



Research Article

ANTIOXIDANT ACTIVITIES OF METHANOLIC EXTRACTS FROM TEN *PLEUROTUS* SPECIES

Sanjit Debnath ^{1*}, Ramesh Chandra Upadhyay ², Panna Das ³ and Ajay Krishna Saha ¹

¹Mycology and Plant Pathology Laboratory, Department of Botany, Tripura University, Suryamaninagar, Tripura, India

²Directorate of Mushroom Research, Chambaghat, Solan, Himachal Pradesh, India

³Microbiology Laboratory, Department of Botany, Tripura University, Suryamaninagar, Tripura, India

*Corresponding Author Email: sanjitdebnath2888@gmail.com

Article Received on: 09/02/17 Approved for publication: 08/03/17

DOI: 10.7897/2230-8407.080335

ABSTRACT

The antioxidant activities of methanolic extract from mycelia of ten *Pleurotus* species were investigated. The main aim of this study was to evaluate and compare the antioxidant activities of methanolic extracts of mushroom mycelium of ten *Pleurotus* species by three different methods. The DPPH (1,1-diphenyl-2-picrylhydrazyl) radical scavenging activity, reducing power, chelating effect on ferrous ion and phenolic content of mycelial extract of ten edible mushrooms were analyzed. *P. sajor-caju* showed highest free radical scavenging activity (92.05 %) and reducing power (1.082 %) at 8 mg/ml concentration. *P. citrinopileatus* showed highest percentage of chelating effect on ferrous ion (90.66 %). The lowest EC₅₀ value of free radical scavenging activity was found in *P. sapidus* which indicated strongest ability of the mycelial extract to act as DPPH radical scavenger. The lowest chelating effect on ferrous ion was noticed in *P. sajor-caju* but EC₅₀ of reducing power was much lower than the synthetic antioxidant (BHT). The revealed data showed that mycelia of all studied *Pleurotus* species possessed potent antioxidant activity and their inclusion in the diet may help to prevent diseases caused by oxidative damage.

Keywords: Antioxidant activity, EC₅₀ value, edible mushroom, *Pleurotus* species, total phenol.

INTRODUCTION

Mushrooms have been reported as therapeutic foods that are useful in preventing diseases such as hypertension, hypercholesterolemia and cancer due to their chemical composition. The genus *Pleurotus* species comprises about 40 species¹, they are ubiquitous, being found in both temperate and tropical part of the world and are now considered to be the second most important cultivated mushroom in the world². *Pleurotus* mushrooms (oyster mushrooms) are quite easily cultivated artificially, most often in liquid medium³ and they are also appreciated as a food due to flavor along with medicinal and bioremediation properties.

Almost all organisms possess antioxidant defense and repair systems that have evolved to protect them against oxidative damage, these systems are not sufficient to prevent the damage entirely⁴. Natural antioxidants are being extensively studied for their capacity to protect organisms and cells from damage brought on by oxidative stress due to ageing and degenerative diseases⁵. Wild mushrooms are traditionally used in many Asian countries and also in other continents as both food and medicine^{6,7}. Wild mushrooms, cultivated mushrooms are becoming more important in our diet for their nutritional and pharmacological properties⁸ and mycelial mushrooms are also important for their secondary metabolite production which contains medicinal property. There are many researches of antioxidant activity on wild and cultivated mushroom in the world.

Our objective was to evaluate and compare the antioxidant activities of methanolic extracts of mushroom mycelium of ten *Pleurotus* species viz. *P. sapidus* (Schulzer) Sacc. (DMRP-4), *P.*

sajor-caju (Fr.) Singer (DMRP-112), *P. membranaceus* Masee. (DMRP-189), *P. ostreatus* (Jacq. ex Fr.) Kummer (DMRP-262), *P. flabellatus* (Berk. & Broome) Sacc (DMRP-5), *P. florida* (Mont.) Singer (DMRP-88), *P. hypsizygus ulmarius* (Bull.) Redhead (DMRP-115), *P. djamor* (Rumph. ex Fr.) Boedijn (DMRP-205), *P. eryngii* (DC.) Quel (DMRP-135) and *P. citrinopileatus* Singer (DMRP-10) by three different methods (DPPH method, Reducing power method and Chelating effect on ferrous ion).

