

INTERNATIONAL RESEARCH JOURNAL OF PHARMACY

www.irjponline.com ISSN 2230 - 8407

Research Article

ESTIMATION OF BIO-ACTIVE COMPOUNDS IN INDIAN DESERTED PLANT EUPHORBIA CADUCIFOLIA (DANDA THOR) BY GAS CHROMATOGRAPHY-MASS SPECTROMETRY

Premlata Singariya *1, Krishan Kumar Mourya 2 and Padma Kumar 1

¹UGC Post-doctoral fellow, Laboratory of Tissue Culture and Secondary Metabolites, Department of Botany, University of Rajasthan, Jaipur, Rajasthan, India

²Assistant Director, Rural Veterinary Polyclinic, Hingonia, Jaipur, Rajasthan, India

Article Received on: 09/05/18 Approved for publication: 28/05/18

DOI: 10.7897/2230-8407.09695

ABSTRACT

Objective: The investigation was carried out to determine the possible bioactive components of methonolic extracts of *Euphorbia caducifolia* (whole plant) using Gas chromatography-Mass spectrometry (GC-MS). Experimental design: All the samples were dried in shade till constant weight was achieved. They were then macerated to powder form with a mixer grinder. The powder was stored in air sealed polythene bags at room temperature before extraction. The chemical compositions of the methonolic extracts of *Euphorbia caducifolia* (whole plant) were investigated using Thermo G C 1300 and "TSQ 8000 "Triple quadrupole GC-MS MS SYSTEM with auto sampler Al 1310 Gas chromatography-Mass spectrometry, while the mass spectra of the compounds found in the extract was matched with the National Institute of Standards and Technology (NIST) library. Observation and Conclusion: GC-MS analysis of the extract reveals the identification of forty four compounds. This is the first report of identification of components from the whole plant of *E. caducifolia* by GC-MS. Most of the compounds in the list are bioactive and possess medicinal properties.

Key words: Euphorbia caducifolia, Gas chromatography-Mass spectrometry, bioactive components.

INTRODUCTION

Taking into consideration of the medicinal importance of this plant, the methonolic extracts of *Euphorbia caducifolia* (whole plant) were analyzed for the first time using Gas chromatography-Mass spectrometry (GC-MS). This work will help to identify the compounds of therapeutic value. GC-MS is one of the best techniques to identify the bioactive constituents of long chain, branched chain hydrocarbons, alcohols, acids, ester, steroids, phenolic compounds etc^{1, 2, 3, 4}.

Various plants are used in traditional treatments to cure variety of diseases. In the last few decades there has been an exponential growth in the field of herbal medicine. Natural products have been a source of drugs for centuries. In the present study methonolic extracts of *E. caducifolia* were analyzed by GC-MS technique to study the major and minor phyto-constituents of the vegetative parts of the whole plant.

The ecological life-history of "leafless" spurge, *E. caducifolia* Haines, which serves as an indicator of rocky and gravelly habitat in Rajasthan (India) is dealt with here. Root decoction is used as an effective abortifacient at initial stages⁵ and dried stem is burnt to produce smoke, affected painful body part is kept in smoke for some time to relive pain⁶. There appear to be two forms in existence: one with red and the other with green inflorescence, which interbreed freely in nature resulting in different shades of red coloured inflorescence. The plant is suitably adapted to xerophytic conditions, but surprisingly the leaves are borne only in summer when there is acute shortage of water⁷.

MATERIAL AND METHOD

Plant material: Euphorbia caducifolia were collected in the month of August 2015 from the Central Arid Zone Research Institute (CAZRI), Jodhpur (Rajasthan). Plants samples were identified and deposited in the herbarium, Department of Botany, University of Rajasthan, Jaipur. The collected plant materials were transferred immediately to the laboratory cleaned with water and selected plant parts were separately shade dried⁸ until weight has been constant.

Preparation of plant extracts: The collected plant materials were shade dried till constant weight was achieved⁹. The plant material were powered with the help of grinder¹⁰ and passed through 40mm meshes and stored in clean container for further use¹¹. The dried powder material was extracted with methanol by using the Soxhlet apparatus¹² for 18 hours at a temperature not exceeding the boiling point of the respective solvent^{13, 14}. The obtained extracts were filtered by using Whatman No. 1 filter paper and then concentrated at 40°C by using an evaporator¹⁵ and stored the residual extracts in refrigerator at 4°C in small and sterile amber colour glass bottles¹⁶ for subsequent use in the further antimicrobial, anti-fungal and phyto-chemical analysis. The extract contains both polar and non-polar phyto-components.

