

INTERNATIONAL RESEARCH JOURNAL OF PHARMACY

www.irjponline.com ISSN 2230 - 8407

Research Article

PHYTOCHEMICAL INVESTIGATION, SPECTRUM ANALYSIS OF SALVADORA PERSICA AND CRESCENTIA CUJETE

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Article Received on: 27/06/21 Approved for publication: 28/07/21

DOI: 10.7897/2230-8407.1207150

ABSTRACT

Salvadorae persica is a well-known medicinal plant which belongs to the family Salvadoraceae. It is commonly known as the Meswak tree. Calabash tree or Crescentia cujete tree belongs to the family of Bignoniaceae. It is also known as the gourd tree. These two plants have been studied by preliminary phytochemical and FTIR analysis. Data gathered on solvent extraction and preliminary phytochemical method suggested that the presence of primary and secondary metabolites in leaf tissue. Two solvents such as ethanol and aqueous are used here to reveal the phytocompounds and extend our work to find out the various functional groups present in these two plants through FTIR analysis was done. Spectrum of Salvadora persica showed 5 peaks that are 599.89, 654.86, 1409.06, 1431.24, 2930.96 and spectrum of Crescentia cujete showed that 13 peaks that are 470.65, 520.80, 630.75, 652.93, 776.38, 1060.89, 1155.41, 1248, 1321.30, 1431.24, 1527.69, 2860.56, 2924.21. The present study revealed that the functional groups of both plants, such as alcohols or phenols, alkanes, amines, esters or carboxylic acid or lactones, aldehyde or ketones, acetates and ethylene.

Keywords: Crescentia cujete, Salvadora persica, FTIR analysis, Functional groups, Phytochemical analysis.

INTRODUCTION

Salvadora persica is a well-known medicinal plant which belongs to the family Salvadoraceae. It is commonly known as the Meswak tree, because the roots and twigs of this tree have been used for oral hygiene and dental care. It is found in shrub savannah, from northwestern India to Africa. It is an evergreen shrub, 4-6 m tall with a short trunk, white bark and smooth green leaves ¹.

Salvadora persica of roots are used for treatment of gonorrhea and relieve the pain due to spleen troubles. Bark is used for treatment of fevers. Leaves are used for treatment of asthma, cough and piles. Carminative and diuretically properties of fruits are used for treatment of rheumatisms². It has a capacity to treat these diseases due to the presence of biologically active compounds such as salvadoricine, salvadourea, trimethyl amine, di-benzyl thiourea, rutin, thioglucoside, chlorine, potash, sulphur etc³. Through traditional knowledge, it is believed that Salvadora persica has capacity to reduce blood cholesterol level. However, it is essential that before its repeated use for medicinal purpose, it should be subjected to botanical inspection to avoid side effects in the body. Since, phytosterols are added to foods because of their capacity to reduce the absorption of cholesterol in the gut and thereby it lowers the blood cholesterol level of the human body⁴.

The seeds of *Salvadora persica* containing about 40% oil with a fatty acid composition (lauric-20%, myristic-55%, palmitic 20% and oleic-5%) which can make an excellent soap. Seeds also contain fluoride and silica⁵. Seed's bark and leaves are reported to have fatty acid methyl ester (FAME), tocopherol (γ -tocopherol, α -tocopherol, vitamin E, and γ -tocotrienols), sterol (phytosterol, sitosterol, β -sitosterol, campesterol), stearic acid and phenolic compound⁶. The plant contains sulphur⁷, organic sulphur

compounds, ascorbic acid^{8, 9} and small amount of saponin. The chemical analysis of miswak sticks showed the presence of fluoride, calcium, phosphorus and silica. Fruits contain large amount of sugar, fat, colouring matter and an alkaloid, oilcake from the seeds contains nitrogen 4.8% potash 2.8% and phosphoric anhydride 1.05% ash contains large amount of chlorine¹⁰.