MATERIALS AND METHODS

Sample

Fungal mycelia cultures of ten *Pleurotus* species viz. *P. sapidus* (Schulzer) Sacc. (DMRP-4), *P. sajor-caju* (Fr.) Singer (DMRP-112), *P. membranaceus* Masee. (DMRP-189), *P. ostreatus* (Jacq. ex Fr.) Kummer (DMRP-262), *P. flabellatus* (Berk. & Broome) Sacc (DMRP-5), *P. florida* (Mont.) Singer (DMRP-88), *P. hypsizygus ulmarius* (Bull.) Redhead (DMRP-115), *P. djamor* (Rumph. ex Fr.) Boedijn (DMRP-205), *P. eryngii* (DC.) Quel (DMRP-135) and *P. citrinopileatus* Singer (DMRP-10) were procured from Directorate of Mushroom Research, Chambaghat, Solan, India.

Mushroom Mycelia Production

Pure cultures of all the *Pleurotus* species was carried out in Potato Dextrose Agar (PDA) medium. To study antioxidant activity, mushroom mycelium was grown sterile conical flask (250 mL) containing 50 ml of Basal Synthetic (BSL) Medium. The BSL Medium contained glucose (30 g/L), yeast extract (2 g/L), peptone (2.5 g/L), MgSO₄ · 7H₂O (0.5 g/L), Ca(NO₃)₂ (0.5 g/L), (NH₄)₂SO₄ (0.25 g/L), KH₂PO₄ (0.25 g/L), FeCl₃

(0.01g/L), ZnSO₄ (0.0001 g/L), Inositol (0.05 mg/L), Thiamine (100 µg/L), Biotin (50 µg/L), Folic acid (100 µg/L), CaCl₂ (0.1 M in 5 mL/L) and distilled water (1 L)⁹. The medium was inoculated with disk of 6 mm diameter of mushroom mycelia obtained from six to eight days old grown culture on Potato Dextrose Agar (PDA) plate. The growth was carried out under stationary condition at 28°C (Shaking Incubator: LSI 4018R). After 30 days incubation in the dark, the liquid medium was filtered and the mycelium separated from the liquid¹⁰.

Mushroom Mycelia Extraction

Preparation of methanolic extracts of mushroom was done by slightly modified method of Mau et al.¹¹. The dried powdered mycelium (5g) was extracted by grinding with 50 mL of methanol with the help of pestle and mortar. After filtering through Whatman No.4 filter paper, the mycelium was then extracted twice with addition of 50 mL of methanol in each. The methanolic extract was then evaporated at 40°C to dryness in rotary evaporator (Rotavap: PBV-7D). The dried extract was used directly for determination of antioxidant activities.

Free Radical Scavenging Activity (DPPH)

Free radical scavenging activity (FRS) activity was measured by a little bit modified method of Shimada et al.¹². 4 ml dried mushroom methanolic extract (0.25- 16 mg/ml) was mixed with 1 ml of (0.0002 M) methanolic solution containing 1,1-diphenyl-2-picrylhydrazyl (DPPH) radical (Sigma). The mixture after shaking vigorously was allowed to stand for 30 min and the absorbance was measured at 517 nm against a blank in a spectrophotometer (Eppendorf AG 22331Hamburg). EC₅₀ (mg/mL) is the valuable concentration at which DPPH radical were scavenged by 50% (w/v) and was obtained by interpolation from linear regression analysis. BHT and ascorbic acid were used as a control. Inhibition of free radical by DPPH in percent was calculated as follows: Percentage of inhibition: $[(A_{\text{Blank}} - A_{\text{Sample}}) / A_{\text{Blank}}] \times 100$, Where, A_{Blank} and A_{Sample} denotes the absorbance of control and test compound respectively.

Reducing Power

Reducing power was determined by the slightly modified method of Oyaizu¹³. Each mycelial extract (0.5-8.0 mg/mL) in methanol (2.5 mL) was mixed with 2.5 mL of 0.2 M sodium phosphate buffer and 2.5 mL of 1% potassium ferricyanide and the mixture was incubated at 50°C for 20 min. Then 2.5 mL of 10 % (w/v) trichloroacetic acid (TCA) were added. After centrifugation at 200 g for 10 min, 5 mL of upper layer was mixed with 5 mL of deionised water and 1 mL of ferric chloride (0.1 %). The absorbance was measured against a blank in 700 nm in spectrophotometer (Eppendorf AG 22331Hamburg). EC₅₀ (mg/mL) is the effective concentration at which the absorbance was 0.5 for reducing power. BHT was used as control.