Gas chromatography-Mass spectrometry analysis: Gas chromatography-Mass spectrometry (GC-MS) analysis of these extracts was carried out by following the method¹⁷. The GC-MS analysis of the extracts was performed using a GC-MS Thermo G C 1300 and "TSQ 8000 "Triple quadrupole GCMSMS SYSTEM with auto sampler Al 1310. Gas Chromatography 1300 with a fused GC column TG-5MS AMINE. The column

^{*}Corresponding Author Email: premlatasingariya@gmail.com

length was 30 m with internal diameter; coated film 0.25 μm with flow rate 10 ml/m in and the condition were as follows: PTV Temp. Program: 70 °C, hold 1.00 min, 10 °C/min to 280 °C, hold 15 min. Carrier gas helium flow rate 1ml/min, split ratio 1:50. GC is equipped with auto-sampler AI 1300 and sample volume was 1μ liter. The elutes were automatically passed into a mass spectrometer. GC mass Spectrum analysis was conducted using TSQ8000 with transfer line temperature 270°C and ion source temperature 230°C in EI mode. Mass scan time was 4 min with full Scan MS. The mass spectrum was also equipped with a computer fed NIST mass Spectra data library.

Identification of Components: Interpretation on mass spectrum of GC-MS was done using the database of National Institute of standard and Technology NIST-08 LIB^{18, 19} and WILEY-8 LIB^{20, 21} library sources were used for matching the identified components from the plant material having more than 62,000 patterns^{22, 23}. The mass spectrum of the unknown component was compared with the spectrum of the known components stored in the NIST library^{24, 25}. The name, molecular weight and structure of the components of the test materials were ascertained²⁶.

Table-1: Total Bio-active compounds of Euphorbia caducifolia by Gas Chromatography- Mass Spectrometry