Calabash tree or Crescentia cujete tree belongs to the family of Bignoniaceae. It is also known as the gourd tree. The calabash tree is 6 to 10 m tall with a wide crown and long branches covered with clusters of tripinnate leaves and gourd-like fruit. The branches have simple elliptical leaves clustered at the anode. The greenish flowers arise from the main trunk and blooms at night¹¹. It is propagated either by seed or stem cuttings. Calabash fruit is a seasonal fruit that develops after pollination by bats. It appears at the end of dry season, and the fruit is up to 12 to 14 cm in diameter. It is globular with smooth hard green woody shell. It takes about six to seven months to ripen and eventually falls to the ground. Small flat seeds are embedded in the pulp¹². The plant principally contains tartaric acid, cianhidric acid, citric acid, crescentic acid, tannins, beta- sitosterol, alpha and beta amyrins, stearic acid, palmitic acid, flavonoids (quercetin and apigenin), iridoid 3-hydroxyoctanol naphthoquinones, glycosides, glycosides¹³.

Calabash is used in various parts of the world for their laxative, expectorant, anthelmintic, analgesic, anti-inflammatory and febrifuge properties. An antimicrobial property of the fruit leaves and bark of the Calabash tree has also been explored in various studies ¹⁴ and has yielded promising results. Of particular interest to this study is the fruit of *Crescentia cujete* or Calabash tree. Phytochemical study on the fruit yielded saponins, flavonoid, cardenolides, tannins, and phenol, as well as the presence of hydrogen cyanide.

Extracts from the various parts of *C. cujete* possessed antiinflammatory¹⁵, antibacterial¹⁶, DPPH radical scavenging¹⁷, antioxidant¹⁸, CNS depressant¹⁹, wound healing activities²⁰. The present investigation aimed to screen out the phytochemicals from leaf extracts of *Salvadora persica* and *Crescentia cujete*

MATERIALS & METHODS

Plant materials

Salvadora persica

Salvadora persica is a small tree or shrub with a crooked trunk, typically 6–7 meters (20–23 ft) in height. Its bark is scabrous and cracked, whitish with pendulous extremities. The root bark of the tree is similar in colour to sand, and the inner surfaces are an even lighter shade of brown. It has a pleasant fragrance, of cress or mustard, as well as a warm and pungent taste. The leaves break with a fine crisp crackle when trodden on. The tree produces small red edible fruits, juicy but pungent, in clusters.

Crescentia cujete

Crescentia cujete is a smooth, much-branched tree growing to a height of 4 to 5 meters. Branches are arching with close-set clusters of leaves. Leaves are alternate, often fascicled at the nodes, oblanceolate, 5 to 17 centimeters long, glossy at the upper surface, blunt at the tip and narrowed at the base. Flowers develop from the buds that grow from the main trunk, yellowish, sometimes veined with purple, with a slightly foetid odor, occurring singly or in pairs at the leaf axils, stalked and about 6 centimeters long, and opens in the evening. Calyx is about 2 centimeters long and split into two lobes. Fruit is short stemmed, rounded, oval or oblong, green or purplish, 15 to 20 centimeters in diameter. Voucher specimen is preserved as herbarium in department of botany (ACH 2018,2019), The American College, Madurai.

Preparation of crude plant extracts

Solvent extraction

The collected plant materials (leaves of *Salvadora persica* and *Crescentia cujete*) were washed with running tap water to remove dusts and other particles. Then the materials were shade dried and blundered as coarse powder. The powder materials (200 gm) were subjected to extraction with solvent (ethanol) in Soxhlet's apparatus with continuous 8 hrs reflux and also. The crude extracts concentrated in room temperature and stored in desiccator.

Aqueous extract

The dust free plant material was ground with mortar and pestle in adequate amount of sterile distilled water. The homogenized tissue was standing out for separation (Cold maceration). The supernatant was taken as aqueous extract.

Qualitative phytochemical analysis

The concentrated extracts were used for preliminary phytochemical screening. The concentrated extracts are dissolved again in solvents (ethanol and aqueous) respectively and used for screening. The following tests were conducted for screening the phytocompounds²¹.

Test for carbohydrates (Molisch's test)

2 ml of plant extract in a boiling tube added with 2 ml of Molisch's reagent and few drops of Conc. Sulphuric acid. Presence of purple or reddish color indicates the presence of carbohydrates.