Chelating Effect on Ferrous Ion

It was determined by the method of Decker & Welch¹⁴. 2 ml of each methanolic extract at various concentration (0.05-1.5 mg/ml) various concentrations of extract in the methanol was added to a solution of 0.002 M FeCl₂ (0.05 ml). The reaction was initiated by the addition of 0.005 M ferrozine (0.2 ml). The total volume was made to 5 ml with methanol. Then, the mixture after shaking vigorously was allowed to stand for 10 min in room temperature and the absorbance was measured at 562 nm in spectrophotometer (Eppendorf AG 22331Hamburg). A

mixture without extract was used as the control. Ethylenediaminetetraacetic acid (EDTA) was used as a standard. The inhibition percentage of Ferrozine Fe²⁺ complex formation was then calculated: Metal Chelating effect (%) = $\{(A_0 - A_1) / A_0\} \times 100$ %, Where, A₀ and A₁ is the absorbance of the control and the sample respectively. EC₅₀ (mg/ml) value was calculated from the graph of ferrous ion inhibition percentage against extract concentration.

Fungal Phenol Estimation

Total phenols was determined according to the method reported by Swain and Hillis, 1959¹⁵ (Folin ciocalteu reagent method) with the Folin-Ciocalteu reagent, using tannic acid in ethanol (80%, w/v) as standard.

Statistical Analysis

For each one of the mushroom species, the assays were carried out in triplicate form. The average data recorded for each replica were subjected to the one way ANOVA technique using Origin 7 followed by Tukey's Least Significant Differences (LSD). The results were expressed as mean values ± standard deviation (SD) and the significance was tested.

RESULTS AND DISCUSSION

Free Radical Scavenging (FRS) Activity

The methanolic extract of mycelia was subjected to screening for possible antioxidant activity by the DPPH free radical scavenging method. Scavenging the stable DPPH radical is widely used method to evaluate the antioxidant activity in comparison to other method because this method is simple, requires short period of time and sensitive. DPPH is a stable free radical that shows a characteristic absorbance at 517 nm, which decreases significantly when exposed to radical scavengers by providing hydrogen atom or electron to be a stable diamagnetic molecule.

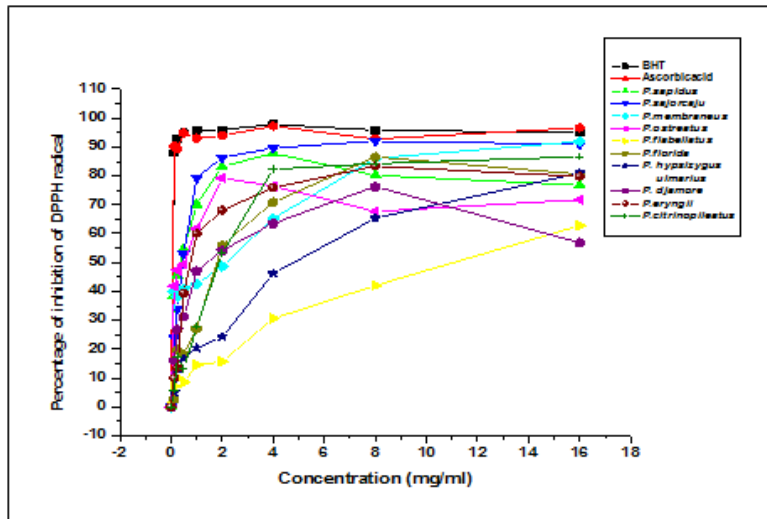
Methanolic extracts of mushroom mycelia of *Pleurotus* species showed varying scavenging ability. Among the ten *Pleurotus* species, *P. sajor-caju* showed highest free radical scavenging effect (92.05±1.37%) at 8.0 mg/ml concentration and followed by *P. membranaceus* (91.92±0.46 %), *P. citrinopileatus* (86.63±0.07), *P. florida* (86.58±0.52 %), *P. eryngii* (83.33±2.32%), *P. hypsizygus ulmarius* (80.98±3.78 %), *P. sapidus* (80.20±1.87 %), *P. ostreatus* (79.29±1.25 %), *P. djamor* (76.31±1.79 %) and *P. flabellatus* (62.75±1.87%). However, highest FRS activities of both the synthetic antioxidants BHT, ascorbic acid were 97.83% at 0.125 mg/ml concentration and 97.25% at 4 mg/ml concentration, respectively. Variations in FRS activities by various *Pleurotus* species were graphically represented in Graph 1.

EC₅₀, is the effective concentration at which the antioxidant activity was 50% and DPPH radicals were scavenged by 50%. EC₅₀ was obtained by interpolation from linear regression analysis. The strongest EC₅₀ of FRS activity had been found in *P. sapidus* at 0.352 mg/ml concentration in comparison to other *Pleurotus* species. Synthetic antioxidant (BHT and ascorbic acid), which was used as a standard, had a superior (*p*<0.05) EC₅₀ (0.049 mg/ml) value in comparison to ten *Pleurotus* species. Table 1. represented the EC₅₀ of FRS activity, reducing power, chelating effect on ferrous ion and total phenol content of all the *Pleurotus* species along with standard.