S. No.	RT	Compound Name	Area	Area %	RSI
1.	4.46	Tetrazolo [1,5-b]pyridazine, 6-chloro-7-methyl-	71716	2.77	859
2.	4.61	1-Phenyl-3,4-dimethyl-4-nitroso-2-pyrazolin-5-one	40496	1.57	683
3.	5.06	Pyrazinamine, 3,4,5,6-tetrahydro-6-imino-N-phenyl-4-(phenylmethyl)-	56464	2.18	909
4.	5.21	8,11 Octadecadiynoic acid, methyl ester	23257	0.90	767
5.	5.25	Sydnone, 3(phenylmethyl)	37550	1.45	830
6.	5.35	Hydrazine, phenylsulfinyl	38604	1.49	797
7.	5.53	1-H Imidazole1ethanol, 2-methylàphenyl	54253	2.10	800
8.	6.24	Sydnone, 3(phenylmethyl) Benzene, 1 nitro2(pmethylphenoxy) 4fluoro	29507	1.14	812
9.	6.51	2,2,2Trichloro1 (2nitrophenylthioamino) ethanol	38275	1.48	787
10.	6.55	2,2-Diphenyl-1,3,6-trioxa-2-sila-cyclooctane	216070	8.36	731
11.	6.68	1,3,5Triazine,1,3,5tricyclohexylhexahydro	16352	0.63	789
12.	8.10	3,7-Decadien-2-one, 10-(3,3-dimethyloxiranyl)-4,8-dimethyl-, (E,E)-ñ-	26676	1.03	743
13.	9.54	Acetic acid, (2,5-dichloro-4-methoxyphenoxy)-, methyl ester	101191	3.91	782
14.	9.83	Manganese, tricarbonyl[(1,2,3,4,5-ü)-1-[(dimethylamino) sulfonyl]-2-(1- hydroxy-1-methylethyl)-2,4-cyclopentadien-1-yl]-	57707	2.23	715
15.	12.09	Diazene, 1-ethyl-2-phenyl-, 1-oxide, (Z)-	76739	2.97	734
16.	12.46	à-D-Xylofuranoside, methyl 5-O-methyl-	58169	2.25	622
17.	12.68	Pyrazinamine, 1,2,3,4Oxatriazolium,5(ethylamino)3phenyl,hydroxide, inner salt	32202	1.25	745
18.	13.00	Tetrazolo[1,5b] pyridazine, 8methyl	34575	1.34	817
19.	14.05	Benzenesulfonic acid, 2-nitro-, hydrazide	55935	2.16	620
20.	14.33	Propanedioic acid, (1-cyclohexen-1-ylmethyl)-, dimethyl ester	37512	1.45	529
21.	15.45	3-Hexene, 1-(1-ethoxyethoxy)-, (Z)-	94449	3.65	806
22.	15.88	Tricyclo[4.3.1.1(3,8)]undecane, 1-bromo-	34567	1.34	592
23.	17.45	1,3-Cyclopentanedione, 4-butyl-	36298	1.40	599
24.	17.78	Copper, bis(4chloro3,5cyclohexadiene1,2dione 2oximatoN2, O1) (pyridine)	47456	1.84	665
25.	18.82	5-Chloro-2,4-dithiahexane 2,2-dioxide	53845	2.08	623
26.	19.79	Oxetane, 2,2,3,3-tetramethyl-4,4-diphenyl-	42651	1.65	768
27.	19.82	1,1-(Phenylthio)-2-isopropyl-cyclobutane	62578	2.42	636
28.	20.18	4-Octene, 2,3,7-trimethyl-, [S-(E)]-	47879	1.85	703
29.	21.23	Chromium, tricarbonyl[(1,2,3,4,5,6\u00fc) 1,3,5,7cyclo octatetraene]	42201	1.63	792
30.	22.52	Phthalic acid, decyl phenyl ester	71547	2.77	771
31.	23.20	Tricyclo[2.2.1.0(1,4)]heptan2one,6nitro	35882	1.39	706
32.	26.19	Cyclopentane, nonyl-	36721	1.42	803
33.	26.44	Methyl N-cyclohexyl-3-phenylpropanimidate	31363	1.21	689
34.	26.76	2Dodecen1ol, 12chloro	80740	3.12	796
35.	26.94	NVinylpyridiniumbromide	54809	2.12	825
36.	28.12	2,4-Dimethyl-6-phenyl-3-thioxo-5-oxo-2,3,4,5-tetrahydro-1,2,4-triazine	113011	4.37	577
37.	30.56	3,5,8Trimethyl1phenyl4,5,6,8tetrahydropyrazolo[3,4b][1,4]diazepin7 (1H)one#	55720	2.16	765
38.	30.72	4Methyl2,4bis(4'trimethylsilyloxyphenyl)pentene1	121940	4.72	740
39.	30.79	3 Phenyl4benzoyl7mercaptomethyl2,6dioxa3azabicyclo [3.3.0] 7octene	40035	1.55	806
40.	31.02	1H1,2,3Triazol1amine, N[(4methoxyphenyl) methylene]4,5dimethyl	99516	3.85	771
41.	31.56	Iron, tricarbonyl[(1,2,3,4\overline{u}) 7methylene1, 3,5cycloheptatriene]	71787	2.78	799
42.	31.61	Trimethyl [4(1,1,3,3,tetramethylbutyl) phenoxy]silane	104508	4.04	782
43.	31.75	2Butanone, 3chloro4hydroxy1,4diphenyl	38394	1.49	785
44.	31.80	4Methyl2,4bis(4'trimethylsilyloxyphenyl)pentene1	63560	2.46	784

Table-2: Major Bio-active compounds of Euphorbia caducifolia by Gas Chromatography- Mass Spectrometry

