, M J Lawrence. Gelatin-stabilised microemulsion-based organogens rheology and application in iontophoretic transdermal drug delivery. *Journal of controlled release* 1999; 60: 2: 355-365.
48. Block LH, Lieberman HA, Banker G S. Pharmaceutical emulsions, and microemulsions *Pharmaceutical Dosage Forms* 2001; 1: 47-110
49. Date AA, Nagarsenker M. Design and evaluation of self-nano emulsifying drug delivery systems for cefpodoxime paroxetine. *International journal of pharmaceutics* 2007; 329: 2: 166-172.

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