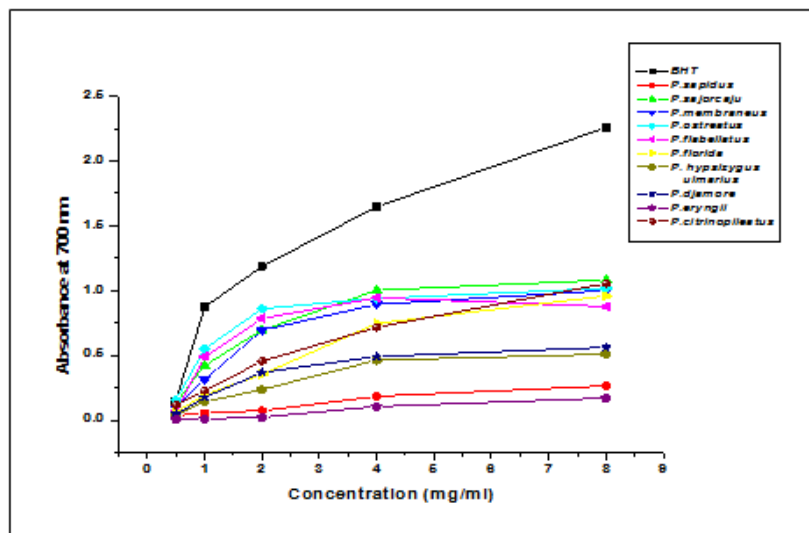
Table 1: EC₅₀ of FRS Activity, Total Phenol Content of all the *Pleurotus* species and Standard

Mushroom samples	EC ₅₀ value (mg/ml)			Total phenol (mg/g)
	Free Radical Scavenging (FRS) activity	Reducing power	Chelating effect of ferrous ions	
BHT	0.049	1.847	-	-
Ascorbic acid	0.049	-	-	-
EDTA	-	-	0.033	-
<i>P. sapidus</i> (DMRP-4)	0.352	< BHT	0.349	3.87±0.08
<i>P. sajor-caju</i> (DMRP-112)	0.498	< BHT	0.031	3.87±0.09
<i>P. membranaceus</i> (DMRP-189)	2.187	< BHT	0.104	1.80±0.05
<i>P. ostreatus</i> (DMRP-262)	0.496	< BHT	0.041	4.33±0.05
<i>P. flabellatus</i> (DMRP-5)	11.190	< BHT	0.349	0.50±0.00
<i>P. florida</i> (DMRP-88)	1.792	< BHT	0.032	2.78±0.01
<i>P. hypsizygus ulmarius</i> (DMRP-115)	4.820	< BHT	0.041	1.48±0.17
<i>P. djamora</i> (DMRP-205)	1.440	< BHT	0.274	2.03±0.03
<i>P. eryngii</i> (DMRP-135)	0.741	< BHT	0.188	2.55±0.07
<i>P. citrinopileatus</i> (DMRP-10)	1.910	< BHT	0.052	1.25±0.01

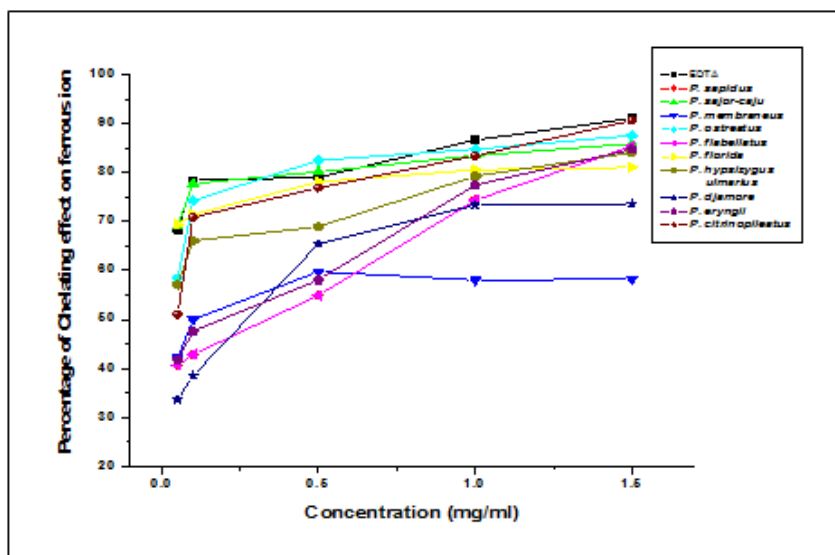
All values are expressed as Mean ± SD (n = 3). Values bearing different letters are significant t-test at P<0.05.