S. No.	RT	Compound Name	Area %				
	Major constituents						
1.	6.55	2,2-Diphenyl-1,3,6-trioxa-2-sila-cyclooctane	8.36				
2.	30.72	4Methyl2,4bis(4'trimethylsilyloxyphenyl)pentene1	4.72				
3.	28.12	2,4-Dimethyl-6-phenyl-3-thioxo-5-oxo-2,3,4,5-tetrahydro-1,2,4-triazine	4.37				
4.	31.61	Trimethyl [4(1,1,3,3,tetramethylbutyl) phenoxy]silane	4.04				
5.	9.54	Acetic acid, (2,5-dichloro-4-methoxyphenoxy)-, methyl ester	3.91				
6.	31.02	1H1,2,3Triazol1amine, N[(4methoxyphenyl) methylene]4,5dimethyl	3.85				
7.	15.45	3-Hexene, 1-(1-ethoxyethoxy)-, (Z)-	3.65				
8.	26.76	2Dodecen1ol, 12chloro	3.12				
9.	12.09	Diazene, 1-ethyl-2-phenyl-, 1-oxide, (Z)-	2.97				
10.	31.56	Iron, tricarbonyl[(1,2,3,4\u00fc) 7methylene1, 3,5cycloheptatriene]	2.78				

Table-3: Minor Bio-active compounds of Euphorbia caducifolia by Gas Chromatography- Mass Spectrometry

S. No.	RT	Compound Name	Area %				
Minor constituents							
1.	17.45	1,3-Cyclopentanedione, 4-butyl-	1.40				
2.	23.20	Tricyclo[2.2.1.0(1,4)]heptan2one,6nitro	1.39				
3.	13.00	Tetrazolo[1,5b] pyridazine, 8methyl	1.34				
4.	15.88	Tricyclo[4.3.1.1(3,8)]undecane, 1-bromo-	1.34				
5.	12.68	Pyrazinamine, 1,2,3,4Oxatriazolium,5(ethylamino)3phenyl,hydroxide, inner salt	1.25				
6.	26.44	Methyl N-cyclohexyl-3-phenylpropanimidate	1.21				
7.	6.24	Sydnone, 3(phenylmethyl) Benzene,1nitro2(pmethylphenoxy) 4fluoro	1.14				
8.	8.10	3,7-Decadien-2-one, 10-(3,3-dimethyloxiranyl)-4,8-dimethyl-, (E,E)-ñ-	1.03				
9.	5.21	8,11 Octadecadiynoic acid, methyl ester	0.90				
10.	6.68	1,3,5Triazine,1,3,5tricyclohexylhexahydro	0.63				

Dept. of USIC University of Rajasthan, Jaipur

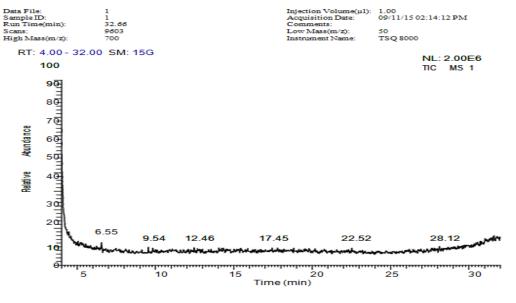
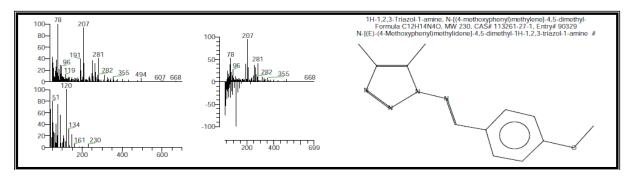
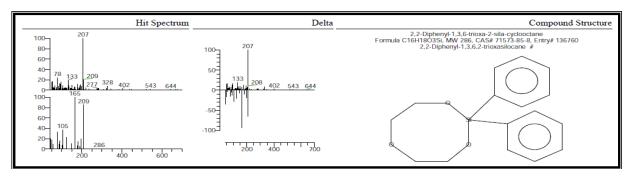


Fig. 1 Chromatogram of Methanolic extract of whole plant of Euphorbia caducifolia by GC-MS

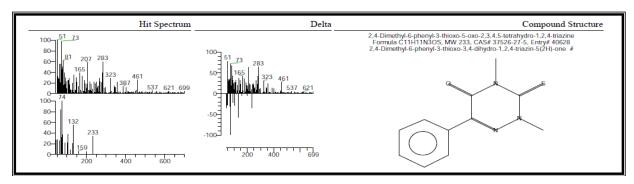


 $1H1, 2, 3Triazol1amine, \\ N[(4methoxyphenyl)\ methylene] 4, 5dimethyl$

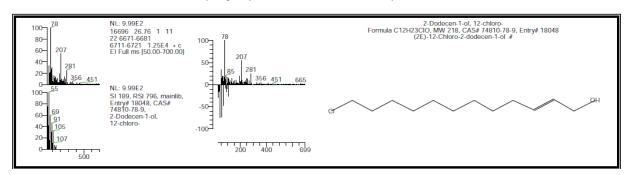
Premlata Singariya et al. Int. Res. J. Pharm. 2018, 9 (6)