Test for tannins (Ferric chloride test)

2 ml of plant extract, 2 ml of 5% ferric chloride is added in a boiling tube. The mixture turned into dark blue or greenish black indicates the presence of tannins.

Test for saponins (Foam test)

2 ml of plant extract and 2 ml of distilled water is added and shaken gently in a graduated cylinder for 15 minutes. Appearance of foaming layer indicates the presence of saponins.

Test for flavonoids

2 ml of plant extract added with NaOH (Sodium hydroxide) solution in a test tube. Appearance of yellow colour indicates the presence of flavonoids.

Test for alkaloids (Mayer's test)

2 ml of plant extract and 2 ml of concentrated hydrochloric acid is added in a test tube. Then few drops of Mayer's reagent (Potassium Iodide + Mercuric Chloride) are added. The appearance of green color or white precipitate indicates the presence of alkaloids.

Test for quinines

2 ml of extract and 2 ml of Concentrated Sulphuric acid is added. The solution turn into red color indicates presence of quinines.

Test for glycosides

2 ml of plant extract, 2 ml of chloroform and 2 ml of acetic acid gradually added alone the wall of test tube. The solution changed blue to green color that indicates presence of glycosides.

Test for Cardiac glycosides (Keller Kiliani test)

2 ml of extract, 2 ml of glacial acetic acid and few drops of 5% Ferric chloride added in a test tube. The mixture will be added 1 ml of concentrated Sulphuric acid on the top layer. The solution mixture changed into brown ring at the middle indicates presence of cardiac glycosides.

Test for Terpenoids

2 ml of extract, 2 ml of chloroform added, and concentrated Sulphuric acid added carefully in a test tube. Appearance of reddish-brown color at the junction of layer indicates presence of terpenoids.

Test for Phenols

2 ml of the extract, 2 ml of Ferric chloride solution added in a tube. Appearance of orange yellow colour indicates presence of phenols.

Test for coumarins

2 ml of extract, 2 ml of 10% sodium hydroxide added. Formation of yellow color indicates presence of coumarins.

Phytosteroids

2 ml of plant extract equal volume of chloroform is added in a test tube and then added few drops of concentrated Sulphuric acid. Appearance of bluish brown ring indicates the presence of phytosteroids.

Phlobatannins

2 ml of plant extract few drops of 2% Hydrochloric acid added in a test tube. The mixture changed into red color precipitate that indicates the presence of phlobatannins.

Anthraquinone

2 ml of plant extract and with few drops of 10% Ammonia solution added. Appearance of pink color precipitate indicates the presence of anthraquinones.

IR Spectrum

The FTIR spectrum of solid sample was obtained from ethanolic extract of plant leaves powder. To run the solution, a drop of the extract was placed on the face of a highly polished salt plate (i.e. 100mg of potassium bromide, Kbr). A second plate was placed on the top of the first plate so as to spread liquid in these layers between the dried plates and clamp the plates together in suitable fashion. Then the spectrum was obtained with the help of spectrophotometer and computer software attached with it²². The frequencies of different compounds present in the sample were analysed. The same procedure was followed for the suitable standard. The samples were run at infrared region between 400nm and 4000nm. FTIR is an effective analytical instrument for detecting functional groups and characterizing covalent bonding information for IR spectroscopic study.

RESULT

The present study subjected to screen the preliminary phytochemical constituents of two plants namely *Salvadora* persica and *Crescentia cujete* by qualitative tests.

Qualitative phytochemical analysis

The qualitative phytochemical analysis of *Salvadora persica* and *Crescentia cujete* with two different solvents, viz., ethanol and aqueous extracts were presented (Table 1 & Table 2). The ethanolic extract of *Salvadora persica* showed the presence of compounds such as flavonoid, alkaloid, glycosides, terpenoids, phenols, anthraquinones. The aqueous extract showed the presence of compounds such as tannins, saponin, flavonoids, alkaloid, cardiac glycosides, phenols, coumarins. Some of the following compounds are totally absent in two extracts also such as carbohydrates, proteins, quinines, steroids, phlobatannins. Among two solvents, the leaf aqueous extract showed the presence of highest phytocompounds in *Salvadora persica* plant (Table1).