Graph 1: Free Radical Scavenging (FRS) activity of methanolic extract of mycelial mushroom on DPPH at various concentrations. Values express as mean ± SD of triplicate experiments.



Graph 2: Reducing power of methanolic extract of mycelial mushrooms at various concentration. Values express as mean ± SD of triplicate experiments.



Graph 3: Chelating effect on ferrous ion of methanolic extract of mycelial mushrooms at various concentrations. Values express as mean \pm SD of triplicate experiments.

The DPPH scavenging activities of methanol extract was more effective in comparison to ethyl acetate and hot water extract¹⁶. Tsai¹⁷, Mau et al.¹⁸ at 10 mg/ml, the methanolic extracts of *Agrocybe cylindracea*, *Ganoderma tsugae* and *Morchella esculenta* mycelia scavenged 91.4%, 95.6% and 94.1% of DPPH radicals, respectively and showed close similarity with the finding of *P. sajor-caju*. Kalyoncu et al.¹⁹ found that ethanolic extract of *Pleurotus eryngii* and *Pleurotus ostreatus* had potent free radical scavenging activity. The Free Radical Scavenging (FRS) activity of standard was higher than all mushroom extracts, which was reported previously^{8,18}. The finding showed that the antioxidant activity of mushroom was strongly correlated with the phenolic content which was similar with the work of Velioglu et al.²⁰. FRS ability of *P. djamor* was 76.31% at 8 mg/ml concentration which closely matched with the finding of Lee et al.²¹ on *Hypsizigus marmoreus* mycelium. *P. ostreatus* showed highest FRS activity in different organic and inorganic extract was ranged 58.76 % to 89.93 % at a concentration of 20 mg/ml³. Crude Extract from *Lentinus squarrosulus* mycelial culture showed highest FRS activity of 86 % at 20 mg/ml concentration²². Highest percentage of DPPH scavenging activity at 4 mg/ml concentration of *Pleurotus pulmonarius* showed 87.17 \pm 0.86; *Pleurotus eryngii* showed 75.00 \pm 1.88 and EC₅₀ was 1.21, 2.67 mg/ml, respectively²³ but Ferreira et al.²⁴ found EC₅₀ value of *Pleurotus eryngii* was 9.21 mg/ml. Somasundaram et al.²⁵, reported that IC₅₀ value of water and methanolic extract of *P. sajor-caju* was 1.80 and 2.50 mg/ml, where as in *P. djamor* it was 3.00 and 1.90 mg/ml, respectively. The EC₅₀ value of DPPH radical of methanol extract of *P. eous* was 4.2 mg/ml¹⁶. The findings showed similarity with those of *P. djamor*, *P. florida* and *P. membranaceus*. Synthetic antioxidants showed higher inhibition in comparison to test samples which showed similarity with this finding²⁶. The synthetic antioxidants ascorbic acid at 0.5 mg/ml, gave the scavenging activity of 90.93 \pm 2.16% with the IC₅₀ values of 0.09 \pm 0.01 mg/ml. This result revealed that methanolic extract of the mushrooms mycelia were free radical scavengers and possibly acting as primary antioxidant. However, the active components in the mycelial extract was responsible for the observed antioxidant activity, are unknown. Further work is

necessary on the isolation and purification of the active components.

Reducing Power Assay

The assay of reducing power, the antioxidant compounds convert into the oxidized form of iron in ferricyanide (Fe³⁺) ion to ferrous (Fe²⁺) ion. The reducing power is related to its electron transfer ability and may serve as a significant indicator of potential antioxidant activity. Here, the change of colour of the test solution from yellow to green is dependent on the reducing power of the sample and absorbance at 700 nm indicates greater reducing power ability.

Reducing powers of methanolic extracts from mushrooms mycelia were excellent and increased steadily increased with lower to higher concentrations (0.5-8 mg/ml), which were shown in Graph 2. The highest reducing powers were found in *P. sajor-caju* (1.082 \pm 0.03), *P. citrinopileatus* (1.054 \pm 0.01), *P. ostreatus* (1.017 \pm 0.02) and *P. eryngii* (1.000 \pm 0.08) at 8 mg/ml concentration. Out of ten species, *P. djamor* showed lowest reducing power of 0.171 \pm 0.03 at highest concentration. However, the reducing power of BHT maintained the level of 0.153 to 2.258 at the concentration of 0.5 to 8.0 mg/ml. The significant ($p < 0.05$) EC₅₀ values of BHT were 1.847 mg/ml of concentration which were more than the maximum values of *Pleurotus* species.