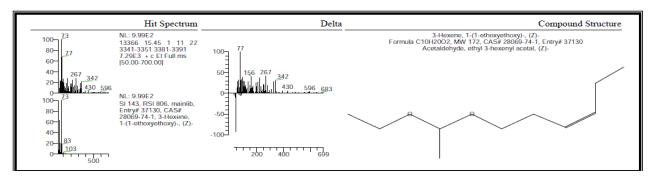
$\hbox{2,2-Diphenyl-1,3,6-trioxa-2-sila-cyclooctane}$



2,4-Dimethyl-6-phenyl-3-thioxo-5-oxo-2,3,4,5-tetrahydro-1,2,4-triazine

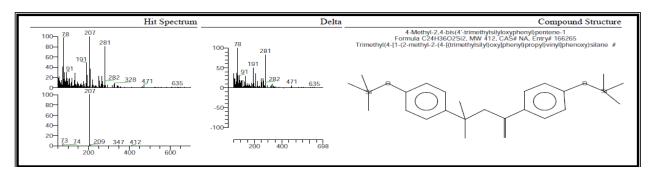


2Dodecen1ol, 12chloro

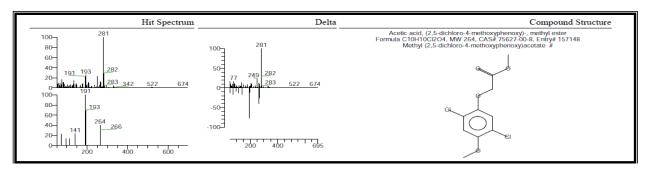


3-Hexene, 1-(1-ethoxyethoxy)-, (Z)-

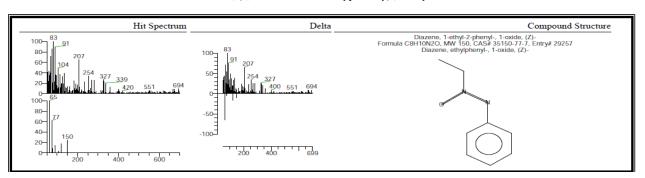
Premlata Singariya et al. Int. Res. J. Pharm. 2018, 9 (6)



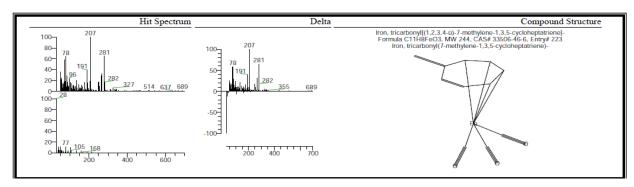
4 Methyl 2, 4 bis (4'trimethyl silyloxyphenyl) pentene 1



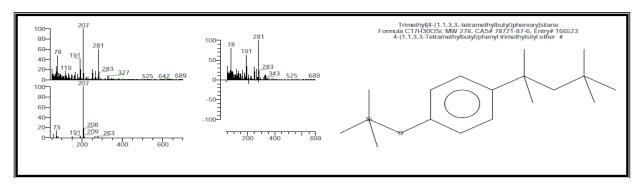
Acetic acid, (2,5-dichloro-4-methoxyphenoxy)-, methyl ester



Diazene, 1-ethyl-2-phenyl-, 1-oxide, (Z)-



 $Iron, tricarbonyl [(1,2,3,4\ddot{u})\ 7methylene 1, 3,5 cycloheptatriene]$



Trimethyl [4(1,1,3,3,tetramethylbutyl) phenoxy|silane

Fig. 2 The best hit for the prevailing compounds in the chromatogram.