The ethanolic extract of *Crescentia cujete* showed the presence of compounds such as Tannins, flavonoids, quinines, glycosides,

Table 1. Qualitative phytochemical analysis of different solvent extracts (Ethanol and Aqueous) of Salvadora persica (leaf)

Phytochemicals	Ethanol	Aqueous
Carbohydrates	-	-
Tannins	-	+
Saponin	-	+
Flavonoid	+	+
Alkaloid	+	+
Quinones	-	_
Glycosides	+	_
Cardiac glycosides	_	+
Terpenoids	+	_
Phenols	+	+
Coumarins	-	+
Proteins	-	_
Steroids	-	_
Phlobatannins	-	_
Anthraquinones	+	_

terpenoids, steroids. The aqueous extract showed the presence of compounds such as Anthraquinones, coumarins, terpenoids, cardiac glycosides, alkaloids, flavonoids, tannins. Some of the following compounds are totally absent in two extracts also such as carbohydrates, saponin, phenols, proteins, phlobatannins. Like *Salvadora persica*, *Crescentia cujete* has highest phytocompounds in aqueous extract while compared to ethanolic extract (Table 2).

FTIR

FTIR spectroscopy was useful for identification of functional groups, when run under IR region from 400 - 4000cm-1. There was a variation in the peaks of the sample which helps to determine the functional groups. Each peak represents single compound based on their density. The typical IR spectra for two plant extract are obtained and shown (Figure 1 and Figure 2). Figure 1 represents the IR spectrum of *Salvadora persica* leaf. In this spectrum 5 peaks are identified that are 599.89, 654.86, 1409.06, 1431.24, 2930.96. Figure 2 represents the IR spectrum of *Crescentia cujete* leaf. In this spectrum 13 peaks are identified that are 470.65, 520.80, 630.75, 652.93, 776.38, 1060.89, 1155.41, 1248, 1321.30, 1431.24, 1527.69, 2860.56, 2924.21.

The broad absorption around 3797.84 cm -1and 2353.16 cm -1 the O-H stretching frequency is due to the presence of alcohols or phenols. Another prominent absorption around 2918.30 cm -1and the weak absorption such as 1371.39cm -1, 1325.10cm -1, is mainly due to alkanes (C-CH 3 stretching vibrations). In the spectrum, the medium range such as 1618.28 cm -1,1631.78 cm -1, is mainly due to amines (C=C stretching vibration). The C=O stretching vibrations at 1728.22 cm-1, due to the presence of esters or carboxylic acid or lactones. At 1442.75 cm -1, CH bending frequency is due to the aldehyde or ketones. Particularly below 1200 cm -1, C-O-C vibrations in ester at the range between 1100 cm-1 to 1200 cm -1 due to the presence of acetates or formats. At the range between 600 cm -1 to 700 cm -1, -CH-CH-bond frequency, is mainly due to the ethylene groups.

Table 2. Qualitative phytochemical analysis of different solvent extracts (Ethanol and Aqueous) of *Crescentia cujete* (leaf)

Phytochemicals	Ethanol	Aqueous
Carbohydrates	-	_
Tannins	+	+
Saponin	-	-
Flavonoid	+	+
Alkaloid	-	+
Quinones	+	-
Glycosides	+	ı
Cardiac glycosides	_	+
Terpenoids	+	+
Phenols	-	1
Coumarins	-	+
Proteins	-	-
Steroids	+	_
Phlobatannins	-	_
Anthraquinones	-	+

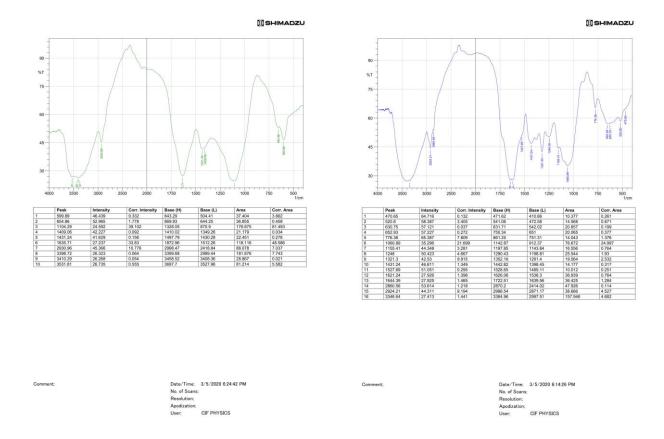


Fig 1. FTIR analysis of Salvadora persica (leaf)