Jeena et al.²⁷ found that cultivated species of *P. sajor-caju* and *P. ostreatus* had an excellent reducing power of 1.980 at 10 mg/ml and 1.780 at 10 mg/ml concentration respectively, which showed close similarity with our finding but *P. sapidus* showed 1.970 at 10 mg/ml concentration which was much more than our finding. The reducing powers of ethyl acetate, methanolic and hot water extracts of *P. eous* showed an excellent reducing power of 1.950, 1.132, and 1.632 at 10 mg/ml, respectively¹⁶. According to Arbaayah & Kalsom²⁸, the greatest ability of ethanolic extract for reducing the ferricyanide complex to ferrous form was observed in different flush of cultivated *P. djamor* var. *djamor* was 0.47-1.23, *P. djamor* var. *roseus* was 0.48-0.96, *P. pulmonarius* was 0.35-0.53 and *P. ostreatus* was

0.29-0.57, respectively at concentration 10 mg/ml. However, the reducing power of BHT (2.258±0.09) at 8 mg/ml concentration which was much higher than the *Pleurotus* species and it also indicated that the synthetic antioxidant had superior reducing power ability compared to antioxidant from *Pleurotus* species²⁷.

Chelating Effect on Ferrous Ion

Chelating effects of methanolic extract from mycelia of ten *Pleurotus* species on ferrous ions increased with increased concentration except *P. membranaceus* (Graph 3). This assay analyzed that mycelial extract of *Pleurotus* species interfered with the formation of ferrous and ferrozine complex, suggesting their ability of chelating activity. Chelating effects of ferrous ion were highest in EDTA of 91.11% in comparison to *Pleurotus* species and lowest in *P. membranaceus* (59.78±2.21%) at 1.5 mg/ml concentration.

Metal ion chelating capacity played a significant role in antioxidant mechanism²⁹. Since ferrous ion were the most effective pro-oxidants in food systems³⁰, the higher chelating effect of methanolic extracts from mycelial mushrooms would be more effective in comparison to DPPH and reducing power method. However, Yen & Wu³¹ used the method of Decker and Welch¹⁴ to determine the chelating effect instead of the method of Shimada et al.¹². Methanolic extracts from ear mushrooms were good chelators for ferrous ions of 85.1–96.5% at 5 mg/ml concentration³². As compared with wild and commercial³³ mushrooms, Omethanolic extracts from mycelial mushrooms would be good chelators for ferrous ions at higher concentrations. The highest chelating activity of 74.88% was found in macerated ethanol extracts of *P. eryngii* with the IC₅₀ value of 1.00 mg/ml³⁴.

Total Phenolic Content

Total phenol content was highest in *P. ostreatus* (4.33±0.05 mg/g) and lowest in *P. flabellatus* (0.50±0.00 mg/g) showed in Table 1. The mycelia of *Pleurotus* species contained phenolic compounds. It can therefore be concluded that these phenolic components might be involved in these antioxidative properties as mentioned in previous studies³. Many studies on spices, vegetables, fruits, and plants extracts had shown a good relationship between phenolic content and antioxidant activity³⁵. According to published data with respect to their relationships between the concentration of phenolic compounds and the antioxidant activity there was much controversy.

CONCLUSION

On basis of the findings, it showed that *Pleurotus* species had low reducing power, moderate free radical scavenging activity and chelating effect of ferrous ion. But *Pleurotus* species had high chelating effect of ferrous ion in comparison to free radical scavenging activity. Out of ten *Pleurotus* species *P. sapidus*, *P. sajor-caju*, *P. ostreatus*, *P. eryngii* and *P. citrinopileatus* showed excellent antioxidant properties. Therefore, on the basis of the outcomes of study it can be concluded that the consumption of mushrooms i.e., *Pleurotus* mushrooms, their assumed antioxidant properties might be beneficial to the antioxidant protection system of the human body against oxidative damage.

The present comparative study of antioxidant activity of different species of *Pleurotus* documented for the first time. This result revealed that methanolic extract of the mushrooms mycelia were free radical scavengers possibly by acting as primary antioxidant. Thus *Pleurotus* mushroom consumers

might be benefited by protecting themselves from oxidative damaged. However, the active components of the mycelial extract responsible for the antioxidant activity are unknown. Further work is necessary on the isolation and purification of the active components.