RESULTS AND DISCUSSION

The active principles with their retention time (RT), molecular formula (MF), molecular weight (MW) and concentration (%) in the methanolic extracts of the whole plant of *Euphorbia caducifolia* are presented in tables 1 followed by^{27, 28}. The GC-MS analysis of the extracts showed the presence of phytocomponents, the phyto-components of the above said plant extract are presented in Table-1 and the GC-MS chromatogram with peak area of each extract is also given²⁹ (figure-1). Totally 44 bio-active constituents were identified in the present study from the acetone extracts of the whole plant of *E. caducifolia* which including both major and minor constituents.

The major constituents were 2,2-Diphenyl-1,3,6-trioxa-2-sila-(8.36%); cvclooctane 4 Methy 12. (4'trimethylsilyloxyphenyl) pentenel (4.72%); 2, 4-Dimethyl-6phenyl-3thioxo-5oxo-2,3,4,5-tetrahydro-1,2,4-triazine (4.37%); Trimethyl [4(1,1,3,3,tetramethylbutyl) phenoxy] silane (4.04%); Acetic acid, (2,5-dichloro-4-methoxyphenoxy)-, methyl ester (3.91%); 1H1,2,3Triazol1amine, N[(4methoxyphenyl) methylene]4,5dimethyl (3.85%); 3-Hexene, 1-(1-ethoxyethoxy)-, (Z)- (3.65%); 2Dodecen1ol, 12chloro (3.12%); Diazene, 1ethyl-2-phenyl-, 1-oxide, (Z)- (2.97%) and Iron, tricarbonyl [(1,2,3,4\u00fc) 7methylene1, 3,5cycloheptatriene] (2.78\u00db) (table-2). Minor constituents were 1,3-Cyclopentanedione, 4-butyl-(1.40%); Tricyclo [2.2.1.0 (1,4)] heptan 2 one, 6 nitro (1.39%); Tetrazolo[1,5b] pyridazine, 8 methyl (1.34%); Tricyclo [4.3.1.1(3,8)] undecane, 1-bromo- (1.34%); Pyrazinamine, 1,2,3,4 Oxatriazolium, 5(ethylamino) 3phenyl, hydroxide, inner salt (1.25%); Methyl N-cyclohexyl-3- phenylpropan imidate (1.21%); Sydnone, 3 (phenylmethyl) Benzene, 1nitro2 (pmethylphenoxy) 4 fluoro (1.14%); 3,7-Decadien-2-one, 10-(3,3dimethyloxiranyl)-4,8-dimethyl-, (E,E)-ñ- (1.03%); 8,11 Octa decadivnoic acid. methvl ester (0.90%)and 1,3,5Triazine,1,3,5tricyclohexyl hexahydro (0.63%) (table-3). The best hit for the prevailing compounds in the chromatogram (Fig. 2).

The GC-MS chromatogram with peak area has shown in fig-1. The aim of the present study is to provide more information about the essential phyto-constituents of *E. caducifolia*. The results from the present investigation were very encouraging and indicates that this plant should be studied more extensively to explore its potential to use as plant medicinal nutritive.

CONCLUSION

Therapeutic mechanism of a plant can be better understood with a proper investigation of its active ingredients. In the present study, 44 components from the methanolic extracts of the whole plant of *E. caducifolia* were identified by GC-MS analysis. The presence of various bioactive compounds justifies the use of this plant for various ailments by traditional practitioners. These active principles provide inspiration for further investigation to achieve lead molecules in the discovery of novel herbal drugs. However, isolation of individual photochemical constituents and subjecting it to biological activity will definitely give fruitful results. It could be concluded that, *E. caducifolia* contains various bioactive compounds. So it is recommended as a plant of phytopharmaceutical importance. However, further studies are needed to undertake its bioactivity and toxicity profile.

There are so many type of genes and proteins present in desertic plants so it can grow easily in stress condition^{30,31}. At the cellular level, plant cell responds to these stresses by the activation of cascades of molecular mechanisms involved in stress perception, signal transduction and the expression of specific stress related genes and metabolites. A number of genes are induced by exposure to such condition, those that protect against environmental stresses directly³².

ACKNOWLEDGEMENT

Authors are expressing their thanks to UGC for providing the funds for the project under Post-doctoral fellowship scheme and also thankful to Department of USIC, University of Rajasthan, Jaipur for providing Gas chromatography-Mass spectrometry (GC-MS) facility.