Fig 2. FTIR analysis of Crescentia cujete (leaf)

DISCUSSION

Phytochemical analysis

Seeds bark and leaves of *Salvadora persica* are reported to phenolic compound²³. Similarly the ethanolic extract of *Salvadora persica* showed the presence of phenols in our study. *Salvadora persica* contained small amount of saponin²⁴. Fruits contain large amount of sugar, and alkaloid from the seeds. In our study aqueous and ethanol extracts both showed the presence of compounds like saponin, alkaloids. But trace amount of sugar (carbohydrates) may be present in crude extract, but it is not responsible to preliminary phytochemical analysis. Several research are going on this particular species around the world. Three lignan glycoside compounds are identified²⁵ and flavonoid compounds such as rutin and quercetin were detected in the stems of *Salvadora persica*²⁶. In overall, crude extract contains the same compounds in both extracts.

Phytochemical study of *Crescentia cujete* fruits are yielded saponins, flavonoid, cardenolides, tannins, and phenol (Stuartxchange). The leaves extract of *C. cujete* and *C. alata* showed glycosides compounds²⁷. Several Anthracene derivatives compounds are coumarins, lignans, triterpenes, steroids mono and diterpenes, anthraquinones that are obtained from the methanolic extract of leaves of *C. cujete*²⁸, similarly the related genus *Kigelia pinnata* showed the same phytocompounds²⁹. Like *Salvadora persica*, *Crescentia cujete* has highest phytocompounds in aqueous extract while compared to ethanolic extract.

FTIR analysis

FTIR spectroscopy was useful for identification of functional groups, when run under IR region from 400 - 4000cm-1. There was a variation in the peaks of the sample which helps to determine the functional groups. Each peak represents single compound based on their density. The typical IR spectra for two plant extract are obtained. The IR spectrum of *Salvadora persica* leaf revealed 5 peaks that are identified as 599.89, 654.86, 1409.06, 1431.24, 2930.96. The IR spectrum of *Crescentia cujete* leaf revealed 13 peaks that are identified as 470.65, 520.80, 630.75, 652.93, 776.38, 1060.89, 1155.41, 1248, 1321.30, 1431.24, 1527.69, 2860.56, 2924.21.

In both plants, IR spectrum showed the broad absorption around 3797.84 cm -1 and 2353.16 cm -1 the O-H stretching frequency is due to the presence of alcohols or phenols. Another prominent absorption around 2918.30 cm -1 and the weak absorption such as 1371.39cm -1, 1325.10cm -1, is mainly due to alkanes (C-CH 3 stretching vibrations). In the spectrum, the medium range such as 1618.28 cm -1,1631.78 cm -1, is mainly due to amines (C=C stretching vibration). The C=O stretching vibrations at 1728.22 cm-1, due to the presence of esters or carboxylic acid or lactones. At 1442.75 cm -1, CH bending frequency is due to the aldehyde or ketones. Particularly below 1200 cm -1, C-O-C vibrations in ester at the range between 1100 cm-1 to 1200 cm -1 due to the presence of acetates or formats. At the range between 600 cm -1 to 700 cm -1, -CH-CH- bond frequency, is mainly due to the ethylene groups.

Similarly, FTIR spectrum analysis of *Caralluma fimbriyata* revealed the peak value at 2970.38 and 2885.51 cm⁻¹ refers to the presence of alkanes (C–H stretch). The peak at 1759.08 and 1666.50 cm⁻¹ corresponds the carboxylic acid group (C=O stretch). A peak of 1327.03 cm⁻¹ showed the presence of aromatic amines (C–N stretch). The peaks of 1273.02, 1087.85 and 1049.28 cm⁻¹ indicate the alcohols, carboxylic acids, esters, ethers (C–O stretch). A peak of 879.54 cm⁻¹ revealed the alkenes (=C–H bend). These basic reports are used to find out found the presence of phenols, alkanes and aromatic amines³⁰. Same absorption spectrums are obtained in extract of *Aerva lanata*³¹, *Albizia lebbeck*³² and *Ardisia blatteri*³³.