ACKNOWLEDGEMENTS

The authors are grateful to the Head, Department of Botany for providing all sorts of facilities. The authors are also thankful to DMR, Chambaghat, Solan, India for providing the mushroom cultures. The first author is thankful to the DBT, Government of India for the financial assistance.

REFERENCES

1. Jose N, Janardhanan KK. Antioxidant and antitumor activity of *Pleurotus florida*. Current Science 2000; 79(7): 941-943.
2. Chang ST. In: Arora DK, Mukerji KG, Marth EH. (Eds). Hand Book of Applied Mycology. New York: Marcel Dekker. Inc.; 1991: 221-240.
3. Vamanu E. Biological activities of the polysaccharides produced in submerged culture of two edible *Pleurotus ostreatus* mushrooms. Journal of Biomedicine and Biotechnology 2012; 2012: 1-8.
4. Simic MG. Mechanisms of inhibition of free-radical processed in mutagenesis and carcinogenesis. Mutation Research 1988; 202: 377-386.
5. Cazzi R, Ricardy R, Aglitti T, Gatta V, Petricone P, De Salvia R. Ascorbic acid and β-carotene as modulators of oxidative damage. Carcinogenesis 1997; 18: 223-228.
6. Sanmee R, Dell B, Lumyong P, Izumori K, Lumyong S. Nutritive value of popular wild edible mushrooms from northern Thailand. Food Chemistry 2003; 82: 527-532.
7. Isildak Ö, Turkecul I, Elmastas M, Tuzen M. Analysis of heavy metals in some wild-grown edible mushrooms from the middle Black Sea region, Turkey. Food Chemistry 2004; 86: 547-552.
8. Elmastas M, Isildak O, Turkecul I, Temur N. Determination of antioxidant activity and antioxidant compounds in wild edible mushrooms. Journal of Food Composition Analysis 2007; 20: 33-45.
9. Saha, AK. Studies on some aspects of mushrooms of hilly tracts and plains of West Bengal. Ph. D Thesis, University of Calcutta, Kolkata, India; 1985: 7-8.
10. Kalyoncu F, Oskay M, Kayalar H. Antioxidant activity of the mycelium of 21 wild mushroom species. Mycology 2010; 1(3): 195-199.
11. Mau LJ, Huang PN, Hung SJ, Chen CC. Antioxidant property of methanolic extracts from two kinds of *Antrodia camphorata* mycelia. Food Chemistry 2004; 86: 25-31.
12. Shimada K, Fujikawa K, Yahara K, Nakamura T. Antioxidative properties of xanthan on the autoxidation of soybean oil in cyclodextrin emulsion. Journal of Agricultural and Food Chemistry 1992; 40(6): 945-948.
13. Oyaizu M. Studies on products of browning reactions: antioxidative activities of products of browning reactions prepared from glucosamine. Japanese Journal of Nutritional Science 1986; 44: 307-315.
14. Decker EA, Welch B. Role of ferritin as a lipid oxidation catalyst in muscle food. Journal of Agricultural and Food Chemistry 1990; 38: 674-677.
15. Swain T, Hillis WE. The phenolic constituents of *Purmus domestica*. I. The quantitative analysis of phenolic constituents. Journal of the Science of Food and Agriculture 1959; 10: 63-68.