REFERENCES

- Singariya P, Mourya KK, Kumar P. Evaluations of Organic Compounds in Ethyl Acetate extracts of Marwar Dhaman By gas chromatography- mass spectrometry. J of Plant Sci. 2017b; 33(1): 17-27.
- Singariya P, Mourya KK, Kumar P. Identification of Bioactive components of Isopropyl alcohol extract of Bird wood grass using gas chromatography- mass spectrometry. Biotech in crop improvement, Chapter-10, 2016c; p. 118-130, Ed. PC. Trivedi. Pointer Publishers, Jaipur. ISBN 978-81-7132-8550.
- Singariya P, Mourya KK, Kumar P. Gas Chromatography-Mass Spectrometric analysis of acetone extract of *Cenchrus ciliaris* (Dhaman grass). I J of Sci and Nature. 2015b; 6(4): 652-661.
- 4. Amakrishnan N. GC-MS Analysis of *Rumex vesicarius* L. I J of Drug Development & Res. 2011; 3(2): 272-279.
- Ishtiaq MC, Khan MA. An ethnomedicinal inventory of plants used for family planning and sex disease in Samahni Valley, Pakisthan. Ind J of traditional knowledge. 2008; 7(2): 277-283.

- Meena KL, Yadav BL. Some ethnomedicinal plants of Southern Rajasthan. Ind J of traditional knowledge. 2010; 9(1): 169-172.
- Sen DN. Leafless Euphorbia on Rajasthan Rocks, India. Ecological Life-History Folia Geobotanica & Phytotaxonomica. 1968; (3) 1: 1-15 (article consists of 21 pages) Published by: <u>Springer</u> Stable. URL: <u>http://www.jstor.org/stable/4179478</u>
- Singariya P, Kumar P, Mourya KK. Estimation of Bioactivity of Arial parts of *Withania somnifera* Against the Bacterial and Fungal Microbes. I J of Pharmacy and Pharm sci. 2012l; 4(3): 553-557.
- Singariya P, Kumar P, Mourya KK. Qualitative and Pharmacological examination of Root extracts of *Withania* somnifera against Human and Plant Pathogens. Asian J of Res in Chem. 2012m; 5(6): 733-737.
- Singariya P, Kumar P, Mourya KK. Ripen Fruits of Indian Ginseng: Phyto-chemical and Pharmacological examination against Human and Plant Pathogens. I J of App Bio and Pharm Tech. 2012g; 3(2): 1-8.
- Singariya P, Kumar P, Mourya KK. Insignificant Antimicrobial Activities and Phyto-chemical screening in different extracts of Indian Ginseng. J of Pharm Negative Results. 2012o; 3(1): 41-45.
- Subramanian SS, Nagarajan S. Flavonoids of the seeds of Crotlaria retusa and C. striata. Curr Sci. 1969; 38: p. 65-68.
- Singariya P, Kumar P, Mourya KK. Absence of Antibiotic Activities of *Cenchrus setigerus* and *Cenchrus ciliaris* Seed extracts in Different Polar Solvents. J of Pharm Negative Results. 2013b; 4(1): 71-75.
- 14. Singariya P, Kumar P, Mourya KK. Evaluation of Indian Ginseng Against Different Bacteria and Fungi. Asian J of Pharm and clin Res. 2012k; 5(2): 145-148.
- Singariya P, Kumar P, Mourya KK. Evaluation of Antimicrobial Activity of Leaf extracts of Winter Cheery (Withania somnifera). I J of Pharm Tech Res. 2012p; 4(3): 1247-1253.
- Singariya P, Kumar P, Mourya KK. Screening for Antimicrobial Potency of Methanolic Extract of Indian Ginseng. I J of Pharmacy and Tech. 2012n; 4 (2): 4537-4548.
- Hema R, Kumaravel S, Gomathi S, Sivasubramaniam C. Gas Chromatography-Mass Spectroscopic analysis of *Lawsonia inermis* leaves. New York Sci J, 2010; 3: 141-143.
- Singariya P, Mourya KK, Kumar P. Identification of some bio-active compounds of iso-propyl alcohol extract of motha dhaman grass by gas chromatography-mass spectrometric analysis. Life Sci Leaflets. 2016a; 72(2): 122-135.
- Mc-Lafferly FW. Registry of mass spectral data, ed. 5, Wiley New York. 1989.
- 20. Singariya P, Mourya KK, Gadi BR. Evaluation of Microcidal and Nitrogen assimilatory enzymes activity and identification of β Sitosterol in C4 Grasses of Thar Desert. Envir Impact on Biodiversity. 2016d; p. 113-131. Editor: B. R. Bamniya and B. R. Gadi. Today & Tomorrow's Printers and Publishers, New Delhi 110 002. ISBN-p- 81-7019-547-7