CONCLUSION

Salvadora persica is a large shrub or a small tree. The plant is highly medicinal plant. Moreover, trees are often cultivated for roots as it is extensively used in preparation of Meswak toothpaste. The tree possesses multiple medicinal benefits. Crescentia cujete is otherwise called as Calabash tree. This plant is widely cultivated as an ornamental shade or specimen tree in tropical areas. This plant has been used as medicinal plant since ancient times. Crescentia cujete possessed various therapeutic activities in related to findout drug in pharmacological aspects. Over exploitation over the last few decades have threatened the existence of both important multipurpose trees (Salvadora persica and Crescentia cujete). These plants need urgent restoration and conservation that is severely required.

ACKNOWLEDGMENTS

Heartfelt thanks to the Principal & Secretary, The American College, Madurai for permitted us to carry out the research work on college premises and support in many phases of academic career. We thank PG Head & UG Head giving us permission to carry out this work and providing all facilities needed to execute this study.

REFERENCES

- 1. Ahmad H and Rajagopal K. Biological Activities of *Salvadora persica* L. (Meswak). Med. Aromat Plants 2013;2: 129.
- 2. Khatak M, Khatak S, Siddqui AA, Vasudeva N, Aggarwal A and Aggarwal P. *Salvadora persica*. Pharmacogn Rev 2010; 4(8): 209–214.
- 3. Malik S, Ahmad SS, Haidar SI and Muzaffar A. Salvadoricine, A new alkaloid from the leaves of *Salvadora persica*. Tetrahedron lett 1987; 28:163-164.
- 4. Hayes KC, Pronczuk A, Wijendran V and Beer M. Free phytosterols effectively reduce plasma and liver cholesterol in gerbils fed cholesterol. J. Nutr 2002; 132(7): 1983-1988.
- 5 Ahmed SS, Gengaihi SEE, Ibrahim MS and Schnug E. Preliminary phytochemical and propagation trial with *Salvadora persica* L. Agriculture and Forestry Research 2008;1(58): 135-138.
- 6. Mariod AA, Mathaus B and Hussein IH. Chemical characterization of the seed and antioxidant activity of various parts of *Salvadora persica*. Journal of the American oil chemist's society, Research Library 2009; 86:9.
- 7. Awasthi CY and Mitra CR. *Salvadoraoleoides*, constituents of the fruit and seed. Indian oilseeds Journal 1964; 8: 289-292.
- 8. Daxenbichler ME, Spencer GF, Carlson DG, Rose GB, Brinker AM and Powell RG. Glucosinolate composition of seeds from 297 species of wild plants. Phytochemistry 1991; 30:2623-2638. 9. Farooqi MIH and Srivastava JG. The toothbrush tree. Quart J Crude Drug Res 1968; 8: 1297-1299.