16. Sudha G, Vadivukkarasi S, Shree RBI, Lakshmanan P. Antioxidant Activity of Various Extracts from an Edible Mushroom *Pleurotus eous*. Food Science and Biotechnology 2012; 21(3): 661-668.
17. Tsai SY. Antioxidant properties and their cytotoxic activities on tumor cells of *Ganoderma tsugae* and *Agrocybe cylindracea* and antimutagenic properties of *Agrocybe cylindracea* [M.Sc thesis]. National Chung Hsing University, Taichung, Taiwan; 2002.
18. Mau JL, Chang CN, Huang SJ, Chen CC. Antioxidant properties of methanolic extracts from *Grifola frondosa*, *Morchella esculenta* and *Termitomyces albuminosus* mycelia. Food Chemistry 2004; 87: 111-118.
19. Kalyoncu F, Oskay M, Saglam H, Erdogan TF, Tamer A. U. Antimicrobial and Antioxidant Activities of Mycelia of 10 Wild Mushroom Species. Journal of Medicinal Food 2010; 13 (2): 415-419.
20. Velioglu YS, Mazza G, Gao L, Oomah BD. Antioxidant activity and total phenolics in selected fruits, vegetables, and grain products. Journal of Agricultural and Food Chemistry 1998; 46: 4113-4117.
21. Lee YL, Jian SY, Lian PY, Mau JL. Antioxidant properties of extracts from a white mutant of the mushroom *Hypsizygus marmoreus*. Journal of Food Composition and Analysis 2008; 21: 116-124.
22. Ahmad R, Muniandy S, Shukri NIA, Alias SMU, Hamid AA, Yusoff WMW, Senafi S, Daud F. Antioxidant Properties and Glucan Compositions of Various Crude Extract from *Lentinus squarrosulus* Mycelial Culture. Advances in Bioscience and Biotechnology 2014; 5: 805-814.
23. Xu WW, Lai ETC, Chen L, Huang JJH, Cheung ALM, Cheung PCK. Water Extract from *Pleurotus pulmonarius* with Antioxidant Activity Exerts In Vivo Chemoprophylaxis and Chemo sensitization for Liver Cancer. Nutrition and Cancer 2014; 66(6): 989-998.
24. Ferreira ICFR, Reis FS, Barros L, Sousa MJ, Martins A. Analytical Methods Applied to the Chemical Characterization and Antioxidant Properties of Three Wild Edible Mushroom Species from Northeastern Portugal. Food Analytical Methods 2014; 7: 645-652.
25. Somasundaram R, Puttaraju NG, Venkateshaiah SU, Dharmesh SM, Urs SMN. Antioxidant Activity of Indigenous Edible Mushrooms. Journal of Agricultural and Food Chemistry 2006; 54: 9764-9772.
26. Woldegiorgis AZ, Abate D, Haki GD, Ziegler GR. Antioxidant property of edible mushrooms collected from Ethiopia. Food Chemistry 2014; 157: 30-36.
27. Jeena GS, Punetha H, Prakash O, Chandra M, Kushwala KPS. Study of invitro antioxidant potential of some cultivated *Pleurotus* species (Oyster Mushrooms). Indian Journal of Natural Product and Resources 2014; 5(1): 56-61.
28. Arbaayah HH, Kalsom YU. Antioxidant properties in the oyster mushrooms (*Pleurotus* spp.) and split gill mushroom (*Schizophyllum commune*) ethanolic extracts. Mycosphere 2013; 4 (4): 661-673.
29. Duh PD, Tu YY, Yen GC. Antioxidant activity of water extract of Harnng Jyur (*Chrysanthemum morifolium* Ramat). LWT-Food Science and Technology 1999; 32: 269-277.
30. Yamaguchi R, Tatsumi MA, Kato K, Yoshimitsu U. Effect of metal salts and fructose on the autoxidation of methyl linoleate in emulsions. Agricultural and Biological Chemistry 1988; 52: 849-850.
31. Yen GC, Wu JY. Antioxidant and radical scavenging properties of extracts from *Ganoderma tsugae*. Food Chemistry 1999; 65: 375-379.
32. Mau JL, Chao GR, Wu KT. Antioxidant properties of methanol extracts from several ear mushrooms. Journal of Agricultural and Food Chemistry 2001; 49: 5461-5467.
33. Yanga JH, Linb HC, Mau JL. Antioxidant properties of several commercial mushrooms. Food Chemistry 2002; 77: 229-235.
34. Yildirim NC, Turkoglu S, Yildirim N, Ince OK. Antioxidant properties of wild edible mushroom *Pleurotus eryngii* collected from tunceli province of turkey. Digest Journal of Nanomaterials and Biostructures 2012; 7: 1647-1654.
35. Dudonn'e S, Vitrac X, Couti'ere P, Woillez M, M'errillon JM. Comparative study of antioxidant properties and total phenolic content of 30 plant extracts of industrial interest using DPPH, ABTS, FRAP, SOD and ORAC assays. Journal of Agricultural and Food Chemistry 2009; 57(5): 1768-1774.

Cite this article as:

Sanjit Debnath et al. Antioxidant activities of methanolic extracts from ten *Pleurotus* species. Int. Res. J. Pharm. 2017;8(3):44-49 <http://dx.doi.org/10.7897/2230-8407.080335>

Source of support: DBT, Government of India, Conflict of interest: None Declared

Disclaimer: IRJP is solely owned by Moksha Publishing House - A non-profit publishing house, dedicated to publish quality research, while every effort has been taken to verify the accuracy of the content published in our Journal. IRJP cannot accept any responsibility or liability for the site content and articles published. The views expressed in articles by our contributing authors are not necessarily those of IRJP editor or editorial board members.