- Stein SE. National Institute of Standards and Technology (NIST) Mass Spectral Database and Software, Version 3.02, USA, 1990.
- 22. Singariya P, Mourya KK, Kumar P. Identification of some bio-active compounds of ethyl acetate extract of *Cenchrus ciliaris* by gas chromatography-mass spectrometric analysis. Life Sci Bull. 2015c; 12(2): 141-148.
- 23. Singariya P, Kumar P, Mourya KK. Comparative Assessment of Bio-efficacy of Leaf Extract of Anjan grass in Polar Solvents and Compound Identification in Ethyl Acetate Extract. Hygeia J for drugs and medicines. 2012s; 4(2): 49-56.
- Singariya P, Mourya KK, Kumar P. Gas Chromatography-Mass Spectrometric Analyses of Acetone extract of Marwar Dhaman grass for bio-active compounds. Plant Arc. 2015a; 15(2): 1065-1074.
- Singariya P, Kumar P, Mourya KK. Isolation of New Steroids and Evaluation of Bio-activity of Kala Dhaman Grass (*Cenchrus setigerus*). Brazilian Arc of Bio and Tech. 2014; 57(1): 62-69.
- 26. Singariya P, Kumar P, Mourya KK. Isolation of Some New Steroids and Evaluation of Bio-activity of *Cenchrus ciliaris*. I J of Res in Pharm Sci. 2012t; 3(4): 678-684.
- Singariya P, Kumar P, Mourya KK. Identification of Steroid Compound using Preparative Thin Layer Chromatography, GC-MS & Anti-microbial and Antioxidant Properties of Cenchrus setigerus (Poaceae). I J of Pharmacy and Life Sci. 2012q; 3(8): 1909-1916.
- 28. Singariya P, Kumar P, Mourya KK. Identification of New Bioactive Compounds by GC-MS and Estimation of Physiological and Biological Activity of Kala Dhaman (Cenchrus setigerus). I J of Pharm and Bio Arc. 2012r; 3(3): 610-616
- 29. Singariya P, Kumar P, Mourya KK. Antimicrobial activity and identification of 4,22-stigmastadiene -3-one and some other compounds in Motha Dhaman grass from Tribal area of Western Rajasthan. Pro of the National Academy of Sci, India Section B: Bio Sci. 2013a; 83(3): 415–421. DOI 10.1007/s40011-012-0135-9.
- Goswami B, Singariya P, Mourya KK, Gadi BR. Abiotic Stress Induced Proteins in Plants. Plant Stress Physiology, Ed. PC. Trivedi. Pointer Publishers, Jaipur. ISBN 13: 9788171328321. 2016b; p. 173-193.
- Singariya P. Effect of Sub-Optimal Environment and PGR's on Metabolic Pattern of Certain Species of *Cenchrus*. Ph. D. Thesis, J. N. Vyas University, Jodhpur (Rajasthan) India. 2009.
- Shinozaki K, Yamaguchi-Shinozaki K. Gene expression and signal transduction in water Stress response. Plant Physiol. 1997; 115(2): 327-334.

Cite this article as:

Premlata Singariya *et al.* Estimation of bio-active compounds in Indian deserted plant *Euphorbia caducifolia* (Danda thor) by gas chromatography-mass spectrometry. Int. Res. J. Pharm. 2018;9(6):87-93 http://dx.doi.org/10.7897/2230-8407.09695

Source of support: Nil, Conflict of interest: None Declared

Disclaimer: IRJP is solely owned by Moksha Publishing House - A non-profit publishing house, dedicated to publish quality research, while every effort has been taken to verify the accuracy of the content published in our Journal. IRJP cannot accept any responsibility or liability for the site content and articles published. The views expressed in articles by our contributing authors are not necessarily those of IRJP editor or editorial board members.