- 10. Von Kampf R and Miswak U. Eine zur Zahnreingigung verwendete Drogeder arabischen Volksmedizin. Pharm Acta hely 1975; 50: 350-352.
- 11. Nadkari K. M. The Indian Matria Medica. Popular Prakashan Publishers;1999.
- 12. Michael A. Trees, Shrubs and Liannas of West Africa Dry Zones. Grad Margae Publishers; 2004.
- 13. Marc NO. Nutritive and anti-nutritive compositions of calabash fruit pulp. Journal of Veterinary Advances 2008;7(9):1069-1072.
- 14. Bussman RW, Glenn A and Sharon D. Antibacterial activity of medicinal plants of Northern Peru can traditional applications provide leads for modern science. Indian Journal of Traditional Knowledge 2010; 9(4): 102-107.
- 15.Parvin MS, Das N, Jahan N, Akhter MA, Nahar L and Islam ME. Evaluation of in vitro anti-inflammatory and antibacterial potential of *Crescentia cujete* leaves and stem bark. BMC Res Notes 2015; 8:412.
- 16. Mahbub KR, Hoq M, Ahmed MM, and Sarker A. In vitro antibacterial activity of *Crescentia cujete* and *Moringa oleifera*. Bangladesh Research Publication Journal 2011;5(4): 337 343.
- 17. Juceni PD, Marilena M, Jorge MD, Hugo NB, Alexsandro B, Fátima MA. Radical scavenging, antioxidant and cytotoxic activity of Brazilian Caatinga plants. Fitoterapia 2007; 78(2):215-218
- 18. Akinmoladun AC, Obuotor EM and Farombi EO. Evaluation of antioxidant and free radical scavenging capacities of some Nigerian indigenous medicinal plants. Journal of Medicinal Food 2010; 13(2):444-451.
- 19. Aderibigbe AO, Olufunmilayo T and Agboola CNSO. Depressant properties of the crude extract of *Crescentia cujete* in mice. Planta Medica 2013; 79(13):423-427.
- 20. Campos GS, de Oliveira Jr SA, Borges LBP, Ribeiro IP, Martinez SB and Ayer IM. Use of extract of coite (*Crescentia cujete*) as a phytotherapic in injuries of horses. Investigacao 2016; 15(4):95-97.
- 21. Harborne JB. Phytochemicals Methods.Chapman and Hall Ltd; 1973.
- 22. Idrees M, Khan S and Riazuddin S. Common genotypes of hepatitis B virus. J.Coll.Phy. Sur. Pak 2004; 14: 344-347.
- 23. Mariod A, Matthäus B. Investigations on fatty acids, tocopherols, sterols, phenolic profiles and oxidative stability of Cucumismelo var. agrestis oil. J Food Lipids 2008; 15(1):42–55.
- 24. Von Kampf R and Miswak U. Eine zur Zahnreingigung verwendete Drogeder arabischen Volksmedizin. Pharm Acta hely 1975; 50: 350-352.
- 25. Kamel MS, Ohtani K, Assaf MH, Kasai R, El-Shanawani MA, Yamasak K, Ali AA and Tanaka O. Lignan glycosides from stems of *Salvadora persica*. Phytochemistry 1992;31: 2469-2471.
- 26. Wahab SMA, Selim MA and El-Fiki NM. Investigation of the flavonoid content of *Salvadora persica* L. Bull, FacPharm.(Cairo Univ) 1990;28: 67-70.
- 27. Agalwal K and Popli SP. The constituents of *Crescentia cujete* leaves. Fitoterapia 1992; 63:476.
- 28. Olaniyi MB, Lawal IO and Olaniyi AA. Proximate, phytochemical screening and mineral analysis of *Crescentia cujete* L. leaves. J. Med. Plants Econ. Dev 2018; 2:28.
- 29. Dhanasekaran M, Abraham GC, Mohan S. Preliminary phytochemical and histochemical investigation on *Kigelia pinnata* DC. International Journal of Pharma Sciences and Research 2014; 5:7.
- 30. Packialakshmi N and Naziya S. Fourier transform infrared spectroscopy analysis of various solvent extracts of *Caralluma fimbriyata*. Asian J Biomed Pharm Sci 2014; 4:20-5.
- 31. Murugan M and Veerabahu RM. Phytochmemical, FTIR and antibacterial activity of whole plant extract of *Aerva lanata*. Journal of Medicinal Plants Studies 2014; 2(3): 51-57.

- 32. Bobby MD, Wesely EG and Johnson M. FTIR studies on the *Albizia lebbeck*. International journal of Pharmacy and Pharmaceutical Sciences 2012; 4(3): 293-296.
- 33. Dhanasekaran M. Phytochemical screening of *Ardisia blatteri* anendemic plant of Southern Western Ghats, Tamilnadu, India. International Research Journal of Pharmacy 2016; 7(2): 31-35.

Cite this article as:

M.Dhanasekaran *et al.* Phytochemical investigation, spectrum analysis of *Salvadora persica* and *Crescentia cujete*. Int. Res. J. Pharm. 2021;12(7):8-13.

http://dx.doi.org/10.7897/2230-8407.1207150

Source of support: Nil, Conflict of interest: None Declared

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